

# Package ‘zinbwave’

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**Type** Package

**Title** Zero-Inflated Negative Binomial Model for RNA-Seq Data

**Version** 1.28.0

**Description** Implements a general and flexible zero-inflated negative binomial model that can be used to provide a low-dimensional representations of single-cell RNA-seq data. The model accounts for zero inflation (dropouts), over-dispersion, and the count nature of the data. The model also accounts for the difference in library sizes and optionally for batch effects and/or other covariates, avoiding the need for pre-normalize the data.

**License** Artistic-2.0

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SingleCellExperiment

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---

`computeDevianceResiduals`  
*Deviance residuals of the zero-inflated negative binomial model*

---

## Description

Given a matrix of counts, this function computes the deviance residuals under a zero-inflated negative binomial (ZINB) model.

## Usage

```
computeDevianceResiduals(model, x, ignoreW = TRUE)
```

## Arguments

<code>model</code>	the zinb model
<code>x</code>	the matrix of counts n cells by J genes
<code>ignoreW</code>	logical, if true matrix W is ignored. Default is TRUE.

## Value

the matrix of deviance residuals of the model.

## Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
computeDevianceResiduals(m, t(assay(se)))
```

---

```
computeObservationalWeights
```

*Observational weights of the zero-inflated negative binomial model for each entry in the matrix of counts*

---

### Description

Given a matrix of counts, this function computes the observational weights of the counts under a zero-inflated negative binomial (ZINB) model. For each count, the ZINB distribution is parametrized by three parameters: the mean value and the dispersion of the negative binomial distribution, and the probability of the zero component.

### Usage

```
computeObservationalWeights(model, x)
```

### Arguments

model	the zinb model
x	the matrix of counts

### Value

the matrix of observational weights computed from the model.

### Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=1), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
computeObservationalWeights(m, assay(se))
```

---

```
getAlpha_mu
```

*Returns the matrix of parameters alpha\_mu*

---

### Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with W for the mean part (mu)

### Usage

```
getAlpha_mu(object, ...)
```

### Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the matrix of alpha\_mu parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getAlpha_mu(a)
```

---

getAlpha_pi	<i>Returns the matrix of parameters alpha_pi</i>
-------------	--

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with W for the zero part (pi)

**Usage**

```
getAlpha_pi(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the matrix of alpha\_pi parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getAlpha_pi(a)
```

---

getBeta_mu	<i>Returns the matrix of parameters beta_mu</i>
------------	---

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with X\_mu

**Usage**

```
getBeta_mu(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the matrix of beta\_mu parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getBeta_mu(a)
```

---

getBeta\_pi

*Returns the matrix of parameters beta\_pi*

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with  $X_{pi}$

**Usage**

```
getBeta_pi(object, ...)
```

**Arguments**

`object` an object that describes a matrix of zero-inflated distributions.  
`...` Additional parameters.

**Value**

the matrix of beta\_pi parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getBeta_pi(a)
```

---

getEpsilon\_alpha

*Returns the vector of regularization parameter for alpha*

---

**Description**

Given an object describing a ZINB model, returns a vector of size the number of rows in the parameter alpha with the regularization parameters associated to each row. Here alpha refers to both alpha\_mu and alpha\_pi, which have the same size and have the same regularization.

**Usage**

```
getEpsilon_alpha(object)
```

**Arguments**

`object` an object that describes a matrix of zero-inflated distributions.

**Value**

the regularization parameters for alpha\_mu and alpha\_pi.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_alpha(a)
```

---

getEpsilon\_beta\_mu      *Returns the vector of regularization parameter for beta\_mu*

---

**Description**

Given an object describing a ZINB model, returns a vector of size the number of rows in the parameter beta\_mu with the regularization parameters associated to each row.

**Usage**

```
getEpsilon_beta_mu(object)
```

**Arguments**

object                  an object that describes a matrix of zero-inflated distributions.

**Value**

the regularization parameters for beta\_mu.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_beta_mu(a)
```

---

getEpsilon\_beta\_pi      *Returns the vector of regularization parameter for beta\_pi*

---

**Description**

Given an object describing a ZINB model, returns a vector of size the number of rows in the parameter beta\_pi with the regularization parameters associated to each row.

**Usage**

```
getEpsilon_beta_pi(object)
```

**Arguments**

object                  an object that describes a matrix of zero-inflated distributions.

**Value**

the regularization parameters for beta\_pi.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_beta_pi(a)
```

---

getEpsilon\_gamma\_mu     *Returns the vector of regularization parameter for gamma\_mu*

---

**Description**

Given an object describing a ZINB model, returns a vector of size the number of columns in the parameter gamma\_mu with the regularization parameters associated to each row.

**Usage**

```
getEpsilon_gamma_mu(object)
```

**Arguments**

object                    an object that describes a matrix of zero-inflated distributions.

**Value**

the regularization parameters for gamma\_mu.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_gamma_mu(a)
```

---

getEpsilon\_gamma\_pi     *Returns the vector of regularization parameter for gamma\_pi*

---

**Description**

Given an object describing a ZINB model, returns a vector of size the number of columns in the parameter gamma\_pi with the regularization parameters associated to each column.

**Usage**

```
getEpsilon_gamma_pi(object)
```

**Arguments**

object                    an object that describes a matrix of zero-inflated distributions.



**Value**

the regularization parameters for gamma\_pi.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_gamma_pi(a)
```

---

getEpsilon_W	<i>Returns the vector of regularization parameter for W</i>
--------------	---

---

**Description**

Given an object describing a ZINB model, returns a vector of size the number of columns in the parameter W with the regularization parameters associated to each column.

**Usage**

```
getEpsilon_W(object)
```

**Arguments**

object            an object that describes a matrix of zero-inflated distributions.

**Value**

the regularization parameters for W.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_W(a)
```

---

getEpsilon_zeta	<i>Returns the regularization parameter for the dispersion parameter</i>
-----------------	--

---

**Description**

The regularization parameter penalizes the variance of zeta, the log of the dispersion parameters across samples.

**Usage**

```
getEpsilon_zeta(object)
```

**Arguments**

object            an object that describes a matrix of zero-inflated distributions.

**Value**

the regularization parameters for zeta.

**Examples**

```
a <- zinbModel(n=5, J=10)
getEpsilon_zeta(a)
```

---

getGamma_mu	<i>Returns the matrix of parameters gamma_mu</i>
-------------	--

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with  $V_{\mu}$

**Usage**

```
getGamma_mu(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the matrix of gamma\_mu parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getGamma_mu(a)
```

---

getGamma_pi	<i>Returns the matrix of parameters gamma_pi</i>
-------------	--

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with  $V_{\pi}$

**Usage**

```
getGamma_pi(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the matrix of gamma\_pi parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getGamma_pi(a)
```

---

getLogitPi

*Returns the matrix of logit of probabilities of zero*

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of logit of probabilities of 0.

**Usage**

```
getLogitPi(object)
```

**Arguments**

`object` an object that describes a matrix of zero-inflated distributions.

**Details**

Note that although the user interface of `zinbFit` requires a  $J \times n$  matrix, internally this is stored as a  $n \times J$  matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of  $n \times J$  dimensions.

**Value**

the matrix of logit-probabilities of 0

**Examples**

```
a <- zinbModel(n=5, J=10)
getLogitPi(a)
```

---

getLogMu	<i>Returns the matrix of logarithm of mean parameters</i>
----------	---

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of logarithm of mean parameters.

**Usage**

```
getLogMu(object)
```

**Arguments**

object            an object that describes a matrix of zero-inflated distributions.

**Details**

Note that although the user interface of `zinbFit` requires a  $J \times n$  matrix, internally this is stored as a  $n \times J$  matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of  $n \times J$  dimensions.

