

# Package ‘vissE’

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**Title** Visualising Set Enrichment Analysis Results

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**Description** This package enables the interpretation and analysis of results from a gene set enrichment analysis using network-based and text-mining approaches. Most enrichment analyses result in large lists of significant gene sets that are difficult to interpret. Tools in this package help build a similarity-based network of significant gene sets from a gene set enrichment analysis that can then be investigated for their biological function using text-mining approaches.

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vissE-package	<i>vissE: Visualising Set Enrichment Analysis Results</i>
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## Description

This package enables the interpretation and analysis of results from a gene set enrichment analysis using network-based and text-mining approaches. Most enrichment analyses result in large lists of significant gene sets that are difficult to interpret. Tools in this package help build a similarity-based network of significant gene sets from a gene set enrichment analysis that can then be investigated for their biological function using text-mining approaches.

## Details

This package supports four workflows to enhance gene set enrichment analysis:

1. Clustering results from a gene set enrichment analysis (e.g. using `limma::fry`, `singscore` or `GSEA`). The functions required for this analysis are `computeMsigOverlap`, `computeMsigNetwork` and `plotMsigNetwork`.
2. Interpreting gene set clusters (identified in the first analysis) by performing text-mining of gene set names and descriptions. The main function required to perform text-mining of gene sets is `plotMsigWordcloud`. Other functions can be used to access intermediate results.
3. Visualise gene-level statistics for gene set clusters identified in the first analysis to link back gene set clusters to the genes of interest. This can be done using the `plotGeneStats` function.
4. Identifying gene sets similar to a list of genes identified from a DE analysis using set overlap measures. This can be done using the `characteriseGeneset` function.

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**See Also**

Useful links:

- <https://davislaboratory.github.io/vissE>
- Report bugs at <https://github.com/DavisLaboratory/vissE/issues>

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bhuvad\_theme

*Custom theme*

---

**Description**

Custom theme

**Usage**

```
bhuvad_theme(r1 = 1.1)
```

**Arguments**

**r1** a numeric, scaling factor to apply to text sizes

**Value**

a ggplot2 theme

**Examples**

```
p1 = ggplot2::ggplot()
p1 + bhuvad_theme()
```

---

characteriseGeneset      *Functionally characterise a list of genes*

---

### Description

This function can be used to perform a network-based enrichment analysis of a list of genes. The list of genes are characterised based on their similarity with gene sets from the MSigDB. A network of similar gene sets is retrieved using this function.

### Usage

```
characteriseGeneset(
  gs,
  thresh = 0.2,
  measure = c("overlapcoef", "jaccard"),
  gscolcs = c("h", "c2", "c5"),
  org = c("auto", "hs", "mm")
)
```

### Arguments

<code>gs</code>	a GeneSet object, representing the list of genes that need to be characterised.
<code>thresh</code>	a numeric, specifying the threshold to discard pairs of gene sets.
<code>measure</code>	a character, specifying the similarity measure to use: <code>ari</code> for the Adjusted Rand Index, <code>jaccard</code> for the Jaccard Index and <code>overlapcoef</code> for the Overlap Coefficient.
<code>gscolcs</code>	a character, listing the MSigDB collections to use as a background (defaults to <code>h</code> , <code>c2</code> , and <code>c5</code> ). Collection types can be retrieved using <code>msigdb::listCollections()</code> .
<code>org</code>	a character, specifying the organism to use. This can either be <code>"auto"</code> (default), <code>"hs"</code> or <code>"mm"</code> .

### Value

an igraph object, containing gene sets that are similar to the query set. The network contains relationships between results of the query too.

### Examples

```
library(GSEABase)
data(hgsc)

#create a geneset using one of the Hallmark gene sets
mySet <- GeneSet(
  geneIds(hgsc[[2]]),
  setName = 'MySet',
  geneIdType = SymbolIdentifier()
)
```

```
#characterise the custom gene set
ig <- characteriseGeneset(mySet)
plotMsigNetwork(ig)
```

---

computeMsigNetwork	<i>Compute a network using computed gene set overlap</i>
--------------------	--

---

### Description

Computes an igraph object using information on gene sets and gene sets computed using the [computeMsigOverlap\(\)](#) function.

