

# Package ‘cardelino’

November 20, 2024

**Type** Package

**Title** Clone Identification from Single Cell Data

**Version** 1.8.0

**Description** Methods to infer clonal tree configuration for a population of cells using single-cell RNA-seq data (scRNA-seq), and possibly other data modalities. Methods are also provided to assign cells to inferred clones and explore differences in gene expression between clones. These methods can flexibly integrate information from imperfect clonal trees inferred based on bulk exome-seq data, and sparse variant alleles expressed in scRNA-seq data. A flexible beta-binomial error model that accounts for stochastic dropout events as well as systematic allelic imbalance is used.

**License** GPL-3

**URL** <https://github.com/single-cell-genetics/cardelino>

**BugReports** <https://github.com/single-cell-genetics/cardelino/issues>

**Depends** R (>= 4.2), stats

**Imports** combinat, GenomeInfoDb, GenomicRanges, ggplot2, ggtree, Matrix, matrixStats, methods, pheatmap, snpStats, S4Vectors, utils, VariantAnnotation, vcfR

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**Enhances** doMC

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---

`assign_cells_to_clones`*Assign cells to clones from cardelino results*

---

**Description**

Assign cells to clones from cardelino results

**Usage**

```
assign_cells_to_clones(prob_mat, threshold = 0.5)
```

**Arguments**

<code>prob_mat</code>	numeric matrix (cells x clones) of clone posterior probabilities as output by <code>clone_id</code>
<code>threshold</code>	numeric(1), posterior probability threshold for cell-clone assignment: if posterior probability is above threshold, assign cell to clone, otherwise leave cell "unassigned"

**Value**

a data.frame with cell ID, assigned clone label and maximum posterior probability across clones.

**Author(s)**

Davis McCarthy

**Examples**

```
data(example_donor)
assignments <- clone_id(A_clone, D_clone, Config = tree$Z, inference = "EM")
df <- assign_cells_to_clones(assignments$prob)
head(df)
table(df$clone)
```

---

`assign_scores`*Scoring the simulation in assignment of singlets and doublets*

---

**Description**

Scoring the simulation in assignment of singlets and doublets

**Usage**

```
assign_scores(prob, I_sim, cutoff = seq(0, 1, 0.001))
```

**Arguments**

prob	Probability matrix for each cell to each component
I_sim	The true identity of assignment from simulation
cutoff	A list of cutoffs from 0 to 1

**Value**

A list with components: df\_sg, the recall/precision data.frame calculated by multiPRC(), AUC\_sg, the AUC calculated by multiPRC(), df\_db, the recall/precision data.frame calculated by binaryPRC() and AUC\_db the AUC calculated by binaryPRC(). Note that multiPRC() is run on a multiclass version of the problem and binaryPRC is run on a binarised version of the problem.

---

A\_clone

*A matrix of read numbers of alternative alleles for clone ID*


---

**Description**

This matrix contains read numbers of alternative alleles for 34 somatic variants across 428 cells, from one example scRNA-seq sample

**Usage**

```
example_donor
```

**Format**

a matrix of float

**Value**

NULL, but makes available a matrix

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

`A_germline`*A matrix of read numbers of alternative alleles*

---

**Description**

This matrix contains read numbers of alternative alleles for 34 germline variants (near the somatic variants) across 428 cells, from one example scRNA-seq sample

**Usage**

```
example_donor
```

**Format**

a matrix of float

**Value**

NULL, but makes available a matrix

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

`binaryPRC`*Precision-recall curve for binary label prediction*

---

**Description**

Precision-recall curve for binary label prediction

**Usage**

```
binaryPRC(  
  scores,  
  labels,  
  cutoff = NULL,  
  cut_direction = ">=",  
  add_cut1 = FALSE,  
  empty_precision = 1  
)
```

**Arguments**

scores	Prediction score for each sample
labels	True labels for each sample, e.g., from simulation
cutoff	A vector of cutoffs; if NULL use all unique scores
cut_direction	A string to compare with cutoff: >=, >, <=, <
add_cut1	Logical value; if True, manually add a cutoff of 1
empty_precision	Float value for default precision if no any recall

**Value**

A data.frame containing recall and precision values at various cutoffs.

**Examples**

```
scores <- 1:10
labels <- c(0, 0, 0, 1, 0, 1, 0, 1, 1, 1)
binaryPRC(scores, labels)

# Extra arguments.
binaryPRC(scores, labels, cutoff = seq(1, 10, by = 2))
binaryPRC(scores, labels, cut_direction = ">")
binaryPRC(scores, labels, add_cut1 = TRUE)
```

---

binaryROC

*ROC curve for binary label prediction*

---

**Description**

ROC curve for binary label prediction

**Usage**

```
binaryROC(
  scores,
  labels,
  cutoff = NULL,
  cut_direction = ">=",
  add_cut1 = TRUE,
  cutoff_point = 0.9
)
```

**Arguments**

scores	Prediction score for each sample
labels	True labels for each sample, e.g., from simulation
cutoff	A vector of cutoffs; if NULL use all unique scores
cut_direction	A string to compare with cutoff: >=, >, <=, <
add_cut1	Logical value; if True, manually add a cutoff of 1
cutoff_point	Numeric value; additional cutoff value

**Value**

A data.frame containing AUC and AUPRC at various cutoffs.