**Value**

the matrix of logarithms of mean parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getLogMu(a)
```

---

getMu	<i>Returns the matrix of mean parameters</i>
-------	--

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of mean parameters.

**Usage**

```
getMu(object)
```

**Arguments**

object            an object that describes a matrix of zero-inflated distributions.

**Details**

Note that although the user interface of `zinbFit` requires a  $J \times n$  matrix, internally this is stored as a  $n \times J$  matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of  $n \times J$  dimensions.

**Value**

the matrix of mean parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getMu(a)
```

---

getPhi

*Returns the vector of dispersion parameters*

---

**Description**

Given an object that describes a matrix of zero-inflated negative binomial distributions, returns the vector of dispersion parameters phi.

**Usage**

```
getPhi(object)
```

**Arguments**

object            an object that describes a matrix of zero-inflated. distributions.

**Value**

the vector of dispersion parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getPhi(a)
```

---

getPi

*Returns the matrix of probabilities of zero*

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of probabilities of 0.

**Usage**

```
getPi(object)
```

**Arguments**

object            an object that describes a matrix of zero-inflated distributions.

## Details

Note that although the user interface of `zinbFit` requires a  $J \times n$  matrix, internally this is stored as a  $n \times J$  matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of  $n \times J$  dimensions.

## Value

the matrix of probabilities of 0

## Examples

```
a <- zinbModel(n=5, J=10)
getPi(a)
```

---

getTheta

*Returns the vector of inverse dispersion parameters*

---

## Description

Given an object that describes a matrix of zero-inflated negative binomial distributions, returns the vector of inverse dispersion parameters `theta`.

## Usage

```
getTheta(object)
```

## Arguments

`object` an object that describes a matrix of zero-inflated distributions.

## Value

the vector of inverse dispersion parameters `theta`

## Examples

```
a <- zinbModel(n=5, J=10)
getTheta(a)
```

---

getV_mu	Returns the gene-level design matrix for mu
---------	---

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the gene-level design matrix for mu

**Usage**

```
getV_mu(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the gene-level design matrix for mu

**Examples**

```
a <- zinbModel(n=5, J=10)
getV_mu(a)
```

---

getV_pi	Returns the gene-level design matrix for pi
---------	---

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the gene-level design matrix for pi

**Usage**

```
getV_pi(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the gene-level design matrix for pi

**Examples**

```
a <- zinbModel(n=5, J=10)
getV_pi(a)
```

---

getW	<i>Returns the low-dimensional matrix of inferred sample-level covariates W</i>
------	---

---

**Description**

Given an object that contains the fit of a ZINB-WaVE model, returns the matrix W of low-dimensional matrix of inferred sample-level covariates.

**Usage**

```
getW(object)
```

**Arguments**

object            a `ZinbModel` object, typically the result of `zinbFit`.

**Value**

the matrix W of inferred sample-level covariates.

**Examples**

```
a <- zinbModel(n=5, J=10)
getW(a)
```

---

getX_mu	<i>Returns the sample-level design matrix for mu</i>
---------	--

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the sample-level design matrix for mu

**Usage**

```
getX_mu(object, ...)
```

**Arguments**

object            an object that describes a matrix of zero-inflated distributions.  
 ...                Additional parameters.

**Value**

the sample-level design matrix for mu

**Examples**

```
a <- zinbModel(n=5, J=10)
getX_mu(a)
```



---

getX_pi	<i>Returns the sample-level design matrix for pi</i>
---------	--

---

**Description**

Given an object that describes a matrix of zero-inflated distributions, returns the sample-level design matrix for pi

**Usage**

```
getX_pi(object, ...)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

**Value**

the sample-level design matrix for pi

**Examples**

```
a <- zinbModel(n=5, J=10)
getX_pi(a)
```

---

getZeta	<i>Returns the vector of log of inverse dispersion parameters</i>
---------	---

---

**Description**

Given an object that describes a matrix of zero-inflated negative binomial distributions, returns the vector zeta of log of inverse dispersion parameters

**Usage**

```
getZeta(object)
```

**Arguments**

object	an object that describes a matrix of zero-inflated distributions.
--------	---

**Value**

the vector zeta of log of inverse dispersion parameters

**Examples**

```
a <- zinbModel(n=5, J=10)
getZeta(a)
```

---

glmWeightedF	<i>Zero-inflation adjusted statistical tests for assessing differential expression.</i>
--------------	---

---

### Description

This function recycles an old version of the `glmLRT` method that allows an F-test with adjusted denominator degrees of freedom to account for the downweighting in the zero-inflation model.

### Usage

```
glmWeightedF(
  glmfit,
  coef = ncol(glmfit$design),
  contrast = NULL,
  ZI = TRUE,
  independentFiltering = TRUE,
  filter = NULL
)
```

### Arguments

<code>glmfit</code>	a <code>DGEGLM-class</code> object, usually output from <code>glmFit</code> .
<code>coef</code>	integer or character vector indicating which coefficients of the linear model are to be tested equal to zero. Values must be columns or column names of design. Defaults to the last coefficient. Ignored if contrast is specified.
<code>contrast</code>	numeric vector or matrix specifying one or more contrasts of the linear model coefficients to be tested equal to zero. Number of rows must equal to the number of columns of design. If specified, then takes precedence over <code>coef</code> .
<code>ZI</code>	logical, specifying whether the degrees of freedom in the statistical test should be adjusted according to the weights in the <code>fit</code> object to account for the downweighting. Defaults to <code>TRUE</code> and this option is highly recommended.
<code>independentFiltering</code>	logical, specifying whether independent filtering should be performed.
<code>filter</code>	vector of values to perform filtering on. Default is the mean of the fitted values from <code>glmfit</code> .

### Details

When ‘`independentFiltering=TRUE`’ (default) an independent filtering step is applied prior to the multiple testing procedure, as described in great details in the ‘DESeq2’ vignette. The values in the ‘`padjFilter`’ column refer to this procedure. They are identical to the ‘`FDR`’ values if the filtering step does not remove any gene, since the function uses the Benjamini-Hochberg correction by default. If the procedure filters some genes, the adjusted p-values will typically result in greater power to detect DE genes. The theory behind independent filtering is described in Bourgon et al. (2010).

### Note

This function uses an adapted version of the `glmLRT` function that was originally written by Gordon Smyth, Davis McCarthy and Yunshun Chen as part of the `edgeR` package. Koen Van den Berge wrote code to adjust residual degree of freedom and added the independent filtering step.

## References

McCarthy, DJ, Chen, Y, Smyth, GK (2012). Differential expression analysis of multifactor RNA-Seq experiments with respect to biological variation. *Nucleic Acids Research* 40, 4288-4297. Bourgon, Richard, Robert Gentleman, and Wolfgang Huber (2010) Independent Filtering Increases Detection Power for High-Throughput Experiments. *PNAS* 107 (21): 9546-51.

## See Also

[glmLRT](#)

---

imputeZeros

*Impute the zeros using the estimated parameters from the ZINB model.*

---

## Description

Given a matrix of counts and a zinb model, this function computes the imputed counts under a zero-inflated negative binomial (ZINB) model.

## Usage

```
imputeZeros(model, x)
```

## Arguments

model	the zinb model
x	the matrix of counts n cells by J genes

## Value

the matrix of imputed counts.

## Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
imputeZeros(m, t(assay(se)))
```

independentFiltering *Perform independent filtering in differential expression analysis.*

---

### Description

This function uses the DESeq2 independent filtering method to increase detection power in high throughput gene expression studies.

### Usage

```
independentFiltering(object, filter, objectType = c("edgeR", "limma"))
```

### Arguments

object	Either a <code>DGELRT-class</code> object or a <code>data.frame</code> with differential expression results.
filter	The characteristic to use for filtering, usually a measure of normalized mean expression for the features.
objectType	Either "edgeR" or "limma". If "edgeR", it is assumed that object is of class <code>DGELRT-class</code> , the output of <code>glmLRT</code> . If "limma", it is assumed that object is a <code>data.frame</code> and the output of a limma-voom analysis.

### Author(s)

Koen Van den Berge

### References

Michael I Love, Wolfgang Huber, and Simon Anders. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology*, 15(12):550, dec 2014.

### See Also

[results](#)

---

loglik *Compute the log-likelihood of a model given some data*

---

### Description

Given a statistical model and some data, this function computes the log-likelihood of the model given the data, i.e., the log-probability of the data under the model.

### Usage

```
loglik(model, x, ...)  
  