### Usage

```
computeMsigNetwork(genesetOverlap, msigGsc)
```

### Arguments

genesetOverlap	a data.frame, containing results of an overlap analysis computed using the <a href="#">computeMsigOverlap()</a> function.
msigGsc	a GeneSetCollection object, containing gene sets used to compute overlap.

### Value

an igraph object

### Examples

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc)
ig <- computeMsigNetwork(ovlap, hgsc)
```

---

computeMsigOverlap	<i>Compute gene set overlap</i>
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---

## Description

Compute overlap between gene sets from a `GeneSetCollection` using the Jaccard index or the overlap coefficient. These values can then be used to compute a network of gene set overlaps.

## Usage

```
computeMsigOverlap(  
  msigGsc1,  
  msigGsc2 = NULL,  
  thresh = 0.25,  
  measure = c("ari", "jaccard", "overlapcoef")  
)
```

## Arguments

<code>msigGsc1</code>	a <code>GeneSetCollection</code> object.
<code>msigGsc2</code>	a <code>GeneSetCollection</code> object or <code>NULL</code> if pairwise overlaps are to be computed.
<code>thresh</code>	a numeric, specifying the threshold to discard pairs of gene sets.
<code>measure</code>	a character, specifying the similarity measure to use: <code>ari</code> for the Adjusted Rand Index, <code>jaccard</code> for the Jaccard Index and <code>overlapcoef</code> for the Overlap Coefficient.

## Value

a `data.frame`, containing the overlap structure of gene sets represented as a network in the simple interaction format (SIF).

## Examples

```
data(hgsc)  
ovlap <- computeMsigOverlap(hgsc)
```

---

computeMsigWordFreq	<i>Compute word frequencies for a single MSigDB collection</i>
---------------------	--

---

## Description

Compute word frequencies for a single MSigDB collection

## Usage

```
computeMsigWordFreq(
  msigGsc,
  weight = NULL,
  measure = c("tfidf", "tf"),
  version = msigdb::getMsigdbVersions(),
  org = c("auto", "hs", "mm"),
  rmwords = getMsigExclusionList(),
  idf = NULL
)
```

## Arguments

msigGsc	a GeneSetCollection object, containing gene sets from the MSigDB. The <a href="#">GSEABase::getBroadSets()</a> function can be used to parse XML files downloaded from MSigDB.
weight	a named numeric vector, containing weights to apply to each gene-set. This can be $-\log_{10}(\text{FDR})$ , $-\log_{10}(\text{p-value})$ or an enrichment score (ideally unsigned).
measure	a character, specifying how frequencies should be computed. "tf" uses term frequencies and "tfidf" (default) applies inverse document frequency weights to term frequencies.
version	a character, specifying the version of msigdb to use (see <a href="#">msigdb::getMsigdbVersions()</a> ).
org	a character, specifying the organism to use. This can either be "auto" (default), "hs" or "mm".
rmwords	a character vector, containing an exclusion list of words to discard from the analysis.
idf	a list of named numeric vectors, specifying inverse document frequencies to use to penalise terms from gene-set names and short descriptions. This should be a vector of length 2 with names "Name" and "Short". Numeric vectors should contain weights and names should represent the term. Precomputed versions can be retrieved using the <a href="#">msigdb::getMsigdbIDF()</a> .

## Value

a list, containing two data.frames summarising the results of the frequency analysis on gene set names and short descriptions.

## Examples

```
data(hgsc)
freq <- computeMsigWordFreq(hgsc, measure = 'tfidf')
```

---

findMsigClusters	<i>Identify gene-set clusters from a gene-set overlap network</i>
------------------	---

---

## Description

This function identifies gene-set clusters from a gene-set overlap network produced using `vissE`. Various graph clustering algorithms from the `igraph` package can be used for clustering. Gene-set clusters identified are then sorted based on their size and a given statistic of interest (absolute of the statistic is maximised per cluster).

## Usage