**Examples**

```
scores <- 1:10
labels <- c(0, 0, 0, 1, 0, 1, 0, 1, 1, 1)
binaryROC(scores, labels)

# Extra arguments.
binaryROC(scores, labels, cutoff = seq(1, 10, by = 2))
binaryROC(scores, labels, cut_direction = ">")
binaryROC(scores, labels, add_cut1 = TRUE)
```

---

Clone ID

*Infer clonal identity of single cells*


---

**Description**

Infer clonal identity of single cells

Assign cells to clones using an EM algorithm

Assign cells to clones using a Gibbs sampling algorithm

**Usage**

```
clone_id(
  A,
  D,
  Config = NULL,
  n_clone = NULL,
  Psi = NULL,
  relax_Config = TRUE,
  relax_rate_fixed = NULL,
  inference = "sampling",
  n_chain = 1,
  n_proc = 1,
  verbose = TRUE,
  ...
)

clone_id_EM(
  A,
  D,
  Config,
  Psi = NULL,
  min_iter = 10,
  max_iter = 1000,
  logLik_threshold = 1e-05,
  verbose = TRUE
)
```

```
clone_id_Gibbs(
  A,
  D,
  Config,
  Psi = NULL,
  relax_Config = TRUE,
  relax_rate_fixed = NULL,
  relax_rate_prior = c(1, 9),
  keep_base_clone = TRUE,
  prior0 = c(0.2, 99.8),
  prior1 = c(0.45, 0.55),
  min_iter = 5000,
  max_iter = 20000,
  buin_frac = 0.5,
  wise = "variant",
  relabel = FALSE,
  verbose = TRUE
)
```

### Arguments

A	variant x cell matrix of integers; number of alternative allele reads in variant i cell j
D	variant x cell matrix of integers; number of total reads covering variant i cell j
Config	variant x clone matrix of binary values. The clone-variant configuration, which encodes the phylogenetic tree structure. This is the output Z of Canopy
n_clone	integer(1), the number of clone to reconstruct. This is in use only if Config is NULL
Psi	A vector of float. The fractions of each clone, output P of Canopy
relax_Config	logical(1), If TRUE, relaxing the Clone Configuration by changing it from fixed value to act as a prior Config with a relax rate.
relax_rate_fixed	numeric(1), If the value is between 0 to 1, the relax rate will be set as a fix value during updating clone Config. If NULL, the relax rate will be learned automatically with relax_rate_prior.
inference	character(1), the method to use for inference, either "sampling" to use Gibbs sampling (default) or "EM" to use expectation-maximization (faster)
n_chain	integer(1), the number of chains to run, which will be averaged as an output result
n_proc	integer(1), the number of processors to use. This parallel computing can largely reduce time when using multiple chains
verbose	logical(1), should the function output verbose information as it runs?
...	arguments passed to <code>clone_id_Gibbs</code> or <code>clone_id_EM</code> (as appropriate)
min_iter	A integer. The minimum number of iterations in the Gibbs sampling. The real iteration may be longer until the convergence.
max_iter	A integer. The maximum number of iterations in the Gibbs sampling, even haven't passed the convergence diagnosis



logLik_threshold	A float. The threshold of logLikelihood increase for detecting convergence.
relax_rate_prior	numeric(2), the two parameters of beta prior distribution of the relax rate for relaxing the clone Configuration. This mode is used when relax_relax is NULL.
keep_base_clone	bool(1), if TRUE, keep the base clone of Config to its input values when relax mode is used.
prior0	numeric(2), alpha and beta parameters for the Beta prior distribution on the inferred false positive rate.
prior1	numeric(2), alpha and beta parameters for the Beta prior distribution on the inferred (1 - false negative) rate.
buin_frac	numeric(1), the fraction of chain as burn-in period
wise	A string, the wise of parameters for theta1: global, variant, element.
relabel	bool(1), if TRUE, relabel the samples of both Config and prob during the Gibbs sampling.

### Details

The two Bernoulli components correspond to false positive and false negative rates. The two binomial components correspond to the read distributions with and without the mutation present.

### Value

If inference method is "EM", a list containing theta, a vector of two floats denoting the parameters of the two components of the base model, i.e., mean of Bernoulli or binomial model given variant exists or not, prob, the matrix of posterior probabilities of each cell belonging to each clone with fitted parameters, and logLik, the log likelihood of the final parameters.