## S4 method for signature 'ZinbModel,matrix'  
loglik(model, x)
```

**Arguments**

model            an object that describes a statistical model.  
x                an object that describes data.  
...              additional arguments.

**Value**

The log-likelihood of the model given the data.

**Methods (by class)**

- `loglik(model = ZinbModel, x = matrix)`: return the log-likelihood of the ZINB model.

**Examples**

```
m <- zinbModel(n=5, J=10)
x <- zinbSim(m)
loglik(m, x$counts)
```

---

nFactors

*Generic function that returns the number of latent factors*

---

**Description**

Given an object that describes a dataset or a model involving latent factors, this function returns the number of latent factors.

**Usage**

```
nFactors(x)
```

**Arguments**

x                an object that describes a dataset or a model involving latent factors

**Value**

the number of latent factors

**Examples**

```
a <- zinbModel()
nFactors(a)
```

---

nFeatures	<i>Generic function that returns the number of features</i>
-----------	---

---

**Description**

Given an object that describes a dataset or a model, it returns the number of features.

**Usage**

```
nFeatures(x)
```

**Arguments**

x                    an object that describes a dataset or a model.

**Value**

the number of features.

**Examples**

```
a <- zinbModel()
nFeatures(a)
```

---

nParams	<i>Generic function that returns the total number of parameters of the model</i>
---------	--

---

**Description**

Given an object that describes a model or a dataset, it returns total number of parameters of the model.

**Usage**

```
nParams(model)

## S4 method for signature 'ZinbModel'
nParams(model)
```

**Arguments**

model                an object that describes a dataset or a model.

**Value**

the total number of parameters of the model.

**Functions**

- nParams(ZinbModel): returns the total number of parameters in the model.

**Examples**

```
a <- zinbModel()
nParams(a)
```

---

nSamples

*Generic function that returns the number of samples*


---

**Description**

Given an object that describes a model or a dataset, it returns the number of samples.

**Usage**

```
nSamples(x)
```

**Arguments**

x                    an object that describes a dataset or a model.

**Value**

the number of samples.

**Examples**

```
a <- zinbModel()
nSamples(a)
```

---

orthogonalizeTraceNorm

*Orthogonalize matrices to minimize trace norm of their product*


---

**Description**

Given two matrices U and V that can be multiplied, this function finds two new matrices U2 and V2 such that their product is conserved ( $U*V = U2*V2$ ) and such that  $a||U||^2 + b||V||^2$  is minimized.

**Usage**

```
orthogonalizeTraceNorm(U, V, a = 1, b = 1)
```

**Arguments**

U                    left matrix  
V                    right matrix  
a                    weight of the norm of U (default=1)  
b                    weight of the norm of V (default=1)

**Value**

A list with the two matrices that solve the problem in the slots U and V.

**Examples**

```
U <- matrix(rnorm(15),5,3)
V <- matrix(rnorm(12),3,4)
o <- orthogonalizeTraceNorm(U,V)
norm( U%*%V - o$U%*%o$V) # should be zero
sum(U^2)+sum(V^2)
sum(o$U^2)+sum(o$V^2) # should be smaller
```

---

penalty

*Compute the penalty of a model*

---

**Description**

Given a statistical model with regularization parameters, compute the penalty.

**Usage**

```
penalty(model)

## S4 method for signature 'ZinbModel'
penalty(model)
```

**Arguments**

model            an object that describes a statistical model with regularization parameters.

**Value**

The penalty of the model.

**Methods (by class)**

- `penalty(ZinbModel)`: return the penalization.

**Examples**

```
m <- zinbModel(K=2)
penalty(m)
```



---

pvalueAdjustment	<i>Perform independent filtering in differential expression analysis.</i>
------------------	---

---

### Description

This function performs independent filtering to increase detection power in high throughput gene expression studies.

### Usage

```
pvalueAdjustment(  
  baseMean,  
  filter,  
  pValue,  
  theta,  
  alpha = 0.05,  
  pAdjustMethod = "BH"  
)
```

### Arguments

baseMean	A vector of mean values.
filter	A vector of stage-one filter statistics.
pValue	A vector of unadjusted p-values, or a function which is able to compute this vector from the filtered portion of data, if data is supplied. The option to supply a function is useful when the value of the test statistic depends on which hypotheses are filtered out at stage one. (The limma t-statistic is an example.)
theta	A vector with one or more filtering fractions to consider. Actual cutoffs are then computed internally by applying quantile to the filter statistics contained in (or produced by) the filter argument.
alpha	A cutoff to which p-values, possibly adjusted for multiple testing, will be compared. Default is 0.05.
pAdjustMethod	The unadjusted p-values contained in (or produced by) test will be adjusted for multiple testing after filtering. Default is "BH".

### Value

a list with pvalues, filtering threshold, theta, number of rejections, and alpha.

### Note

This function is an adapted version of the pvalueAdjustment function that was originally written by Michael I. Love as part of the DESeq2 package. Koen Van den Berge adapted the function.

---

`solveRidgeRegression` *Solve ridge regression or logistic regression problems*

---

### Description

This function solves a regression or logistic regression problem regularized by a L2 or weighted L2 penalty. Contrary to `lm.ridge` or `glmnet`, it works for any number of predictors.

### Usage

```
solveRidgeRegression(
  x,
  y,
  beta = rep(0, NCOL(x)),
  epsilon = 1e-06,
  family = c("gaussian", "binomial"),
  offset = rep(0, NROW(x))
)
```

### Arguments

<code>x</code>	a matrix of covariates, one sample per row, one covariate per column.
<code>y</code>	a vector of response (continuous for regression, 0/1 binary for logistic regression)
<code>beta</code>	an initial solution where optimization starts (null vector by default)
<code>epsilon</code>	a scalar or vector of regularization parameters (default 1e-6)
<code>family</code>	a string to choose the type of regression (default family="gaussian")
<code>offset</code>	a vector of offsets (default null vector)

### Details

When family="gaussian", we solve the ridge regression problem that finds the  $\beta$  that minimizes:

$$\|y - x\beta\|^2 + \epsilon\|\beta\|^2/2.$$

When family="binomial" we solve the ridge logistic regression problem

$$\min \sum_i [-y_i(x\beta)_i + \log(1 + \exp(x\beta)_i)] + \epsilon\|\beta\|^2/2.$$

When epsilon is a vector of size equal to the size of beta, then the penalty is a weighted L2 norm  $\sum_i \epsilon_i \beta_i^2 / 2$ .

### Value

A vector solution of the regression problem

---

toydata	<i>Toy dataset to check the model</i>
---------	---------------------------------------

---

**Description**

Toy dataset to check the model

**Format**

A matrix of integers (counts) with 96 samples (rows) and 500 genes (columns).

---

zinb.loglik	<i>Log-likelihood of the zero-inflated negative binomial model</i>
-------------	--

---

**Description**

Given a vector of counts, this function computes the sum of the log-probabilities of the counts under a zero-inflated negative binomial (ZINB) model. For each count, the ZINB distribution is parametrized by three parameters: the mean value and the dispersion of the negative binomial distribution, and the probability of the zero component.

**Usage**