```
findMsigClusters(
  ig,
  genesetStat = NULL,
  minSize = 2,
  alg = igraph::cluster_walktrap,
  alparams = list()
)
```

## Arguments

<code>ig</code>	an <code>igraph</code> object, containing a network of gene set overlaps computed using <code>computeMsigNetwork()</code> .
<code>genesetStat</code>	a named numeric, containing statistics for each gene-set that are to be used in cluster prioritisation. If <code>NULL</code> , clusters are prioritised based on their size (number of gene-sets in them).
<code>minSize</code>	a numeric, stating the minimum size a cluster can be (default is 2).
<code>alg</code>	a function, from the <code>igraph</code> package that should be used to perform graph-clustering (default is <code>igraph::cluster_walktrap</code> ). The function should produce a <code>communities</code> object.
<code>alparams</code>	a list, specifying additional parameters that are to be passed to the graph clustering algorithm.

## Details

Gene-sets clusters are identified using graph clustering and are prioritised based on a combination of cluster size and optionally, a statistic of interest (e.g., enrichment scores). A product-of-ranks approach is used to prioritise clusters when gene-set statistics are available. In this approach, clusters are ranked based on their cluster size (largest to smallest) and on the median absolute statistic



of gene-sets within it (largest to smallest). The product of these ranks is computed and clusters are ranked based on these product-of-rank statistic (smallest to largest).

When prioritising using cluster size and gene-set statistics, if statistics for some gene-sets in the network are missing, only the size is used in cluster prioritisation.

**Value**

a list, containing gene-sets that belong to each cluster. Items in the list are organised based on prioritisation.

**Examples**

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc, thresh = 0.25)
ig <- computeMsigNetwork(ovlap, hgsc)
findMsigClusters(ig)
```

---

getMsigExclusionList	<i>Exclusion list of words for MSigDB gene set text mining</i>
----------------------	--

---

**Description**

List of words to discard when performing text mining MSigDB gene set names and short descriptions.

**Usage**

```
getMsigExclusionList(custom = c())
```

**Arguments**

custom	a character vector, containing list of words to add onto existing exclusion list.
--------	---

**Value**

a character vector, containing words to be excluded from the text mining analysis.

**Examples**

```
getMsigExclusionList('remove')
```

hgsc

*The Hallmark collection from the MSigDB***Description**

The molecular signatures database (MSigDB) is a collection of over 25000 gene expression signatures. Signatures in v7.2 are divided into 9 categories. The Hallmarks collection contains gene expression signatures representing molecular processes that are hallmarks in cancer development and progression.

**Usage**

hgsc

**Format**

A GeneSetCollection object with 50 GeneSet objects representing the 50 Hallmark gene expression signatures.

**References**

Subramanian, A., Tamayo, P., Mootha, V. K., Mukherjee, S., Ebert, B. L., Gillette, M. A., ... & Mesirov, J. P. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proceedings of the National Academy of Sciences*, 102(43), 15545-15550.

Liberzon, A., Subramanian, A., Pinchback, R., Thorvaldsdóttir, H., Tamayo, P., & Mesirov, J. P. (2011). Molecular signatures database (MSigDB) 3.0. *Bioinformatics*, 27(12), 1739-1740.

Liberzon, A., Birger, C., Thorvaldsdóttir, H., Ghandi, M., Mesirov, J. P., & Tamayo, P. (2015). The molecular signatures database hallmark gene set collection. *Cell systems*, 1(6), 417-425.

plotGeneStats

*Plot gene statistics for clusters of gene sets***Description**

This function plots gene statistics against gene frequencies for any given cluster of gene sets. The plot can be used to identify genes that are over-represented in a cluster of gene-sets (identified based on gene-set overlaps) and have a strong statistic (e.g. log fold-change or p-value).

**Usage**

```
plotGeneStats(  
  geneStat,  
  msigGsc,  
  groups,  
  statName = "Gene-level statistic",  
  topN = 5  
)
```

**Arguments**

geneStat	a named numeric, containing the statistic to be displayed. The vector must be named with either gene Symbols or Entrez IDs depending on annotations in msigGsc.
msigGsc	a GeneSetCollection object, containing gene sets from the MSigDB. The <a href="#">GSEABase::getBroadSets()</a> function can be used to parse XML files downloaded from MSigDB.
groups	a named list, of character vectors or numeric indices specifying node groupings. Each element of the list represent a group and contains a character vector with node names.
statName	a character, specifying the name of the statistic.
topN	a numeric, specifying the number of genes to label. The top genes are those with the largest count and statistic.

