

Package ‘autonomics’

November 19, 2025

Type Package

Title Unified Statistical Modeling of Omics Data

Version 1.19.0

Description This package unifies access to Statistical Modeling of Omics Data.

Across linear modeling engines (lm, lme, lmer, limma, and wilcoxon).

Across coding systems (treatment, difference, deviation, etc).

Across model formulae (with/without intercept, random effect, interaction or nesting).

Across omics platforms (microarray, rnaseq, msproteomics, affinity proteomics, metabolomics).

Across projection methods (pca, pls, sma, lda, spls, oplis).

Across clustering methods (hclust, pam, cmeans).

Across survival methods (coxph, survdiff, coin).

It provides a fast enrichment analysis implementation.

License GPL-3

Encoding UTF-8

LazyData true

VignetteBuilder knitr

biocViews Software, DataImport, Preprocessing, DimensionReduction,
PrincipalComponent, Regression, DifferentialExpression,
GeneSetEnrichment, Transcriptomics, Transcription,
GeneExpression, RNASeq, Microarray, Proteomics, Metabolomics,
MassSpectrometry,

BugReports

<https://gitlab.uni-marburg.de/fb20/ag-graumann/software/autonomics/issues>

RoxygenNote 7.3.3

Depends R (>= 4.0)

Imports abind, arrow, BiocFileCache, BiocGenerics, bit64, cluster,
codingMatrices, colorspace, data.table, dplyr, edgeR, ggforce,
ggplot2, ggrepel, graphics, grDevices, grid, gridExtra, limma,
lme4, magrittr, matrixStats, methods, MultiAssayExperiment,
parallel, RColorBrewer, rlang, R.utils, readxl, S4Vectors,
scales, stats, stringi, SummarizedExperiment, survival, tidyr,
tidyselect, tools, utils, vsn

Suggests affy, AnnotationDbi, AnnotationHub, apcluster, Biobase, BiocManager, BiocStyle, Biostrings, coin, diagram, DBI, e1071, ensemblDb, GenomicDataCommons, GenomicRanges, GEOquery, ggstance, ggridges, ggtext, hgu95av2.db, ICSNP, jsonlite, knitr, lmerTest, MASS, mclust, mixOmics, mixtools, mpm, nlme, OlinkAnalyze, org.Hs.eg.db, org.Mm.eg.db, patchwork, pcaMethods, pheatmap, progeny, propagate, RCurl, RSQLite, remotes, rmarkdown, ropis, Rsubread, readODS, rtracklayer, statmod, testthat, UniProt.ws, writexl, XML

git_url <https://git.bioconductor.org/packages/autonomics>

git_branch devel

git_last_commit 9fc1276

git_last_commit_date 2025-10-29

Repository Bioconductor 3.23

Date/Publication 2025-11-19

Author Aditya Bhagwat [aut, cre],
Richard Cotton [aut],
Vanessa Beutgen [ctb],
Witold Szymanski [ctb],
Shahina Hayat [ctb],
Laure Cougnaud [ctb],
Hinrich Goehlmann [sad],
Karsten Suhre [sad],
Johannes Graumann [aut, sad]

Maintainer Aditya Bhagwat <aditya.bhagwat@uni-marburg.de>

Contents

.coxph	7
.densities	8
.extract_p_features	8
.fit_survival	11
.merge	14
.read_compounddiscoverer	15
.read_compounddiscoverer_masslist	15
.read_diann_precursors	16
.read_maxquant_proteingroups	19
.read_metabolon	20
.read_rectangles	22
.read_rnaseq_bams	24
.read_somascan	27
abstract_fit	29
add_adjusted_pvalues	30
add_assay_means	31
add_facetvars	32