```
zinb.loglik(Y, mu, theta, logitPi)
```

**Arguments**

Y	the vector of counts
mu	the vector of mean parameters of the negative binomial
theta	the vector of dispersion parameters of the negative binomial, or a single scalar is also possible if the dispersion parameter is constant. Note that theta is sometimes called inverse dispersion parameter (and $\phi=1/\theta$ is then called the dispersion parameter). We follow the convention that the variance of the NB variable with mean $\mu$ and dispersion $\theta$ is $\mu + \mu^2/\theta$ .
logitPi	the vector of logit of the probabilities of the zero component

**Value**

the log-likelihood of the model.

**Examples**

```
n <- 10
mu <- seq(10,50,length.out=n)
logitPi <- rnorm(10)
zeta <- rnorm(10)
Y <- rnbino(n=n, size=exp(zeta), mu=mu)
zinb.loglik(Y, mu, exp(zeta), logitPi)
zinb.loglik(Y, mu, 1, logitPi)
```

---

`zinb.loglik.dispersion`

*Log-likelihood of the zero-inflated negative binomial model, for a fixed dispersion parameter*

---

### Description

Given a unique dispersion parameter and a set of counts, together with a corresponding set of mean parameters and probabilities of zero components, this function computes the sum of the log-probabilities of the counts under the ZINB model. The dispersion parameter is provided to the function through `zeta = log(theta)`, where `theta` is sometimes called the inverse dispersion parameter. The probabilities of the zero components are provided through their logit, in order to better numerical stability.

### Usage

```
zinb.loglik.dispersion(zeta, Y, mu, logitPi)
```

### Arguments

<code>zeta</code>	a scalar, the log of the inverse dispersion parameters of the negative binomial model
<code>Y</code>	a vector of counts
<code>mu</code>	a vector of mean parameters of the negative binomial
<code>logitPi</code>	a vector of logit of the probabilities of the zero component

### Value

the log-likelihood of the model.

### See Also

[zinb.loglik.](#)

### Examples

```
mu <- seq(10,50,length.out=10)
logitPi <- rnorm(10)
zeta <- rnorm(10)
Y <- rnbinom(n=10, size=exp(zeta), mu=mu)
zinb.loglik.dispersion(zeta, Y, mu, logitPi)
```

---

`zinb.loglik.dispersion.gradient`*Derivative of the log-likelihood of the zero-inflated negative binomial model with respect to the log of the inverse dispersion parameter*

---

**Description**

Derivative of the log-likelihood of the zero-inflated negative binomial model with respect to the log of the inverse dispersion parameter

**Usage**

```
zinb.loglik.dispersion.gradient(zeta, Y, mu, logitPi)
```

**Arguments**

<code>zeta</code>	the log of the inverse dispersion parameters of the negative binomial
<code>Y</code>	a vector of counts
<code>mu</code>	a vector of mean parameters of the negative binomial
<code>logitPi</code>	a vector of the logit of the probability of the zero component

**Value**

the gradient of the inverse dispersion parameters.

**See Also**

[zinb.loglik](#), [zinb.loglik.dispersion](#).

---

`zinb.loglik.matrix`*Log-likelihood of the zero-inflated negative binomial model for each entry in the matrix of counts*

---

**Description**

Given a matrix of counts, this function computes the log-probabilities of the counts under a zero-inflated negative binomial (ZINB) model. For each count, the ZINB distribution is parametrized by three parameters: the mean value and the dispersion of the negative binomial distribution, and the probability of the zero component.

**Usage**

```
zinb.loglik.matrix(model, x)
```

**Arguments**

<code>model</code>	the zinb model
<code>x</code>	the matrix of counts

**Value**

the matrix of log-likelihood of the model.

---

```
zinb.loglik.regression
```

*Penalized log-likelihood of the ZINB regression model*

---

**Description**

This function computes the penalized log-likelihood of a ZINB regression model given a vector of counts.

**Usage**

```
zinb.loglik.regression(
  alpha,
  Y,
  A.mu = matrix(nrow = length(Y), ncol = 0),
  B.mu = matrix(nrow = length(Y), ncol = 0),
  C.mu = matrix(0, nrow = length(Y), ncol = 1),
  A.pi = matrix(nrow = length(Y), ncol = 0),
  B.pi = matrix(nrow = length(Y), ncol = 0),
  C.pi = matrix(0, nrow = length(Y), ncol = 1),
  C.theta = matrix(0, nrow = length(Y), ncol = 1),
  epsilon = 0
)
```

**Arguments**

alpha	the vectors of parameters c(a.mu, a.pi, b) concatenated
Y	the vector of counts
A.mu	matrix of the model (see Details, default=empty)
B.mu	matrix of the model (see Details, default=empty)
C.mu	matrix of the model (see Details, default=zero)
A.pi	matrix of the model (see Details, default=empty)
B.pi	matrix of the model (see Details, default=empty)
C.pi	matrix of the model (see Details, default=zero)
C.theta	matrix of the model (see Details, default=zero)
epsilon	regularization parameter. A vector of the same length as alpha if each coordinate of alpha has a specific regularization, or just a scalar is the regularization is the same for all coordinates of alpha. Default=0.

**Details**

The regression model is parametrized as follows:

$$\log(\mu) = A_\mu * a_\mu + B_\mu * b + C_\mu$$

$$\text{logit}(\Pi) = A_\pi * a_\pi + B_\pi * b$$

$$\log(\theta) = C_\theta$$

where  $\mu, \Pi, \theta$  are respectively the vector of mean parameters of the NB distribution, the vector of probabilities of the zero component, and the vector of inverse dispersion parameters. Note that the  $b$  vector is shared between the mean of the negative binomial and the probability of zero. The log-likelihood of a vector of parameters  $\alpha = (a_\mu; a_\pi; b)$  is penalized by a regularization term  $\epsilon \|\alpha\|^2/2$  is  $\epsilon$  is a scalar, or  $\sum_i \epsilon_i \alpha_i^2/2$  is  $\epsilon$  is a vector of the same size as  $\alpha$  to allow for differential regularization among the parameters.

**Value**

the penalized log-likelihood.

---

zinb.loglik.regression.gradient

*Gradient of the penalized log-likelihood of the ZINB regression model*

---

**Description**

This function computes the gradient of the penalized log-likelihood of a ZINB regression model given a vector of counts.

**Usage**

```
zinb.loglik.regression.gradient(
  alpha,
  Y,
  A.mu = matrix(nrow = length(Y), ncol = 0),
  B.mu = matrix(nrow = length(Y), ncol = 0),
  C.mu = matrix(0, nrow = length(Y), ncol = 1),
  A.pi = matrix(nrow = length(Y), ncol = 0),
  B.pi = matrix(nrow = length(Y), ncol = 0),
  C.pi = matrix(0, nrow = length(Y), ncol = 1),
  C.theta = matrix(0, nrow = length(Y), ncol = 1),
  epsilon = 0
)
```

**Arguments**

alpha	the vectors of parameters c(a.mu, a.pi, b) concatenated
Y	the vector of counts
A.mu	matrix of the model (see Details, default=empty)
B.mu	matrix of the model (see Details, default=empty)
C.mu	matrix of the model (see Details, default=zero)

A.pi	matrix of the model (see Details, default=empty)
B.pi	matrix of the model (see Details, default=empty)
C.pi	matrix of the model (see Details, default=zero)
C.theta	matrix of the model (see Details, default=zero)
epsilon	regularization parameter. A vector of the same length as alpha if each coordinate of alpha has a specific regularization, or just a scalar is the regularization is the same for all coordinates of alpha. Default=0.

### Details

The regression model is described in [zinb.loglik.regression](#).

### Value

The gradient of the penalized log-likelihood.

### See Also

[zinb.loglik.regression](#)

---

zinb.regression.parseModel

*Parse ZINB regression model*

---

### Description

Given the parameters of a ZINB regression model, this function parses the model and computes the vector of  $\log(\mu)$ ,  $\text{logit}(\pi)$ , and the dimensions of the different components of the vector of parameters. See [zinb.loglik.regression](#) for details of the ZINB regression model and its parameters.

### Usage