If inference method is "sampling", a list containing: theta0, the mean of sampled false positive parameter values; theta1 the mean of sampled (1 - false negative rate) parameter values; theta0\_all, all sampled false positive parameter values; theta1\_all, all sampled (1 - false negative rate) parameter values; element; logLik\_all, log-likelihood for model for all sampled parameter sets; prob\_all; prob, matrix with mean of sampled cell-clone assignment posterior probabilities (the key output of the model); prob\_variant.

a list containing theta, a vector of two floats denoting the binomial rates given variant exists or not, prob, the matrix of posterior probabilities of each cell belonging to each clone with fitted parameters, and logLik, the log likelihood of the final parameters.

### Author(s)

Yuanhua Huang and Davis McCarthy

Yuanhua Huang

### Examples

```
data(example_donor)
assignments <- clone_id(A_clone, D_clone,
  Config = tree$Z,
  min_iter = 800, max_iter = 1200
)
prob_heatmap(assignments$prob)
```

```

assignments_EM <- clone_id(A_clone, D_clone,
  Config = tree$Z,
  inference = "EM"
)
prob_heatmap(assignments_EM$prob)

```

---

colMatch	<i>Column match between two matrices by minimum mean absolute difference</i>
----------	--

---

### Description

Column match between two matrices by minimum mean absolute difference

### Usage

```
colMatch(A, B, force = FALSE)
```

### Arguments

A	The first matrix which will be matched
B	The second matrix, the return index will be used on
force	bool(1), If TRUE, force traversing all permutations of B to find the optimised match to A with computing cost of $O(n!)$ . Otherwise, use greedy search with computing cost of $O(n^2)$ .

### Value

idx, the column index of B to be matched to A

### Examples

```

matA <- matrix(sample(seq(12)), nrow = 3)
col_idx <- sample(4)
matB <- matA[, col_idx]
colMatch(matB, matA)

```

---

Config_all	<i>A list of tree configuration</i>
------------	-------------------------------------

---

### Description

This list of tree configuration between 3 clones to 10 clones, each element is a list with all possible tree matrix

### Usage

```
config_all
```

**Format**

a list of list of matrix

**Value**

NULL, but makes available a list

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

PASTRI Python package

---

devianceIC

*Deviance Information Criterion for cardelino model*

---

**Description**

Deviance Information Criterion for cardelino model

**Usage**

```
devianceIC(logLik_all, logLik_post)
```

**Arguments**

logLik\_all      A vector of numeric; the log likelihood of posterior sample, i.e., posterior samples of deviance

logLik\_post     numeric(1); the log likelihood of mean posterior parameters, i.e., deviance of posterior means

**Value**

DIC, a float of deviance information criterion

**Author(s)**

Yuanhua Huang

---

donor\_read\_simulator *Reads simulator for donor identification*

---

### Description

Reads simulator for donor identification

### Usage

```
donor_read_simulator(
  GT,
  D_seed,
  sample_variants = FALSE,
  donor_size = NULL,
  beta_shapes = NULL,
  n_cell = 5000,
  doublet_rate = NULL
)
```

### Arguments

GT	Variant-by-donor matrix for genotypes
D_seed	Variant-by-cell matrix for read coverage for generating depth, which be row sample and column sample both with replacement
sample_variants	logical(1), if TRUE, sample variants with replacement to the same size, otherwise not
donor_size	Vector of float for the fractions of each donor; default NULL means uniform
beta_shapes	A 3-by-2 matrix of beta parameters for genotypes: 0, 1, and 2; default NULL means matrix(c(0.2, 0.5, 99.8, 99.8, 0.5, 0.2), nrow = 3)
n_cell	An integer for number of total cells
doublet_rate	A float from 0 to 1 for doublet rate; default NULL means rate n_cell / 100000

### Value

A list of various components of the simulated dataset.

---

D\_clone *A matrix of sequencing depths for clone ID*

---

### Description

This matrix contains sequencing depths for 34 somatic variants across 428 cells, from one example scRNA-seq sample

### Usage

```
example_donor
```

**Format**

a matrix of float

**Value**

NULL, but makes available a matrix

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

<i>D_germline</i>	<i>A matrix of sequencing depths</i>
-------------------	--------------------------------------

---

**Description**

This matrix contains sequencing depths for 34 germline variants (near the somatic variants) across 428 cells, from one example scRNA-seq sample

**Usage**

example\_donor

**Format**

a matrix of float

**Value**

NULL, but makes available a matrix

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

D_input	<i>A matrix of sequencing depths</i>
---------	--------------------------------------

---

**Description**

This matrix contains sequencing depths for 439 somatic variants across 151 cells, from one particular scRNA-seq sample, can be used to generate sequencing depths

**Usage**

```
simulation_input
```

**Format**

a matrix of float

**Value**

NULL, but makes available a matrix

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

get_logLik	<i>Log likelihood of clone_id model It returns <math>P(A, D \mid C, I, \theta_0, \theta_1)</math></i>
------------	---