add_opentargets_by_uniprot	33
add_psp	34
add_smiles	35
altenrich	35
analysis	36
analyze	37
annotate_compounddiscoverer	39
annotate_maxquant	39
annotate_uniprot_rest	41
assert_is_valid_sumexp	41
AUTONOMICS_DATASETS	42
awblinmod	42
biplot	44
biplot_corrections	45
biplot_covariates	46
block2limma	47
block2lm	48
block2lme	49
block2lmer	50
block_has_two_levels	51
center	51
code	52
collapsed_entrezg_to_symbol	55
COMPOUNDDISCOVERER_PATTERNS	55
contrastdt	56
contrast_coefs	57
contrast_subgroup_cols	58
counts	58
counts2cpm	59
counts2tpm	60
count_in	61
cpm	62
create_design	63
DATADIR	64
defaultmsigfile	66
default_formula	66
default_geom	67
default_sfile	68
demultiplex	68
dequantify	69
dequantify_compounddiscoverer	70
DIMREDUN	71
download_gtf	71
download_mcclain21	72
dt2mat	73
enrichment	73
ens2org	75
entrezg_to_symbol	76

extract_rectangle	76
factorize	78
fcluster	81
fdata	82
fdr2p	84
filter_exprs_replicated_in_some_subgroup	85
filter_features	86
filter_medoid	86
filter_samples	87
fits	88
fix_xlgenes	89
flevels	89
fnames	90
formula2str	91
ftype	91
fvalues	92
fvars	93
genome_to_orgdb	93
group_by_level	94
guess_compounddiscoverer_quantity	95
guess_fitsep	96
guess_maxquant_quantity	96
guess_sep	97
has_multiple_levels	98
hdlproteins	100
impute	100
installed	102
invert_subgroups	103
is_character_matrix	103
is_collapsed_subset	104
is_compounddiscoverer_output	105
is_correlation_matrix	105
is_diann_report	106
is_fastadt	107
is_file	107
is_fraction	108
is_fragpipe_tsv	109
is_imputed	109
is_maxquant_phosphosites	110
is_maxquant_proteingroups	111
is_non_numeric	111
is_positive_number	112
is_scalar_subset	113
is_sig	114
is_valid_formula	114
keep_estimable_features	116
label2index	116
left.vars	117

LINMOD	118
LINMODENGINES	122
list2mat	122
list_files	123
log2counts	123
log2cpm	124
log2diffs	125
log2proteins	126
log2sites	126
log2tpm	127
log2transform	128
logical2factor	129
make_alpha_palette	130
make_colors	131
make_volcano_dt	131
map_fvalues	132
matrix2sumexp	133
MAXQUANT_PATTERNS	133
mclust_breaks	134
mdsplot	135
merge_compounddiscoverer	136
merge_sample_excel	136
merge_sample_file	137
merge_sdata	138
message_df	139
modelvar	140
MSIGCOLLECTIONSHUMAN	146
MSIGDIR	146
nfactors	147
object1	148
OPENTARGETSDIR	148
order_on_p	149
overall_parameters	150
pca	151
pg_to_canonical	153
plot_coef_densities	154
plot_contrastogram	154
plot_contrast_venn	155
plot_data	156
plot_densities	157
plot_densities_transforms	159
plot_design	162
plot_exprs	163
plot_exprs_per_coef	166
plot_fit_summary	167
plot_heatmap	168
plot_matrix	169
plot_sample_nas	170

plot_subgroup_points	171
plot_summary	172
plot_venn	173
plot_venn_heatmap	174
plot_violins	174
plot_volcano	177
plot_x_density	179
PRECURSOR_QUANTITY	181
preprocess_rnaseq_counts	181
pull_columns	183
pvalues_estimable	183
read_affymetrix	185
read_compounddiscoverer	186
read_diann_pgmatrix	187
read_fragpipe	188
read_maxquant_phosphosites	188
read_maxquant_proteingroups	190
read_msigdt	192
read_olink	193
read_salmon	194
read_uniprottdt	194
reexports	195
reset_fit	196
rm_diann_contaminants	196
rm_missing_in_all_samples	197
rm_unmatched_samples	197
sbind	198
scaledlibsizes	199
scoremat	200
slevels	201
snames	201
split_samples	202
stepauc	203
stri_any_regex	203
stri_detect_fixed_in_collapsed	204
subgroup_array	205
subtract_baseline	205
sumexplist_to_longdt	207
sumexp_to_tsv	208
sumexp_to_widedt	208
summarize_fit	209
survobj	210
svalues	211
svars	212
systematic_nas	213
tag_features	213
tag_hdlproteins	214
TAXON_TO_ORGNAME	215