```
zinb.regression.parseModel(alpha, A.mu, B.mu, C.mu, A.pi, B.pi, C.pi)
```

### Arguments

alpha	the vectors of parameters $c(a.mu, a.pi, b)$ concatenated
A.mu	matrix of the model (see above, default=empty)
B.mu	matrix of the model (see above, default=empty)
C.mu	matrix of the model (see above, default=zero)
A.pi	matrix of the model (see above, default=empty)
B.pi	matrix of the model (see above, default=empty)
C.pi	matrix of the model (see above, default=zero)

### Value

A list with slots `logMu`, `logitPi`, `dim.alpha` (a vector of length 3 with the dimension of each of the vectors `a.mu`, `a.pi` and `b` in `alpha`), and `start.alpha` (a vector of length 3 with the starting indices of the 3 vectors in `alpha`)



**See Also**

[zinb.loglik.regression](#)

---

 zinbAIC

---

*Compute the AIC or BIC of a model given some data*


---

**Description**

Given a statistical model and some data, these functions compute the AIC or BIC of the model given the data, i.e., the AIC/BIC of the data under the model.

**Usage**

```
zinbAIC(model, x)
```

```
zinbBIC(model, x)
```

```
## S4 method for signature 'ZinbModel,matrix'
zinbAIC(model, x)
```

```
## S4 method for signature 'ZinbModel,matrix'
zinbBIC(model, x)
```

**Arguments**

`model` an object that describes a statistical model.  
`x` an object that describes data.

**Value**

the AIC/BIC of the model.

**Functions**

- `zinbAIC(model = ZinbModel, x = matrix)`: returns the AIC of the ZINB model.
- `zinbBIC(model = ZinbModel, x = matrix)`: returns the BIC of the ZINB model.

**Examples**

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
zinbAIC(m, t(assay(se)))
zinbBIC(m, t(assay(se)))
```

---

`zinbFit`*Fit a ZINB regression model*

---

**Description**

Given an object with the data, it fits a ZINB model.

**Usage**

```
zinbFit(Y, ...)  
  
## S4 method for signature 'SummarizedExperiment'  
zinbFit(  
  Y,  
  X,  
  V,  
  K,  
  which_assay,  
  commondispersion = TRUE,  
  zeroinflation = TRUE,  
  verbose = FALSE,  
  nb.repeat.initialize = 2,  
  maxiter.optimize = 25,  
  stop.epsilon.optimize = 1e-04,  
  BPPARAM = BiocParallel::bpparam(),  
  ...  
)  
  
## S4 method for signature 'matrix'  
zinbFit(  
  Y,  
  X,  
  V,  
  K,  
  commondispersion = TRUE,  
  zeroinflation = TRUE,  
  verbose = FALSE,  
  nb.repeat.initialize = 2,  
  maxiter.optimize = 25,  
  stop.epsilon.optimize = 1e-04,  
  BPPARAM = BiocParallel::bpparam(),  
  ...  
)  
  
## S4 method for signature 'dgCMatrix'  
zinbFit(Y, ...)
```

**Arguments**

`Y` The data (genes in rows, samples in columns).  
`...` Additional parameters to describe the model, see [zinbModel](#).

<code>X</code>	The design matrix containing sample-level covariates, one sample per row. If missing, <code>X</code> will contain only an intercept. If <code>Y</code> is a <code>SummarizedExperiment</code> object, <code>X</code> can be a formula using the variables in the <code>colData</code> slot of <code>Y</code> .
<code>V</code>	The design matrix containing gene-level covariates, one gene per row. If missing, <code>V</code> will contain only an intercept. If <code>Y</code> is a <code>SummarizedExperiment</code> object, <code>V</code> can be a formula using the variables in the <code>rowData</code> slot of <code>Y</code> .
<code>K</code>	integer. Number of latent factors.
<code>which_assay</code>	numeric or character. Which assay of <code>Y</code> to use (only if <code>Y</code> is a <code>SummarizedExperiment</code> ).
<code>commondispersion</code>	Whether or not a single dispersion for all features is estimated (default <code>TRUE</code> ).
<code>zeroinflation</code>	Whether or not a ZINB model should be fitted. If <code>FALSE</code> , a negative binomial model is fitted instead.
<code>verbose</code>	Print helpful messages.
<code>nb.repeat.initialize</code>	Number of iterations for the initialization of <code>beta_mu</code> and <code>gamma_mu</code> .
<code>maxiter.optimize</code>	maximum number of iterations for the optimization step (default 25).
<code>stop.epsilon.optimize</code>	stopping criterion in the optimization step, when the relative gain in likelihood is below <code>epsilon</code> (default 0.0001).
<code>BPPARAM</code>	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.

## Details

By default, i.e., if no arguments other than `Y` are passed, the model is fitted with an intercept for the regression across-samples and one intercept for the regression across genes, both for `mu` and for `pi`.

This means that by default the model is fitted with  $X_{\mu} = X_{\pi} = 1_n$  and  $V_{\mu} = V_{\pi} = 1_J$ . If the user explicitly passes the design matrices, this behavior is overwritten, i.e., the user needs to explicitly include the intercept in the design matrices.

If `Y` is a `SummarizedExperiment`, the function uses the assay named "counts", if any, or the first assay.

Currently, if `Y` is a `sparseMatrix`, this calls the `zinbFit` method on `as.matrix(Y)`

## Value

An object of class `ZinbModel` that has been fitted by penalized maximum likelihood on the data.

## Methods (by class)

- `zinbFit(SummarizedExperiment)`: `Y` is a [SummarizedExperiment](#).
- `zinbFit(matrix)`: `Y` is a matrix of counts (genes in rows).
- `zinbFit(dgCMatrix)`: `Y` is a sparse matrix of counts (genes in rows).

## See Also

[model.matrix](#).

**Examples**

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
                             colData = data.frame(bio = gl(2, 3)))

m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
             BPPARAM=BiocParallel::SerialParam())
bio <- gl(2, 3)
m <- zinbFit(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
             X=model.matrix(~bio), BPPARAM=BiocParallel::SerialParam())
```

---

zinbInitialize

*Initialize the parameters of a ZINB regression model*


---

**Description**

The initialization performs quick optimization of the parameters with several simplifying assumptions compared to the true model: non-zero counts are modeled as log-Gaussian, zeros are modeled as dropouts. The dispersion parameter is not modified.

**Usage**

```
zinbInitialize(
  m,
  Y,
  nb.repeat = 2,
  it.max = 100,
  BPPARAM = BiocParallel::bpparam()
)
```

**Arguments**

m	The model of class ZinbModel
Y	The matrix of counts.
nb.repeat	Number of iterations for the estimation of beta_mu and gamma_mu.
it.max	Maximum number of iterations in softImpute.
BPPARAM	object of class bpparamClass that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.

**Value**

An object of class ZinbModel similar to the one given as argument with modified parameters alpha\_mu, alpha\_pi, beta\_mu, beta\_pi, gamma\_mu, gamma\_pi, W.

**Examples**

```
Y <- matrix(rpois(60, lambda=2), 6, 10)
bio <- gl(2, 3)
time <- rnorm(6)
gc <- rnorm(10)
m <- zinbModel(Y, X=model.matrix(~bio + time), V=model.matrix(~gc),
              which_X_pi=1L, which_V_mu=1L, K=1)
m <- zinbInitialize(m, Y, BPPARAM=BiocParallel::SerialParam())
```

---

zinbModel

*Initialize an object of class ZinbModel*


---

### Description

Initialize an object of class ZinbModel

### Usage

