# Package 'CGHnormaliter'

November 20, 2024

Type Package
Title Normalization of array CGH data with imbalanced aberrations.
<b>Version</b> 1.60.0
<b>Date</b> 2012-06-12
Author Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling
Maintainer Bart P.P. van Houte  bvhoute@few.vu.nl>
<b>Depends</b> CGHcall (>= 2.17.0), CGHbase (>= 1.15.0)
Imports Biobase, CGHbase, CGHcall, methods, stats, utils
<b>Description</b> Normalization and centralization of array comparative genomic hybridization (aCGH) data. The algorithm uses an iterative procedure that effectively eliminates the influence of imbalanced copy numbers. This leads to a more reliable assessment of copy number alterations (CNAs).
License GPL (>= 3)
biocViews Microarray, Preprocessing
git_url https://git.bioconductor.org/packages/CGHnormaliter
git_branch RELEASE_3_20
git_last_commit a252ff1
git_last_commit_date 2024-10-29
Repository Bioconductor 3.20
Date/Publication 2024-11-20
Contents
CGHnormaliter-package
CGHnormaliter-internal
CGHnormaliter.write.table
Leukemia
Index

CGHnormaliter-package This package implements the CGHnormaliter algorithm which is a strategy for improved normalization of array Comparative Genomic Hybridization (aCGH) data

# **Description**

CGHnormaliter is an iterative algorithm for normalization of aCGH data displaying imbalanced aberrations. First, provisory balanced copy numbers are identified and subsequently used for normalization based on LOWESS. These two steps are then iterated to refine the normalization. The assumption here is that the temporary exclusion of aberrations allows for a more appropriate calculation of the LOWESS regression curve. As a result, after normalization, the log2 intensity ratios of the normals will generally be closer to zero and better reflect the biological reality.

#### **Details**

Package: CGHnormaliter

Type: Package Version: 1.5.6 Date: 2011-04-06

Date: 2011-04-06 License: GPL (>= 3)

The package contains two public functions. The function CGHnormaliter performs the iterative normalization of aCGH data, while the function CGHnormaliter.write.table prints normalized aCGH data to a file. See function documentation for details.

# Author(s)

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling

#### References

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling, Walter Pirovano and Jaap Heringa. CGH-normaliter: an iterative strategy to enhance normalization of array CGH data with imbalanced aberrations. BMC Genomics, 10:401, 2009.

#### See Also

CGHcall, DNAcopy

CGHnormaliter 3

naliter Iterative normalization of aCGH da
--

# **Description**

Implementation of an iterative algorithm for normalization of aCGH data displaying imbalanced aberrations.

#### Usage

```
CGHnormaliter(data, nchrom = 24, cellularity = 1, max.losses = 0.3, plot.MA = TRUE, ...)
```

#### **Arguments**

data	either a dataframe or character string containing a filename. See section Details for the format.
nchrom	number of chromosomes.
cellularity	vector of cellularities ranging from $0$ to $1$ to define the contamination of your sample with healthy cells ( $1 = \text{no contamination}$ ). See Details for more information.
max.losses	vector of maximum fraction of losses in your samples, ranging from 0 to 1. In case this fraction is exceeded after calling, losses are deemed normals and centralization will be around the (lower) peak at the left of the log2 ratio distribution. See Details for more information.
plot.MA	logical. If 'plot.MA = TRUE', MA-plots before and after normalization are generated of each sample. The plots are saved into a PDF file.
	arguments for segment (from package DNAcopy) and/or CGHcall.

#### **Details**

The input should be either a data. frame or the file name of a tabseparated text file (text files must contain a header). The first four columns should contain the name, chromosome and the start and end position in bp for each array target respectively. The position columns must contain numbers only. Following these are two columns with the raw test and reference intensities for each of your samples. These intensities must be numeric as well. If the input type is a text file, missing values should be represented as 'NA' or an empty field.

The cellularity and max.losses parameters should both be a vector of length n where n is the number of samples in your dataset. Each vector is recycled if there are not enough values in it, or truncated if there are too many.

There is a CGHnormaliter.write.table method that prints the results in a tabular format.

# Value

This function returns a matrix of objects of class cghCall with dimension (number of clones) \* (number of samples). Each object contains the following components (See section Examples on how to access them):

normalized data

A matrix with the normalized log2 intensity ratios for each profile.

segments A matrix with the segments for each profile.