---

**Description**

Log likelihood of clone\_id model It returns  $P(A, D \mid C, I, \theta_0, \theta_1)$

**Usage**

```
get_logLik(A1, B1, Config, Assign, theta0, theta1)
```

**Arguments**

A1	variant x cell matrix of integers; number of alternative allele reads in variant i cell j
B1	variant x cell matrix of integers; number of reference allele reads in variant i cell j
Config	variant x clone matrix of float values. The clone-variant configuration probability, averaged by posterior samples
Assign	cells x clone matrix of float values. The cell-clone assignment probability, averaged by posterior samples
theta0	the binomial rate for alternative allele from config = 0
theta1	the binomial rate for alternative allele from config = 1

**Value**

logLik, a float of log likelihood

**Author(s)**

Yuanhua Huang

---

get_snp_matrices	<i>Get SNP data matrices from VCF object(s)</i>
------------------	---

---

**Description**

Get SNP data matrices from VCF object(s)

**Usage**

```
get_snp_matrices(vcf_cell, vcf_donor = NULL, verbose = TRUE, donors = NULL)
```

**Arguments**

vcf_cell	a <a href="#">CollapsedVCF</a> object containing variant data for cells
vcf_donor	an optional <a href="#">CollapsedVCF</a> object containing genotype data for donors
verbose	logical(1), should the function output verbose information as it runs?
donors	optional character vector providing a set of donors to use, by subsetting the donors present in the donor_vcf_file; if NULL (default) then all donors present in VCF will be used.

**Value**

a list containing A, a matrix of integers. Number of alteration reads in SNP i cell j. D, a matrix of integers. Number of reads depth in SNP i cell j. R, a matrix of integers. Number of reference reads in SNP i cell j. GT\_cells, a matrix of integers for genotypes. The cell-SNP configuration. GT\_donors, a matrix of integers for genotypes. The donor-SNP configuration.

**Examples**

```
vcf_cell <- read_vcf(system.file("extdata", "cells.donorid.vcf.gz",
                               package = "cardelino"))
vcf_donor <- read_vcf(system.file("extdata", "donors.donorid.vcf.gz",
                                 package = "cardelino"))
snp_data <- get_snp_matrices(vcf_cell, vcf_donor)
```

---

`get_tree`*Get a clonal tree from a configuration matrix*

---

**Description**

Get a clonal tree from a configuration matrix

**Usage**

```
get_tree(Config, P = NULL, strictness = "lax")
```

**Arguments**

<code>Config</code>	variant x clone matrix of binary values. The clone-variant configuration, which encodes the phylogenetic tree structure. This is the output Z of Canopy
<code>P</code>	a one-column numeric matrix encoding the (observed or estimated) prevalence (or frequency) of each clone
<code>strictness</code>	character(1), a character string defining the strictness of the function if there are all-zero rows in the Config matrix. If "lax" then the function silently drops all-zero rows and proceeds. If "warn" then the function warns of dropping all-zero rows and proceeds. If "error" then the function throws an error if all-zero rows are detected.

**Details**

Output tree may be nonsensical if the input Config matrix does not define a coherent tree structure.

**Value**

An object of class "phylo" describing the tree structure. The output object also contains an element "sna" defining the clustering of variants onto the branches of the tree, and if P is non-null it also contains VAF (variant allele frequency), CCF (cell clone fraction) and clone prevalence values (computed from the supplied P argument).

**Author(s)**

Davis McCarthy

**Examples**

```
Config3 <- matrix(c(
  rep(0, 15), rep(1, 8), rep(0, 7), rep(1, 5), rep(0, 3),
  rep(1, 7)
), ncol = 3)
tree_k3 <- get_tree(Config = Config3, P = matrix(rep(1 / 3, 3), ncol = 1))
plot_tree(tree_k3)
```



---

Geweke_Z	<i>Geweke diagnostic for MCMC sampling.</i>
----------	---

---

**Description**

Geweke diagnostic for MCMC sampling.

**Usage**

```
Geweke_Z(X, first = 0.1, last = 0.5)
```

**Arguments**

X	A matrix of MCMC samples for N samples per K variables
first	A float between 0 and 1. The initial region of MCMC chain.
last	A float between 0 and 1. The final region of MCMC chain.

**Value**

Z, a vector of absolute value of Z scores for each variable. When  $|Z| \leq 2$ , the sampling could be taken as converged.