```
zinbModel(
  X,
  V,
  O_mu,
  O_pi,
  which_X_mu,
  which_X_pi,
  which_V_mu,
  which_V_pi,
  W,
  beta_mu,
  beta_pi,
  gamma_mu,
  gamma_pi,
  alpha_mu,
  alpha_pi,
  zeta,
  epsilon,
  epsilon_beta_mu,
  epsilon_gamma_mu,
  epsilon_beta_pi,
  epsilon_gamma_pi,
  epsilon_W,
  epsilon_alpha,
  epsilon_zeta,
  epsilon_min_logit,
  n,
  J,
  K
)
```

### Arguments

X	matrix. The design matrix containing sample-level covariates, one sample per row.
V	matrix. The design matrix containing gene-level covariates, one gene per row.
O_mu	matrix. The offset matrix for mu.
O_pi	matrix. The offset matrix for pi.
which_X_mu	integer. Indices of which columns of X to use in the regression of mu.
which_X_pi	integer. Indices of which columns of X to use in the regression of pi.

<code>which_V_mu</code>	integer. Indices of which columns of $V$ to use in the regression of $\mu$ .
<code>which_V_pi</code>	integer. Indices of which columns of $V$ to use in the regression of $\pi$ .
<code>W</code>	matrix. The factors of sample-level latent factors.
<code>beta_mu</code>	matrix or NULL. The coefficients of $X$ in the regression of $\mu$ .
<code>beta_pi</code>	matrix or NULL. The coefficients of $X$ in the regression of $\pi$ .
<code>gamma_mu</code>	matrix or NULL. The coefficients of $V$ in the regression of $\mu$ .
<code>gamma_pi</code>	matrix or NULL. The coefficients of $V$ in the regression of $\pi$ .
<code>alpha_mu</code>	matrix or NULL. The coefficients of $W$ in the regression of $\mu$ .
<code>alpha_pi</code>	matrix or NULL. The coefficients of $W$ in the regression of $\pi$ .
<code>zeta</code>	numeric. A vector of log of inverse dispersion parameters.
<code>epsilon</code>	nonnegative scalar. Regularization parameter.
<code>epsilon_beta_mu</code>	nonnegative scalar. Regularization parameter for <code>beta_mu</code> .
<code>epsilon_gamma_mu</code>	nonnegative scalar. Regularization parameter for <code>gamma_mu</code> .
<code>epsilon_beta_pi</code>	nonnegative scalar. Regularization parameter for <code>beta_pi</code> .
<code>epsilon_gamma_pi</code>	nonnegative scalar. Regularization parameter for <code>gamma_pi</code> .
<code>epsilon_W</code>	nonnegative scalar. Regularization parameter for <code>W</code> .
<code>epsilon_alpha</code>	nonnegative scalar. Regularization parameter for <code>alpha</code> (both <code>alpha_mu</code> and <code>alpha_pi</code> ).
<code>epsilon_zeta</code>	nonnegative scalar. Regularization parameter for <code>zeta</code> .
<code>epsilon_min_logit</code>	scalar. Minimum regularization parameter for parameters of the logit model, including the intercept.
<code>n</code>	integer. Number of samples.
<code>J</code>	integer. Number of genes.
<code>K</code>	integer. Number of latent factors.

## Details

This is a wrapper around the `new()` function to create an instance of class `ZinbModel`. Rarely, the user will need to create a `ZinbModel` object from scratch, as typically this is the result of `zinbFit`.

If any of  $X$ ,  $V$ ,  $W$  matrices are passed,  $n$ ,  $J$ , and  $K$  are inferred. Alternatively, the user can specify one or more of  $n$ ,  $J$ , and  $K$ .

The regularization parameters can be set by a unique parameter `epsilon` or specific values for the different regularization parameters can also be provided. If only `epsilon` is specified, the other parameters take the following values:

- `epsilon_beta` = `epsilon/J`
- `epsilon_gamma` = `epsilon/n`
- `epsilon_W` = `epsilon/n`
- `epsilon_alpha` = `epsilon/J`
- `epsilon_zeta` = `epsilon`

We empirically found that large values of epsilon provide a more stable estimation of  $W$ .

A call with no argument has the following default values:  $n = 50$ ,  $J = 100$ ,  $K = 0$ ,  $\text{epsilon} = J$ .

Although it is possible to create new instances of the class by calling this function, this is not the most common way of creating ZinbModel objects. The main use of the class is within the `zinbFit` function.

### Value

an object of class `ZinbModel`.

### Examples

```
a <- zinbModel()
nSamples(a)
nFeatures(a)
nFactors(a)
nParams(a)
```

---

ZinbModel-class	<i>Class ZinbModel</i>
-----------------	------------------------

---

### Description

Objects of this class store all the values needed to work with a zero-inflated negative binomial (ZINB) model, as described in the vignette. They contain all information to fit a model by penalized maximum likelihood or simulate data from a model.

### Usage

```
## S4 method for signature 'ZinbModel'
show(object)

## S4 method for signature 'ZinbModel'
nSamples(x)

## S4 method for signature 'ZinbModel'
nFeatures(x)

## S4 method for signature 'ZinbModel'
nFactors(x)

## S4 method for signature 'ZinbModel'
getX_mu(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
getX_pi(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
getV_mu(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
```

```
getV_pi(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
getLogMu(object)

## S4 method for signature 'ZinbModel'
getMu(object)

## S4 method for signature 'ZinbModel'
getLogitPi(object)

## S4 method for signature 'ZinbModel'
getPi(object)

## S4 method for signature 'ZinbModel'
getZeta(object)

## S4 method for signature 'ZinbModel'
getPhi(object)

## S4 method for signature 'ZinbModel'
getTheta(object)

## S4 method for signature 'ZinbModel'
getEpsilon_beta_mu(object)

## S4 method for signature 'ZinbModel'
getEpsilon_gamma_mu(object)

## S4 method for signature 'ZinbModel'
getEpsilon_beta_pi(object)

## S4 method for signature 'ZinbModel'
getEpsilon_gamma_pi(object)

## S4 method for signature 'ZinbModel'
getEpsilon_W(object)

## S4 method for signature 'ZinbModel'
getEpsilon_alpha(object)

## S4 method for signature 'ZinbModel'
getEpsilon_zeta(object)

## S4 method for signature 'ZinbModel'
getW(object)

## S4 method for signature 'ZinbModel'
getBeta_mu(object)

## S4 method for signature 'ZinbModel'
getBeta_pi(object)
```



```
## S4 method for signature 'ZinbModel'
getGamma_mu(object)