4 CGHnormaliter-internal

calls A data.frame with the calls for each profile. Values are -1 (loss), 0 (normal) or 1 (gain).

probabilities A data.frame with 3 columns of probe information (name, chromosome and

position), followed by 3 columns with aberration probabilities for each sample.

#### Author(s)

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling

#### References

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling, Walter Pirovano and Jaap Heringa. CGH-normaliter: an iterative strategy to enhance normalization of array CGH data with imbalanced aberrations. BMC Genomics, 10:401, 2009.

# **Examples**

```
data(Leukemia)
## Normalize the raw intensity values of the first 4 chromosomes.
result <- CGHnormaliter(Leukemia, nchrom=4)
## Get the normalized log2 intensity ratios, segments and calls
normalized.data <- copynumber(result)
segmented.data <- segmented(result)
called.data <- calls(result)
## Plot the normalization result of sample 2
plot(result[, 2])
## Write the normalized log2 intensity ratios to file
CGHnormaliter.write.table(result)</pre>
```

CGHnormaliter-internal

Internal CGHnormaliter functions

# **Description**

Internal CGHnormaliter functions.

# **Details**

These are not to be called by the user.

CGHnormaliter.write.table 5

```
CGHnormaliter.write.table
```

Print normalized aCGH data to a file

# **Description**

This function stores the results of a CGHnormaliter normalization into a plain text file.

#### Usage

```
CGHnormaliter.write.table(input, data.type=c("normalized", "segmented", "called"), file=paste(dat
```

# **Arguments**

input result object of a CGHnormaliter normalization.

data.type type of aCGH data to store. file file name to store the data in.

#### **Details**

Available aCGH data types are:

- normalized: The normalized log2 intensity ratios
- segmented: The segmented log2 intensity ratios (after normalization)
- called: The calls (after normalization)

The selected data are stored into a tabseparated file with the specified file name.

#### Author(s)

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling

#### References

Bart P.P. van Houte, Thomas W. Binsl, Hannes Hettling, Walter Pirovano and Jaap Heringa. CGH-normaliter: an iterative strategy to enhance normalization of array CGH data with imbalanced aberrations. BMC Genomics, 10:401, 2009.

# See Also

CGHnormaliter

# **Examples**

```
data(Leukemia)
## Normalize the intensity ratios
## Not run: result <- CGHnormaliter(Leukemia)
## Write the result to a file
## Not run: CGHnormaliter.write.table(result)
## Not run: CGHnormaliter.write.table(result, type="segmented")
## Not run: CGHnormaliter.write.table(result, type="called", file="mycalls.txt")</pre>
```

6 Leukemia

Leukemia Array CGH experiment data on childhood acute lymphoblastic leukemia (ALL) in humans

#### **Description**

A dataframe containing 30180 rows and 10 columns, representing the array CGH data of 3 ALL samples.

# Usage

Leukemia

#### **Format**

A dataframe containing the following 10 columns:

**CloneID** The unique identifiers of array elements.

Chromosome Chromosome number of each array element.

Start Chromosomal start position in bp of each array element.

End Chromosomal end position in bp of each array element.

Case1.test Background corrected test intensity values for sample 1.

Case1.ref Background corrected reference intensity values for sample 1.

Case2.test Background corrected test intensity values for sample 2.

Case2.ref Background corrected reference intensity values for sample 2.

Case3.test Background corrected test intensity values for sample 3.

Case3.ref Background corrected reference intensity values for sample 3.

# Source

Provided by the authors (see references).

#### References

Paulsson K, Heidenblad M, Morse H, Borg A, Fioretos T, Johansson B: Identication of cryptic aberrations and characterization of translocation breakpoints using array CGH in high hyperdiploid childhood acute lymphoblastic leukemia. Leukemia 2006, 20:2002-2007.

# **Index**

```
* IO
    CGHnormaliter.write.table, 5
* datasets
    Leukemia, 6
* file
    CGHnormaliter.write.table, 5
* internal
    CGHnormaliter-internal, 4
* iteration
    CGHnormaliter, 3
*\ methods
    CGHnormaliter, 3
* package
    CGHnormaliter-package, 2
.calculateMA (CGHnormaliter-internal), 4
.localLowess (CGHnormaliter-internal), 4
.plotMA (CGHnormaliter-internal), 4
. \verb|readCghRaw| (\verb|CGHnormal| iter-internal), 4
.runCGHcall (CGHnormaliter-internal), 4
CGHnormaliter, 3, 5
CGHnormaliter-internal, 4
CGHnormaliter-package, 2
{\tt CGHnormaliter.write.table,5}
Leukemia, 6
```