TESTS

tpm

TRANSFORMENGINES

twofactor_sumexp

uncollapse

values

varlevels_dont_clash

venn_detects

weights

write_xl

X

zero_to_na

215

216

217

217

218

218

219

220

221

222

223

224

Index

226

.coxph	<i>Fit onefeature survival</i>
--------	--------------------------------

Description

Fit onefeature survival

Usage

```
.coxph(sd, formula)

.survdiff(sd, formula)

.logrank(sd, formula)
```

Arguments

sd	data.table
formula	model formula

Examples

```
# Dataset
sd <- survobj()
sd %<>% sumexp_to_longdt( svars = c('timetoevent', 'event', 'age', 'sex'), assay = 'exprs2levels')
sd[, value := code(factor(value), 'code_control')]
sd[, age := code(factor(age ), 'code_control')]
sd[, sex := code(factor(sex ), 'code_control')]

# Singlefactor - coxph, survdiff, logrank
.survdiff(sd, survival::Surv(timetoevent, event) ~ value)
.logrank(sd, survival::Surv(timetoevent, event) ~ value)
.coxph(sd, survival::Surv(timetoevent, event) ~ value)
.coxph(sd, survival::Surv(timetoevent, event) ~ age/value)
```

.densities	Densities
------------	-----------

Description

Densities

Usage

```
.densities(x, xpred = x)

densities(x, xpred = x, plot = TRUE, color = "#F8766D")
```

Arguments

x	numeric vector: data points
xpred	numeric vector: prediction points
plot	whether to plot
color	string

Value

numeric vector with same length as xpred

Examples

```
set.seed(1)
x <- c(rnorm(20, 3), rnorm(20,7), rnorm(20, 11))
xpred <- seq(min(x), max(x), length.out = 100)
.densities(x, xpred) # innerfun
densities(x, xpred) # outerfun
```

.extract_p_features	Extract coefficient features
---------------------	------------------------------

Description

Extract coefficient features

Usage

```
.extract_p_features(  
  object,  
  coefs,  
  p = 0.05,  
  fit = fits(object),  
  combiner = "|",  
  features = NULL,  
  verbose = TRUE  
)  
  
.extract_fdr_features(  
  object,  
  coefs,  
  fdr = 0.05,  
  fit = fits(object),  
  combiner = "|",  
  features = NULL,  
  verbose = TRUE  
)  
  
.extract_effectsize_features(  
  object,  
  coefs,  
  effectsize = 1,  
  fit = fits(object),  
  combiner = "|",  
  features = NULL,  
  verbose = TRUE  
)  
  
.extract_n_features(  
  object,  
  coefs,  
  combiner = "|",  
  n,  
  fit = fits(object)[1],  
  features = NULL,  
  verbose = TRUE  
)  
  
extract_contrast_features(  
  object,  
  fit = fits(object)[1],  
  coefs = autonomics::coefs(object, fit = fit),  
  combiner = "|",  
  decreasing = FALSE,  
  p = 1,
```

```

    fdr = 1,
    effectsize = 0,
    sign = c(-1, +1),
    n = 4,
    features = NULL,
    verbose = TRUE
  )