## S4 method for signature 'ZinbModel'
getGamma_pi(object)

## S4 method for signature 'ZinbModel'
getAlpha_mu(object)

## S4 method for signature 'ZinbModel'
getAlpha_pi(object)
```

### Arguments

object	an object of class ZinbModel.
x	an object of class ZinbModel.
intercept	logical. Whether to return the intercept (ignored if the design matrix has no intercept). Default TRUE

### Details

For the full description of the model see the model vignette. Internally, the slots are checked so that the matrices are of the appropriate dimensions: in particular,  $X$ ,  $O_{\mu}$ ,  $O_{\pi}$ , and  $W$  need to have  $n$  rows,  $V$  needs to have  $J$  rows,  $\zeta$  must be of length  $J$ .

### Value

`nSamples` returns the number of samples; `nFeatures` returns the number of features; `nFactors` returns the number of latent factors.

### Methods (by generic)

- `show(ZinbModel)`: show useful info on the object.
- `nSamples(ZinbModel)`: returns the number of samples.
- `nFeatures(ZinbModel)`: returns the number of features.
- `nFactors(ZinbModel)`: returns the number of latent factors.
- `getX_mu(ZinbModel)`: returns the sample-level design matrix for  $\mu$ .
- `getX_pi(ZinbModel)`: returns the sample-level design matrix for  $\pi$ .
- `getV_mu(ZinbModel)`: returns the gene-level design matrix for  $\mu$ .
- `getV_pi(ZinbModel)`: returns the sample-level design matrix for  $\pi$ .
- `getLogMu(ZinbModel)`: returns the logarithm of the mean of the non-zero component.
- `getMu(ZinbModel)`: returns the mean of the non-zero component.
- `getLogitPi(ZinbModel)`: returns the logit-probability of zero.
- `getPi(ZinbModel)`: returns the probability of zero.
- `getZeta(ZinbModel)`: returns the log of the inverse of the dispersion parameter.
- `getPhi(ZinbModel)`: returns the dispersion parameter.
- `getTheta(ZinbModel)`: returns the inverse of the dispersion parameter.

- `getEpsilon_beta_mu(ZinbModel)`: returns the regularization parameters for `beta_mu`.
- `getEpsilon_gamma_mu(ZinbModel)`: returns the regularization parameters for `gamma_mu`.
- `getEpsilon_beta_pi(ZinbModel)`: returns the regularization parameters for `beta_pi`.
- `getEpsilon_gamma_pi(ZinbModel)`: returns the regularization parameters for `gamma_pi`.
- `getEpsilon_W(ZinbModel)`: returns the regularization parameters for `W`.
- `getEpsilon_alpha(ZinbModel)`: returns the regularization parameters for `alpha`.
- `getEpsilon_zeta(ZinbModel)`: returns the regularization parameters for `zeta`.
- `getW(ZinbModel)`: returns the matrix `W` of inferred sample-level covariates.
- `getBeta_mu(ZinbModel)`: returns the matrix `beta_mu` of inferred parameters.
- `getBeta_pi(ZinbModel)`: returns the matrix `beta_pi` of inferred parameters.
- `getGamma_mu(ZinbModel)`: returns the matrix `gamma_mu` of inferred parameters.
- `getGamma_pi(ZinbModel)`: returns the matrix `gamma_pi` of inferred parameters.
- `getAlpha_mu(ZinbModel)`: returns the matrix `alpha_mu` of inferred parameters.
- `getAlpha_pi(ZinbModel)`: returns the matrix `alpha_pi` of inferred parameters.

### Slots

`X` matrix. The design matrix containing sample-level covariates, one sample per row.

`V` matrix. The design matrix containing gene-level covariates, one gene per row.

`O_mu` matrix. The offset matrix for `mu`.

`O_pi` matrix. The offset matrix for `pi`.

`which_X_mu` integer. Indices of which columns of `X` to use in the regression of `mu`.

`which_V_mu` integer. Indices of which columns of `V` to use in the regression of `mu`.

`which_X_pi` integer. Indices of which columns of `X` to use in the regression of `pi`.

`which_V_pi` integer. Indices of which columns of `V` to use in the regression of `pi`.

`X_mu_intercept` logical. TRUE if `X_mu` contains an intercept.

`X_pi_intercept` logical. TRUE if `X_pi` contains an intercept.

`V_mu_intercept` logical. TRUE if `V_mu` contains an intercept.

`V_pi_intercept` logical. TRUE if `V_pi` contains an intercept.

`W` matrix. The factors of sample-level latent factors.

`beta_mu` matrix or NULL. The coefficients of `X` in the regression of `mu`.

`gamma_mu` matrix or NULL. The coefficients of `V` in the regression of `mu`.

`alpha_mu` matrix or NULL. The coefficients of `W` in the regression of `mu`.

`beta_pi` matrix or NULL. The coefficients of `X` in the regression of `pi`.

`gamma_pi` matrix or NULL. The coefficients of `V` in the regression of `pi`.

`alpha_pi` matrix or NULL. The coefficients of `W` in the regression of `pi`.

`zeta` numeric. A vector of log of inverse dispersion parameters.

`epsilon_beta_mu` nonnegative scalar. Regularization parameter for `beta_mu`

`epsilon_gamma_mu` nonnegative scalar. Regularization parameter for `gamma_mu`

`epsilon_beta_pi` nonnegative scalar. Regularization parameter for `beta_pi`

`epsilon_gamma_pi` nonnegative scalar. Regularization parameter for `gamma_pi`

epsilon\_W nonnegative scalar. Regularization parameter for W  
 epsilon\_alpha nonnegative scalar. Regularization parameter for alpha (both alpha\_mu and alpha\_pi)  
 epsilon\_zeta nonnegative scalar. Regularization parameter for zeta  
 epsilon\_min\_logit scalar. Minimum regularization parameter for parameters of the logit model, including the intercept.

---

zinbOptimize *Optimize the parameters of a ZINB regression model*

---

## Description

The parameters of the model given as argument are optimized by penalized maximum likelihood on the count matrix given as argument. It is recommended to call `zinb_initialize` before this function to have good starting point for optimization, since the optimization problem is not convex and can only converge to a local minimum.

## Usage

```

zinbOptimize(
  m,
  Y,
  comondispersion = TRUE,
  maxiter = 25,
  stop.epsilon = 1e-04,
  verbose = FALSE,
  BPPARAM = BiocParallel::bpparam()
)

```

## Arguments

<code>m</code>	The model of class <code>ZinbModel</code>
<code>Y</code>	The matrix of counts.
<code>comondispersion</code>	Whether the dispersion is the same for all features (default=TRUE)
<code>maxiter</code>	maximum number of iterations (default 25)
<code>stop.epsilon</code>	stopping criterion, when the relative gain in likelihood is below epsilon (default 0.0001)
<code>verbose</code>	print information (default FALSE)
<code>BPPARAM</code>	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.

## Value

An object of class `ZinbModel` similar to the one given as argument with modified parameters `alpha_mu`, `alpha_pi`, `beta_mu`, `beta_pi`, `gamma_mu`, `gamma_pi`, `W`.

**Examples**

```

Y = matrix(10, 3, 5)
m = zinbModel(n=NROW(Y), J=NCOL(Y))
m = zinbInitialize(m, Y, BPPARAM=BiocParallel::SerialParam())
m = zinbOptimize(m, Y, BPPARAM=BiocParallel::SerialParam())

```

---

zinbOptimizeDispersion

*Optimize the dispersion parameters of a ZINB regression model*


---

**Description**

The dispersion parameters of the model are optimized by penalized maximum likelihood on the count matrix given as argument.

**Usage**

```

zinbOptimizeDispersion(
  J,
  mu,
  logitPi,
  epsilon,
  Y,
  commondispersion = TRUE,
  BPPARAM = BiocParallel::bpparam()
)

```

**Arguments**

J	The number of genes.
mu	the matrix containing the mean of the negative binomial.
logitPi	the matrix containing the logit of the probability parameter of the zero-inflation part of the model.
epsilon	the regularization parameter.
Y	The matrix of counts.
commondispersion	Whether or not a single dispersion for all features is estimated (default TRUE)
BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.

**Value**

An object of class `ZinbModel` similar to the one given as argument with modified parameters `zeta`.

**Examples**

```

Y = matrix(10, 3, 5)
m = zinbModel(n=NROW(Y), J=NCOL(Y))
m = zinbInitialize(m, Y, BPPARAM=BiocParallel::SerialParam())
m = zinbOptimizeDispersion(NROW(Y), getMu(m), getLogitPi(m),
  getEpsilon_zeta(m), Y, BPPARAM=BiocParallel::SerialParam())

```

---

zinbSim	<i>Simulate counts from a zero-inflated negative binomial model</i>
---------	---

---

### Description

Given an object that describes zero-inflated negative binomial distribution, simulate counts from the distribution.

### Usage

```
zinbSim(object, seed, ...)  
  