**Author(s)**

Yuanhua Huang

---

heatmap.theme	<i>The theme of heatmaps for prob_heatmap and sites_heatmap</i>
---------------	---

---

**Description**

The theme of heatmaps for prob\_heatmap and sites\_heatmap

**Usage**

```
heatmap.theme(legend.position = "bottom", size = 12)
```

**Arguments**

legend.position	character, describes where to place legend on plot (passed to <a href="#">theme_gray</a> )
size	numeric, base font size for plot (passed to <a href="#">theme_gray</a> )

**Value**

a ggplot theme based on [theme\\_gray](#)

---

heat_matrix	<i>Plot heatmap from a matrix</i>
-------------	-----------------------------------

---

**Description**

Plot heatmap from a matrix

**Usage**

```
heat_matrix(mat, base_size = 12, digits = 2, show_value = FALSE)
```

**Arguments**

mat	A matrix to show, column by x-axis and row by y-axis
base_size	Numeric value for the base size in theme_bw
digits	Integer value for the number of digits to show
show_value	Logical value for showing the value for each element or not

**Value**

A ggplot heatmap visualization of the passed matrix.

**Examples**

```
mat <- matrix(rnorm(9), ncol = 3, nrow = 3) + diag(rnorm(3, 2, 0.1))
rownames(mat) <- paste0("sample_", letters[1:3])
colnames(mat) <- paste0("var_", 1:3)
heat_matrix(mat)

# Additional arguments.
heat_matrix(mat, base_size = 6)
heat_matrix(mat, show_value = TRUE)
heat_matrix(mat, show_value = TRUE, digits = 4)
```

---

load_cellSNP_vcf	<i>Load sparse matrices A and D from cellSNP VCF file with filtering SNPs</i>
------------------	---

---

**Description**

Load sparse matrices A and D from cellSNP VCF file with filtering SNPs

**Usage**

```
load_cellSNP_vcf(
  vcf_file,
  min_count = 0,
  min_MAF = 0,
  max_other_allele = NULL,
  rowname_format = "full",
  keep_GL = FALSE
)
```

**Arguments**

vcf_file	character(1), path to VCF file generated from cellSNP
min_count	minimum count across all cells, e.g., 20
min_MAF	minimum minor allele fraction, e.g., 0.1
max_other_allele	maximum ratio of other alleles comparing to REF and ALT alleles; for cellSNP vcf, we recommend 0.05
rowname_format	the format of rowname: NULL is the default from vcfR, short is CHROM_POS, and full is CHROM_POS_REF_ALT
keep_GL	logical(1), if TRUE, check if GL (genotype probability) exists it will be returned

**Value**

A list with elements the matrices A and D and GL, the genotype probability. If keep\_GL is false the GL element will be an empty list.

**Examples**

```
vcf_file <- system.file("extdata", "cellSNP.cells.vcf.gz",
  package = "cardelino"
)
input_data <- load_cellSNP_vcf(vcf_file)
```

---

load_GT_vcf	<i>Load genotype VCF into numeric values: 0, 1, or 2</i>
-------------	--

---

**Description**

Note, the genotype VCF can be very big for whole genome. It would be more efficient to only keep the wanted variants and samples. bcftools does such jobs nicely.

**Usage**

```
load_GT_vcf(vcf_file, rowname_format = "full", na.rm = TRUE, keep_GP = TRUE)
```

**Arguments**

vcf_file	character(1), path to VCF file for donor genotypes
rowname_format	the format of rowname: NULL is the default from vcfR, short is CHROM_POS, and full is CHROM_POS_REF_ALT
na.rm	logical(1), if TRUE, remove the variants with NA values
keep_GP	logical(1), if TRUE, check if GP (genotype probability) exists it will be returned

**Value**

A list representing the loaded genotype information with two components: GT, the usual numeric representation of genotype and GP the genotype probabilities. Note that if keep\_GP is false the GP component will be NULL.

**Examples**

```
vcf_file <- system.file("extdata", "cellSNP.cells.vcf.gz",
  package = "cardelino"
)
GT_dat <- load_GT_vcf(vcf_file, na.rm = FALSE)
```

---

mixBinom

*EM algorithm for estimating binomial mixture model*


---

**Description**

EM algorithm for estimating binomial mixture model

**Usage**

```
mixBinom(
  k,
  n,
  n_components = 2,
  p_init = NULL,
  learn_p = TRUE,
  min_iter = 10,
  max_iter = 1000,
  logLik_threshold = 1e-05
)
```

**Arguments**

k	A vector of integers. number of success
n	A vector of integers. number of trials
n_components	A number. number of components
p_init	A vector of floats with length n_components, the initial value of p
learn_p	bool(1) or a vector of bool, whether learn each p
min_iter	integer(1). number of minimum iterations
max_iter	integer(1). number of maximum iterations
logLik_threshold	A float. The threshold of logLikelihood increase for detecting convergence

**Value**

a list containing p, a vector of floats between 0 and 1 giving the estimated success probability for each component, psi, estimated fraction of each component in the mixture, and prob, the matrix of fitted probabilities of each observation belonging to each component.