**Value**

a ggplot object, plotting the gene-level statistic against gene frequencies in the cluster of gene sets.

**Examples**

```
library(GSEABase)  
  
data(hgsc)  
groups <- list('g1' = names(hgsc)[1:25], 'g2' = names(hgsc)[26:50])  
  
#create statistics  
allgenes = unique(unlist(geneIds(hgsc)))  
gstats = rnorm(length(allgenes))  
names(gstats) = allgenes  
  
#plot  
plotGeneStats(gstats, hgsc, groups)
```

---

plotMsigNetwork	<i>Plot a gene set overlap network</i>
-----------------	--

---

### Description

Plots a network of gene set overlap with overlap computed using the `computeMsigOverlap()` and a graph created using `computeMsigNetwork()`.

### Usage

```
plotMsigNetwork(
  ig,
  markGroups = NULL,
  genesetStat = NULL,
  nodeSF = 1,
  edgeSF = 1,
  lytFunc = "graphopt",
  lytParams = list(),
  rmUnmarkedGroups = FALSE,
  maxGrp = 12
)
```

### Arguments

<code>ig</code>	an igraph object, containing a network of gene set overlaps computed using <code>computeMsigNetwork()</code> .
<code>markGroups</code>	a named list, of character vectors. Each element of the list represent a group and contains a character vector with node names. Up to 12 groups can be visualised in the plot.
<code>genesetStat</code>	a named numeric, statistic to project onto the nodes. These could be p-values, log fold-changes or gene set score from a singscore-based analysis.
<code>nodeSF</code>	a numeric, indicating the scaling factor to apply to node sizes.
<code>edgeSF</code>	a numeric, indicating the scaling factor to apply to edge widths.
<code>lytFunc</code>	a character, specifying the layout to use (see <code>ggraph::create_layout()</code> ).
<code>lytParams</code>	a named list, containing additional parameters needed for the layout (see <code>ggraph::create_layout()</code> ).
<code>rmUnmarkedGroups</code>	a logical, indicating whether unmarked groups should be removed from the network (TRUE) or retained (FALSE - default).
<code>maxGrp</code>	a numeric, specifying the maximum number of groups to plot.

### Value

a ggplot2 object

## Examples

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc, thresh = 0.15)
ig <- computeMsigNetwork(ovlap, hgsc)
groups <- list(
  'g1' = c("HALLMARK_HYPOXIA", "HALLMARK_GLYCOLYSIS"),
  'g2' = c("HALLMARK_INTERFERON_GAMMA_RESPONSE")
)

plotMsigNetwork(ig, markGroups = groups)
```

---

plotMsigPPI

---

*Plot PPI network for gene-set clusters identified using vissE*


---

## Description

This function plots the protein-protein interaction (PPI) network for a gene-set cluster identified using vissE. The international molecular exchange (IMEx) PPI is used to obtain PPIs for genes present in a gene-set cluster.

## Usage