## S4 method for signature 'ZinbModel'  
zinbSim(object, seed)
```

### Arguments

object	an object that describes a matrix of zero-inflated negative binomial.
seed	an optional integer to specify how the random number generator should be initialized with a call to <code>set.seed</code> . If missing, the random generator state is not changed.
...	additional arguments.

### Value

A list with the following elements.

- `counts` the matrix with the simulated counts.
- `dataNB` the data simulated from the negative binomial.
- `dataDropout` the data simulated from the binomial process.
- `zeroFraction` the fraction of zeros.

### Methods (by class)

- `zinbSim(ZinbModel)`: simulate from a ZINB distribution.

### Examples

```
a <- zinbModel(n=5, J=10)  
zinbSim(a)
```

---

zinbsurf	<i>Perform dimensionality reduction using a ZINB regression model for large datasets.</i>
----------	---

---

### Description

Given an object with the data, it performs dimensionality reduction using a ZINB regression model with gene and cell-level covariates on a random subset of the data. It then projects the remaining data onto the lower dimensional space.

### Usage

```
zinbsurf(Y, ...)

## S4 method for signature 'SummarizedExperiment'
zinbsurf(
  Y,
  X,
  V,
  K,
  which_assay,
  which_genes,
  zeroinflation = TRUE,
  prop_fit = 0.1,
  BPPARAM = BiocParallel::bpparam(),
  verbose = FALSE,
  ...
)
```

### Arguments

Y	The data (genes in rows, samples in columns). Currently implemented only for SummarizedExperiment.
...	Additional parameters to describe the model, see <a href="#">zinbModel</a> .
X	The design matrix containing sample-level covariates, one sample per row. If missing, X will contain only an intercept. If Y is a SummarizedExperiment object, X can be a formula using the variables in the colData slot of Y.
V	The design matrix containing gene-level covariates, one gene per row. If missing, V will contain only an intercept. If Y is a SummarizedExperiment object, V can be a formula using the variables in the rowData slot of Y.
K	integer. Number of latent factors. Specify $K = 0$ if only computing observational weights.
which_assay	numeric or character. Which assay of Y to use. If missing, if 'assayNames(Y)' contains "counts" then that is used. Otherwise, the first assay is used.
which_genes	character. Which genes to use to estimate W (see details). Ignored if fitted_model is provided.
zeroinflation	Whether or not a ZINB model should be fitted. If FALSE, a negative binomial model is fitted instead.
prop_fit	numeric between 0 and 1. The proportion of cells to use for the zinbwave fit.

BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.
verbose	Print helpful messages.

### Details

This function implements an approximate strategy, in which the full `zinbwave` model is fit only on a random subset of the data (controlled by the `prop_fit` parameter). The rest of the samples are subsequently projected onto the low-rank space. This strategy is much faster and uses less memory than the full `zinbwave` method. It is recommended with extremely large datasets.

By default `zinbwave` uses all genes to estimate  $W$ . However, we recommend to use the top 1,000 most variable genes for this step. In general, a user can specify any custom set of genes to be used to estimate  $W$ , by specifying either a vector of gene names, or a single character string corresponding to a column of the `rowData`.

### Value

An object of class `SingleCellExperiment`; the dimensionality reduced matrix is stored in the `reducedDims` slot.

### Methods (by class)

- `zinbwave(SummarizedExperiment)`:  $Y$  is a [SummarizedExperiment](#).

### Examples

```
se <- SingleCellExperiment(assays = list(counts = matrix(rpois(60, lambda=5),
                                                       nrow=10, ncol=6)),
                          colData = data.frame(bio = gl(2, 3)))
colnames(se) <- paste0("sample", 1:6)
m <- zinbwave(se, X="~bio", K = 1, prop_fit = .5, which_assay = 1,
              BPPARAM=BiocParallel::SerialParam())
```

---

<code>zinbwave</code>	<i>Perform dimensionality reduction using a ZINB regression model with gene and cell-level covariates.</i>
-----------------------	--

---

### Description

Given an object with the data, it performs dimensionality reduction using a ZINB regression model with gene and cell-level covariates.

### Usage

```
zinbwave(Y, ...)

## S4 method for signature 'SummarizedExperiment'
zinbwave(
  Y,
  X,
  V,
  K = 2,
```

```

fitted_model,
which_assay,
which_genes,
commondispersion = TRUE,
zeroinflation = TRUE,
verbose = FALSE,
nb.repeat.initialize = 2,
maxiter.optimize = 25,
stop.epsilon.optimize = 1e-04,
BPPARAM = BiocParallel::bpparam(),
normalizedValues = FALSE,
residuals = FALSE,
imputedValues = FALSE,
observationalWeights = FALSE,
...
)

```

### Arguments

Y	The data (genes in rows, samples in columns). Currently implemented only for SummarizedExperiment.
...	Additional parameters to describe the model, see <a href="#">zinbModel</a> .
X	The design matrix containing sample-level covariates, one sample per row. If missing, X will contain only an intercept. If Y is a SummarizedExperiment object, X can be a formula using the variables in the colData slot of Y.
V	The design matrix containing gene-level covariates, one gene per row. If missing, V will contain only an intercept. If Y is a SummarizedExperiment object, V can be a formula using the variables in the rowData slot of Y.
K	integer. Number of latent factors. Specify $K = 0$ if only computing observational weights.
fitted_model	a <a href="#">ZinbModel</a> object.
which_assay	numeric or character. Which assay of Y to use. If missing, if 'assayNames(Y)' contains "counts" then that is used. Otherwise, the first assay is used.
which_genes	character. Which genes to use to estimate W (see details). Ignored if fitted_model is provided.
commondispersion	Whether or not a single dispersion for all features is estimated (default TRUE).
zeroinflation	Whether or not a ZINB model should be fitted. If FALSE, a negative binomial model is fitted instead.
verbose	Print helpful messages.
nb.repeat.initialize	Number of iterations for the initialization of beta_mu and gamma_mu.
maxiter.optimize	maximum number of iterations for the optimization step (default 25).
stop.epsilon.optimize	stopping criterion in the optimization step, when the relative gain in likelihood is below epsilon (default 0.0001).
BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See <a href="#">bpparam</a> for details.



normalizedValues	indicates whether or not you want to compute normalized values for the counts after adjusting for gene and cell-level covariates.
residuals	indicates whether or not you want to compute the residuals of the ZINB model. Deviance residuals are computed.
imputedValues	indicates whether or not you want to compute the imputed counts of the ZINB model.
observationalWeights	indicates whether to compute the observational weights for differential expression (see vignette).

## Details

For visualization (heatmaps, ...), please use the normalized values. It corresponds to the deviance residuals when the  $W$  is not included in the model but the gene and cell-level covariates are. As a result, when  $W$  is not included in the model, the deviance residuals should capture the biology. Note that we do not recommend to use the normalized values for any downstream analysis (such as clustering, or differential expression), but only for visualization.

If one has already fitted a model using `ZinbModel`, the object containing such model can be used as input of `zinbwave` to save the resulting  $W$  into a `SummarizedExperiment` and optionally compute residuals and normalized values, without the need for re-fitting the model.

By default `zinbwave` uses all genes to estimate  $W$ . However, we recommend to use the top 1,000 most variable genes for this step. In general, a user can specify any custom set of genes to be used to estimate  $W$ , by specifying either a vector of gene names, or a single character string corresponding to a column of the `rowData`.

Note that if both `which_genes` is specified and at least one among `observationalWeights`, `imputedValues`, `residuals`, and `normalizedValues` is `TRUE`, the model needs to be fit twice.

## Value

An object of class `SingleCellExperiment`; the dimensionality reduced matrix is stored in the `reducedDims` slot and optionally normalized values and residuals are added in the list of assays.

## Methods (by class)

- `zinbwave(SummarizedExperiment)`:  $Y$  is a `SummarizedExperiment`.

## Examples

```
se <- SingleCellExperiment(assays = list(counts = matrix(rpois(60, lambda=5),
                                                    nrow=10, ncol=6)),
                          colData = data.frame(bio = gl(2, 3)))

m <- zinbwave(se, X="~bio", BPPARAM=BiocParallel::SerialParam())
```

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