**Examples**

```
n1 <- array(sample(1:30, 50, replace = TRUE))
n2 <- array(sample(1:30, 200, replace = TRUE))
k1 <- apply(n1, 1, rbinom, n = 1, p = 0.5)
k2 <- apply(n2, 1, rbinom, n = 1, p = 0.01)
RV <- mixBinom(c(k1, k2), c(n1, n2))
```

---

mtx_to_df	<i>Convert a matrix to data frame</i>
-----------	---------------------------------------

---

**Description**

Convert a matrix to data frame

**Usage**

```
mtx_to_df(X)
```

**Arguments**

X                    A matrix of values

**Value**

A data.frame version of the passed matrix.

**Examples**

```
mtx_to_df(matrix(seq(12), nrow = 3))
```

---

multiPRC	<i>Precision-recall curve for multi-class prediction</i>
----------	--

---

**Description**

Precision-recall curve for multi-class prediction

**Usage**

```
multiPRC(
  prob_mat,
  simu_mat,
  marginal_mode = "best",
  cutoff = NULL,
  multiLabel.rm = TRUE,
  add_cut1 = FALSE
)
```

**Arguments**

prob_mat	Probability matrix for each cell to each component
simu_mat	The true identity of assignment from simulation
marginal_mode	A string for the mode to marginalize the column: best, second, or delta
cutoff	A list of cutoff; if NULL use all unique scores
multiLabel.rm	Logical value; if True, remove the samples with multiple labels
add_cut1	Logical value; if True, manually add a cutoff of 1

**Value**

A list with two components: `df`, a `data.frame` containing precision and recall values at various cutoffs and AUC, the overall AUC.

---

<code>plot_config_diffs</code>	<i>Define a publication-style plot theme</i>
--------------------------------	--

---

**Description**

Define a publication-style plot theme

**Usage**

```
plot_config_diffs(Config1, Config2, show_variant_names = FALSE)
```

**Arguments**

`Config1` variant by clone matrix defining the first clonal structure

`Config2` variant by clone matrix defining the second clonal structure

`show_variant_names` `logical(1)`, should the variant names (rownames of Config matrices) be shown on the plot? Default is FALSE.

**Value**

a `ggplot` heatmap style plot showing the differences between the two Config matrices, specifically the differences `Config1 - Config2`.

**Examples**

```
Config1 <- matrix(c(
  rep(0, 15), rep(1, 8), rep(0, 7),
  rep(1, 5), rep(0, 3), rep(1, 7)
), ncol = 3)
Config2 <- matrix(c(
  rep(0, 15), rep(1, 8), rep(1, 7),
  rep(0, 5), rep(1, 3), rep(1, 7)
), ncol = 3)
rownames(Config1) <- rownames(Config2) <- paste0("var", 1:nrow(Config1))
colnames(Config1) <- colnames(Config2) <- paste0("clone", 1:ncol(Config1))
plot_config_diffs(Config1, Config2)
```

---

plot_tree	<i>Plot a phylogenetic tree</i>
-----------	---------------------------------

---

### Description

Plot a phylogenetic tree

### Usage

```
plot_tree(tree, orient = "h")
```

### Arguments

tree	A phylogenetic tree object of class "phylo"
orient	A string for the orientation of the tree: "v" (vertical) or "h" (horizontal)

### Details

This function plots a phylogenetic tree from an object of class "phylo", as produced, for example, by the Canopy package.

### Value

a ggtree object

### Author(s)

Davis McCarthy and Yuanhua Huang

### References

This function makes use of the [ggtree](#) package:

Guangchuang Yu, David Smith, Huachen Zhu, Yi Guan, Tommy Tsan-Yuk Lam. ggtree: an R package for visualization and annotation of phylogenetic trees with their covariates and other associated data. *Methods in Ecology and Evolution* 2017, 8(1):28-36, doi:10.1111/2041-210X.12628

### Examples

```
data(example_donor)
plot_tree(tree, orient = "v")
```

---

predMixBinom                      *Predicted probability from learned binomial mixture model*

---

### Description

Predicted probability from learned binomial mixture model

### Usage

```
predMixBinom(k, n, p, psi)
```

### Arguments

k                      A vector of integers. number of success  
n                      A vector of integers. number of trials  
p                      a vector of binomial success probabilities  
psi                    A float between 0 and 1. fraction of each component

### Value

A list with two components: prob, a matrix representing the probability of each of the passed values coming from each component of the mixture and logLik, the total log-likelihood of the new samples.