```

Arguments

<code>object</code>	SummarizedXExperiment
<code>coefs</code>	NULL/character: subset of coefs(object)
<code>p</code>	p threshold
<code>fit</code>	character: subset of fits(object)
<code>combiner</code>	' ' or '&': how to combine multiple fits/coefs
<code>features</code>	features to include no matter what (character vector)
<code>verbose</code>	TRUE or FALSE
<code>fdr</code>	fdr threshold
<code>effectsize</code>	effectsize threshold
<code>n</code>	number of top features (Inf means all)
<code>decreasing</code>	TRUE or FALSE
<code>sign</code>	effect sign

Value

SummarizedExperiment

Examples

```

# Read and Fit
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma()
fdr(object) %<>% add_adjusted_pvalues('fdr')

# Single coef
object0 <- object
object %<>% .extract_p_features(      coefs = 't1-t0', p = 0.05)
object %<>% .extract_fdr_features(    coefs = 't1-t0', fdr = 0.05)
object %<>% .extract_effectsize_features(coefs = 't1-t0', effectsize = 1)
object %<>% .extract_n_features(      coefs = 't1-t0', n = 1)
object0 <- object
object %<>% extract_contrast_features(coefs = 't1-t0', p = 0.05, fdr = 0.05, effectsize = 1, sign = -1, n = 1)

# Multiple coefs
object <- object0
object %<>% .extract_p_features(      coefs = c('t1-t0', 't2-t0'), p = 0.05)
object %<>% .extract_fdr_features(    coefs = c('t1-t0', 't2-t0'), fdr = 0.01)
object %<>% .extract_effectsize_features(coefs = c('t1-t0', 't2-t0'), effectsize = 1)

```

```

    object %<>% .extract_n_features(          coefs = c('t1-t0', 't2-t0'), n = 1)
    object <- object0
    object %<>% extract_contrast_features(coefs = c('t1-t0', 't2-t0'), p = 0.05, fdr = 0.01, effectsize = 1, sign = -1)

```

*.fit_survival**Fit/Plot survival***Description**

Fit/Plot survival

Usage

```

.fit_survival(
  object,
  formula = as.formula(sprintf("~%s", assayNames(object)[1])),
  coefs = NULL,
  engine = c("coxph", "survdif", "logrank")[1],
  drop = TRUE,
  coding = "code_control",
  verbose = TRUE
)

fit_survival(
  object,
  formula = as.formula(sprintf("~%s", assayNames(object)[1])),
  engine = c("coxph", "survdif", "logrank")[1],
  drop = TRUE,
  coding = "code_control",
  coefs = NULL,
  verbose = TRUE,
  outdir = NULL,
  plot = FALSE,
  order = coefs(object, fit = engine)[1],
  stats = coefs(object, fit = engine),
  dodge = 0,
  n = if (svar_formula(formula, object)) 1 else min(nrow(object), 2),
  n_col = n %>% min(nrow(object)) %>% sqrt() %>% ceiling() %>% min(4),
  n_row = n %>% min(ncol(object)) %>% sqrt() %>% floor() %>% min(4),
  width = 3 * n_col,
  height = 3 * n_row,
  writefunname = "write_xl"
)

prep_survival(
  object,
  formula = as.formula(sprintf("~%s", assayNames(object)[1])),

```

```

    assaylevels = NULL,
    engine = c("coxph", "survdif", "logrank") %>% intersect(fits(object)) %>%
      extract(1),
    order = autonomics::coefs(object, fit = engine)[1],
    stats = autonomics::coefs(object, fit = engine),
    n = if (svar_formula(formula, object)) 1 else min(nrow(object), 9)
  )

plot_survival(
  object,
  formula = as.formula(sprintf("~%s", assayNames(object)[1])),
  assaylevels = NULL,
  engine = c("coxph", "survdif", "logrank") %>% intersect(fits(object)) %>%
    extract(1),
  order = autonomics::coefs(object, fit = engine)[1],
  stats = autonomics::coefs(object, fit = engine),
  title = sprintf("%s ~ %s", engine, formula2str(formula) %>% substr(2, nchar(.))),
  dodge = 0,
  file = NULL,
  n = if (svar_formula(formula, object)) 1 else min(nrow(object), 4),
  n_col = n %>% min(nrow(object)) %>% sqrt() %>% ceiling() %>% min(4),
  n_row = n %>% min(ncol(object)) %>% sqrt() %>% floor() %>% min(4),
  width = 3 * n_col,
  height = 3 * n_row
)