```
plotMsigPPI(
  ppidf,
  msigGsc,
  groups,
  geneStat = NULL,
  statName = "Gene-level statistic",
  threshConfidence = 0,
  threshFrequency = 0.25,
  threshStatistic = 0,
  threshUseAbsolute = TRUE,
  topN = 5,
  nodeSF = 1,
  edgeSF = 1,
  lytFunc = "graphopt",
  lytParams = list()
)
```

## Arguments

ppidf	a data.frame, containing a protein-protein interaction from the IMEx database. This can be retrieved from the <code>msigdb::getIMEX()</code> function.
msigGsc	a GeneSetCollection object, containing gene sets from the MSigDB. The <code>GSEABase::getBroadSets()</code> function can be used to parse XML files downloaded from MSigDB.

<code>groups</code>	a named list, of character vectors or numeric indices specifying node groupings. Each element of the list represent a group and contains a character vector with node names.
<code>geneStat</code>	a named numeric, containing the statistic to be displayed. The vector must be named with either gene Symbols or Entrez IDs depending on annotations in <code>msigGsc</code> .
<code>statName</code>	a character, specifying the name of the statistic.
<code>threshConfidence</code>	a numeric, specifying the confidence threshold to apply to determine high confidence interactions. This should be a value between 0 and 1 (default is 0).
<code>threshFrequency</code>	a numeric, specifying the frequency threshold to apply to determine more frequent genes in the gene-set cluster. The frequency of a gene is computed as the proportion of gene-sets to which the gene belongs. This should be a value between 0 and 1 (default is 0.25).
<code>threshStatistic</code>	a numeric, specifying the threshold to apply to gene-level statistics (e.g. a log fold-change). This should be a value between 0 and 1 (default is 0).
<code>threshUseAbsolute</code>	a logical, indicating whether the <code>threshStatistic</code> threshold should be applied to absolute values (default TRUE). This can be used to threshold on statistics such as the log fold-change from a differential expression analysis.
<code>topN</code>	a numeric, specifying the number of genes to label. The top genes are those with the largest count and statistic.
<code>nodeSF</code>	a numeric, indicating the scaling factor to apply to node sizes.
<code>edgeSF</code>	a numeric, indicating the scaling factor to apply to edge widths.
<code>lytFunc</code>	a character, specifying the layout to use (see <code>ggraph::create_layout()</code> ).
<code>lytParams</code>	a named list, containing additional parameters needed for the layout (see <code>ggraph::create_layout()</code> ).

## Value

a `ggplot` object with the protein-protein interaction networks plot for each gene-set cluster.

## Examples

```
data(hgsc)
grps = list('early' = 'HALLMARK_ESTROGEN_RESPONSE_EARLY', 'late' = 'HALLMARK_ESTROGEN_RESPONSE_LATE')
ppi = msigdb::getIMEX(org = 'hs', inferred = TRUE)
plotMsigPPI(ppi, hgsc, grps)
```

---

plotMsigWordcloud	<i>Compute and plot word frequencies for multiple MSigDB collections</i>
-------------------	--

---

## Description

Given a gene set collection, this function computes the word frequency of gene set names from the Molecular Signatures Database (MSigDB) collection (split by `_`). Word frequencies are also computed using short descriptions attached with each gene set object.

## Usage

```
plotMsigWordcloud(
  msigGsc,
  groups,
  weight = NULL,
  measure = c("tfidf", "tf"),
  version = msigdb::getMsigdbVersions(),
  org = c("auto", "hs", "mm"),
  rmwords = getMsigExclusionList(),
  type = c("Name", "Short"),
  idf = NULL
)
```

## Arguments

<code>msigGsc</code>	a <code>GeneSetCollection</code> object, containing gene sets from the MSigDB. The <code>GSEABase::getBroadSets()</code> function can be used to parse XML files downloaded from MSigDB.
<code>groups</code>	a named list, of character vectors or numeric indices specifying node groupings. Each element of the list represent a group and contains a character vector with node names.
<code>weight</code>	a named numeric vector, containing weights to apply to each gene-set. This can be $-\log_{10}(\text{FDR})$ , $-\log_{10}(\text{p-value})$ or an enrichment score (ideally unsigned).
<code>measure</code>	a character, specifying how frequencies should be computed. "tf" uses term frequencies and "tfidf" (default) applies inverse document frequency weights to term frequencies.
<code>version</code>	a character, specifying the version of msigdb to use (see <code>msigdb::getMsigdbVersions()</code> ).
<code>org</code>	a character, specifying the organism to use. This can either be "auto" (default), "hs" or "mm".
<code>rmwords</code>	a character vector, containing an exclusion list of words to discard from the analysis.
<code>type</code>	a character, specifying the source of text mining. Either gene set names (Name) or descriptions (Short) can be used.

**idf** a list of named numeric vectors, specifying inverse document frequencies to use to penalise terms from gene-set names and short descriptions. This should be a vector of length 2 with names "Name" and "Short". Numeric vectors should contain weights and names should represent the term. Precomputed versions can be retrieved using the `msigdb::getMsigdbIDF()`.

**Value**

a ggplot object.

**Examples**

```
data("hgsc")
groups <- list('g1' = names(hgsc)[1:25], 'g2' = names(hgsc)[26:50])
plotMsigWordcloud(hgsc, groups, rmwords = getMsigExclusionList())
```



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