### Examples

```
n1 <- array(sample(1:30, 50, replace = TRUE))
n2 <- array(sample(1:30, 200, replace = TRUE))
k1 <- apply(n1, 1, rbinom, n = 1, p = 0.5)
k2 <- apply(n2, 1, rbinom, n = 1, p = 0.01)
RV <- mixBinom(c(k1, k2), c(n1, n2))
RV_pred <- predMixBinom(3, 10, RV$p, RV$psi)
```

---

prob\_heatmap                      *Plot a heatmap for probability of clone assignment*

---

### Description

Plot a heatmap for probability of clone assignment

### Usage

```
prob_heatmap(prob_mat, threshold = 0.5, mode = "best", cell_idx = NULL)
```

### Arguments

prob\_mat              A matrix (M x K), the probability of cell j to clone k  
threshold            A float value, the threshold for assignable cells  
mode                  A string, the method for defining scores for filtering cells: best and delta. best: highest probability of a cell to K clones, delta: the difference between the best and second.  
cell\_idx              A vector the indices of the input cells. If NULL, order by the probability of each clone



**Value**

a ggplot object

**Examples**

```
data(example_donor)
assignments <- clone_id(A_clone, D_clone, Config = tree$Z, inference = "EM")
fig <- prob_heatmap(assignments$prob)
```

---

pub.theme

*Define a publication-style plot theme*

---

**Description**

Define a publication-style plot theme

**Usage**

```
pub.theme(size = 12)
```

**Arguments**

size            numeric, base font size for adapted ggplot2 theme

**Details**

This theme modifies the [theme\\_classic](#) theme in ggplot2.

**Value**

a ggplot theme based on theme\_classic

**Examples**

```
library(ggplot2)
x <- sample(10)
y <- x + runif(10) - 0.5
df <- data.frame(x = x, y = y)
fig <- ggplot(df, aes(x = x, y = y)) +
  geom_point() +
  pub.theme()
```

---

read_vcf	<i>Read a VCF file into R session</i>
----------	---------------------------------------

---

**Description**

Read a VCF file into R session

**Usage**

```
read_vcf(
  vcf_file,
  genome = "GRCh37",
  seq_levels_style = "Ensembl",
  verbose = TRUE
)
```

**Arguments**

vcf_file	character(1), path to VCF file to read into R session as a <a href="#">CollapsedVCF</a> object
genome	character(1), string indicating the genome build used in the VCF file(s) (default: "GRCh37")
seq_levels_style	character(1), string passed to <a href="#">seqlevelsStyle</a> the style to use for chromosome/contig names (default: "Ensembl")
verbose	logical(1), should messages be printed as function runs?

**Value**

a vcf object

**Examples**

```
vcf <- read_vcf(system.file("extdata", "cells.donorid.vcf.gz",
  package = "cardelino"))
```

---

rowArgmax	<i>Column index of the maximum value for each row in a matrix</i>
-----------	---

---

**Description**

Column index of the maximum value for each row in a matrix

**Usage**

```
rowArgmax(X)
```

**Arguments**

X	A matrix of floats.
---	---------------------

**Value**

a vector of the index of column for each row. Note, when multiple columns have the same value, only the earliest column will be returned.

**Examples**

```
matA <- matrix(sample(seq(12)), nrow = 3)
rowArgmax(matA)
```

---

rowMax	<i>Maximum value for each row in a matrix</i>
--------	---

---

**Description**

Maximum value for each row in a matrix

**Usage**

```
rowMax(X, mode = "best")
```

**Arguments**

X	A matrix of floats.
mode	A string, the method for defining scores for filtering cells: best, second and delta. best: highest value for each row, similarly for the second. delta is the difference between the best and the second.

**Value**

a vector of the collapsed value for each row, depending on the mode used.

**Examples**

```
matA <- matrix(sample(seq(12)), nrow = 3)
rowMax(matA)
```

---

sample_seq_depth	<i>Update matrix D with manually selected missing rate</i>
------------------	--

---

**Description**

Given missing rate, the NA will be generated first. For none NA element, sequencing depth with uniformly sampled from D, row wisely. Namely, the depth is variant specific.

**Usage**

```
sample_seq_depth(D, n_cells = NULL, n_sites = NULL, missing_rate = NULL)
```

**Arguments**

D	A matrix (N variants x M cells), the original sequencing coverage, NA means missing
n_cells	A integer, the number of the cells to generate
n_sites	A integer, the number of variants to generate
missing_rate	A float value, if NULL, use the same missing rate as D

**Value**

a n\_sites by n\_cells matrix sampled from input D.

**Examples**

```
data(simulation_input)
D1 <- sample_seq_depth(D_input,
  n_cells = 500, n_sites = 50,
  missing_rate = 0.85
)
```

---

sample\_tree\_SNV

*Down sample number of SNVs in the tree*

---

**Description**

Down sample number of SNVs in the tree

**Usage**

```
sample_tree_SNV(tree, n_SNV = NULL)
```

**Arguments**

tree	A tree object from Canopy
n_SNV	A integer, the number of SNVs to keep in the output tree