```

Arguments

<code>object</code>	SummarizedExperiment
<code>formula</code>	model formula: contains svars/assayNames
<code>coefs</code>	NULL or character (coefs to be stored in object)
<code>engine</code>	'coxph', 'survdif' or 'logrank'
<code>drop</code>	TRUE or FALSE : whether to drop var in coefname
<code>coding</code>	string: codingfunname
<code>verbose</code>	TRUE or FALSE
<code>outdir</code>	output directory
<code>plot</code>	TRUE or FALSE
<code>order</code>	NULL/character (coefs to order plots on)
<code>stats</code>	coefs to print stats for
<code>dodge</code>	number
<code>n</code>	number of features to plot
<code>n_col</code>	number of columns
<code>n_row</code>	number of rows
<code>width</code>	number

height	number
writefunname	'write_xl' or 'write_ods'
assaylevels	NULL or vector: assaylevels to be used (for plotting)
title	string
file	filepath

Value

SummarizedExperiment/ggplot

Examples

```
# Formula
# Samplevar-based
  fit_survival(survobj(), ~age)           # age
  fit_survival(survobj(), ~sex)          # sex
  fit_survival(survobj(), ~age + sex)    # age across sexlevels, sex across agelevels
  fit_survival(survobj(), ~age / sex)    # sex within agelevel
  fit_survival(survobj(), ~age * sex)    # sex between agelevels (=age between sexlevels)

# Assayvar-based
  fit_survival(survobj(), ~exprs)        # numerical coding
  fit_survival(survobj(), ~exprs2bins)   # integer coding
  fit_survival(survobj(), ~exprs2levels) # categorical coding

# Samplevar/Assayvar-based
  fit_survival(survobj(), ~age+exprs2levels, order = 'senior-junior' ) # age effect across exprlevels
  fit_survival(survobj(), ~age+exprs2levels, order = '2-1'           ) # expr effect across agelevels
  fit_survival(survobj(), ~age/exprs2levels, order = 'senior:2-1'    ) # expr effect within agelevel
  fit_survival(survobj(), ~age*exprs2levels, order = 'senior-junior:2-1' ) # expr effect differences between agelevels

# Other arguments
# engine: 'coxph' -> 'survdiff'
  fit_survival(survobj(), ~ exprs2levels)           # coxph
  fit_survival(survobj(), ~ exprs2levels, engine = 'survdiff') # survdiff

# drop: drop varname in coefnames -> dont
  fit_survival(survobj(), ~ exprs2levels)           # 2-1
  fit_survival(survobj(), ~ exprs2levels, drop = FALSE) # exprs2levels2-1

# coding: code_control -> contr.treatment
  fit_survival(survobj(), ~ exprs2levels)           # code_control
  fit_survival(survobj(), ~ exprs2levels, coding = 'contr.treatment') # contr.treatment

# outdir: print to object/screen -> print to xlsx/pdf
  fit_survival(survobj(), ~ exprs2levels)           # print to object/screen
  fit_survival(survobj(), ~ exprs2levels, outdir = tempdir()) # print to xlsx/pdf
  fit_survival(survobj(), ~ exprs2levels, outdir = tempdir(), writefunname = 'write_ods') # print to ods/pdf

# plot: plot -> dont
  fit_survival(survobj(), ~ exprs2levels)           # plot
```

```

fit_survival(survobj(), ~ exprs2levels, plot = FALSE) # dont

# order: order on first coef -> order on custom coef
fit_survival(survobj(), ~ age+exprs2levels)           # order on 'senior-junior'
fit_survival(survobj(), ~ age+exprs2levels, order = '2-1') # order on '2-1'

# stats: show stats for all coefs -> show stats for custom coefs
fit_survival(survobj(), ~ age+exprs2levels)           # show stats for 'senior-junior' and 'bin2-bin'
fit_survival(survobj(), ~ age+exprs2levels, stats = 'senior-junior') # show stats for 'senior-junior'

# dodge: overlap curves -> dodge curves
fit_survival(survobj(), ~ age+exprs2levels)           # overlap curves
fit_survival(survobj(), ~ age+exprs2levels, dodge = 2) # dodge curves

# n: (plot) top2 -> top4
fit_survival(survobj(), ~ age+exprs2levels)           # top2
fit_survival(survobj(), ~ age+exprs2levels, n = 4)     # top4

# n_row n_col: 1 row 2 col -> 2 row 1 col
fit_survival(survobj(), ~ age+exprs2levels)           # 1 row 2 col
fit_survival(survobj(), ~ age+exprs2levels, n_row = 2, n_col = 1) # 2 row 1 col