**Value**

a phylo tree with down sampled variants

**Examples**

```
data(simulation_input)
tree_lite <- sample_tree_SNV(tree_4clone, n_SNV = 10)
```

sim\_read\_count

*Synthetic reads generator for genetic variants***Description**

There are following steps to generate the simulated reads counts for variants in single cells: 1) given the clonal genotype and the clonal prevalence, the genotypes (i.e, the clone) of cells will be generated following a multinomial distribution. Note, one cell may contain variants from two clones when it is a doublet. 2) given the distribution of reads coverage, e.g., a matrix of read coverage from real data, (variant specific), the total reads of each variant will be generated by random sampling. Note, the missing rate is governed by this matrix. 3) the allelic frequency of each variant will be generated by following a beta distribution with parameters of mean and variance. 4) Given the genotype of a cell, if the mutation exists in a cell, the alteration read counts will be generated by a binomial distribution, parameterized the allelic frequency, sampled from step 3. 5) Given the genotype of a cell, if the mutation does not exist in a cell, the alteration read counts will be generated by a binomial distribution, parameterized by the technical error rate.

**Usage**

```
sim_read_count(
  Config,
  D,
  Psi = NULL,
  means = c(0.002, 0.45),
  vars = c(100, 1),
  wise0 = "element",
  wise1 = "variant",
  cell_num = 300,
  permute_D = FALSE,
  sample_cell = TRUE,
  doublet = 0
)
```

**Arguments**

Config	A matrix of binary values. The clone-variant configuration, which encodes the phylogenetic tree structure, and the genotype of each clone
D	A matrix of integers. Sequencing depth for N variants across x cells (ideally >100 cells). NA means 0 here.
Psi	A vector of float. The fractions of each clone. If NULL, set a uniform distribution.
means	A vector of two floats. The mean theta_1 (false positive rate) and the mean theta_2 (true positive rate).
vars	A vector of two floats. The variance of theta_1 and theta_2.
wise0	A string, the beta-binomial parameter specificity for theta0: global, variant, element.
wise1	A string, the beta-binomial parameter specificity for theta1: global, variant, element.
cell_num	A integer. The number of cells to generate.

permute_D	A Boolean value. If True permute variants in D.
sample_cell	A Boolean value. If True and $M > \text{ncol}(D)$ , sample cells.
doublet	A float between 0 and 1, the rate of doublets

**Value**

a list containing `A_sim`, a matrix for alteration reads, `A_sim`, a matrix for total reads, `I_sim`, a matrix for clonal label, `H_sim`, a matrix for genotype, `theta0`, a matrix of expected false positive rate, `theta1`, a matrix of expected true positive rate, `theta0_binom`, `theta0` as binomial parameter, `theta1_binom`, `theta0` as binomial parameter, and `is_doublet`, a vector of Boolean value if a cell is a doublet

**Examples**

```
data(simulation_input)
D2 <- sample_seq_depth(D_input, n_cells = 500, n_sites = nrow(tree_4clone$Z))
simu <- sim_read_count(tree_4clone$Z, D2, Psi = NULL, cell_num = 500)
```

---

tree

*A tree object*

---

**Description**

This tree object contains clonal tree information, inferred from bulk exome-seq data

**Usage**

```
example_donor
```

**Format**

a tree object

**Value**

NULL, but makes available a tree object

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

tree_3clone	<i>A tree object</i>
-------------	----------------------

---

**Description**

This tree object with 3 clones contains clonal tree information, inferred from bulk exome-seq data

**Usage**

simulation\_input

**Format**

a tree object

**Value**

NULL, but makes available a tree object

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

tree_4clone	<i>A tree object</i>
-------------	----------------------

---

**Description**

This tree object with 4 clones contains clonal tree information, inferred from bulk exome-seq data

**Usage**

simulation\_input

**Format**

a tree object

**Value**

NULL, but makes available a tree object

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

tree_5clone	<i>A tree object</i>
-------------	----------------------

---

**Description**

This tree object with 5 clones contains clonal tree information, inferred from bulk exome-seq data

**Usage**

```
simulation_input
```

**Format**

a tree object

**Value**

NULL, but makes available a tree object

**Author(s)**

Yuanhua Huang, Davis McCarthy, 2018-06-25

**Source**

A fibroblast sample from HipSci project

---

vc_heatmap	<i>Plot a variant-cell heatmap for cell clonal assignment</i>
------------	---

---

**Description**

Plot a variant-cell heatmap for cell clonal assignment

**Usage**

```
vc_heatmap(mat, prob, Config, show_legend = FALSE)
```

**Arguments**

mat	A matrix for heatmap: N variants x M cells. row and column will be sorted automatically.
prob	A matrix of probability of clonal assignment: M cells x K clones
Config	A binary matrix of clonal Configuration: N variants x K clones
show_legend	A bool value: if TRUE, show the legend

**Value**

a pheatmap object  
a ggplot object



## **References**

This function makes use of the [pheatmap](#) packages

## **Examples**

```
data(example_donor)
assignments <- clone_id(A_clone, D_clone, Config = tree$Z)
fig <- vc_heatmap(assignments$prob_variant, assignments$prob, tree$Z)
```

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