```

.merge

Clean Merge

Description

Clean Merge

Usage

```
.merge(dt1, dt2, by)
```

Arguments

dt1	data.table
dt2	data.table
by	string

Examples

```

require(data.table)
dt1 <- data.table(feature_id = c('PG1', 'PG2'), gene = c('G1', 'G2'))
dt2 <- data.table(feature_id = c('PG1', 'PG2'), protein = c('P1', 'P2'))
dt1 %<>% .merge(dt2, by = 'feature_id')
dt1

```

`.read_compounddiscoverer`*Read compound discoverer files as-is*

Description

Read compound discoverer files as-is

Usage

```
.read_compounddiscoverer(  
  file,  
  quantity = guess_compounddiscoverer_quantity(file),  
  colname_format = NULL,  
  mod_extract = NULL,  
  verbose = TRUE  
)
```

Arguments

<code>file</code>	compound discoverer file
<code>quantity</code>	string
<code>colname_format</code>	function to reformat column names
<code>mod_extract</code>	function to extract MS modi from sample names
<code>verbose</code>	TRUE / FALSE

Value

data.table

`.read_compounddiscoverer_masslist`*Read compound discoverer masslist files as-is*

Description

Read compound discoverer masslist files as-is

Usage

```
.read_compounddiscoverer_masslist(file, verbose = TRUE)
```

Arguments

file	compound discoverer masslist file
verbose	TRUE / FALSE

Value

data.table

.read_diann_precursors

Read diann

Description

Read diann

Usage

```
.read_diann_precursors(  
  file,  
  Global.Q = 0.01,  
  Q = 0.01,  
  Global.PG.Q = 0.01,  
  PG.Q = 0.05,  
  Global.Peptidoform.Q = 0.01,  
  Peptidoform.Q = 0.01,  
  Lib.Q = 0.01,  
  Lib.PG.Q = 0.01,  
  Lib.Peptidoform.Q = 0.01,  
  verbose = TRUE  
)  
  
.read_diann_proteingroups(  
  file,  
  Global.Q = 0.01,  
  Q = 0.01,  
  Global.PG.Q = 0.01,  
  PG.Q = 0.05,  
  Global.Peptidoform.Q = 0.01,  
  Peptidoform.Q = 0.01,  
  Lib.Q = 0.01,  
  Lib.PG.Q = 0.01,  
  Lib.Peptidoform.Q = 0.01,  
  verbose = TRUE  
)
```

```

read_diann_proteingroups(
  file,
  Global.Q = 0.01,
  Q = 0.01,
  Global.PG.Q = 0.01,
  PG.Q = 0.05,
  Global.Peptidoform.Q = 0.01,
  Peptidoform.Q = 0.01,
  Lib.Q = 0.01,
  Lib.PG.Q = 0.01,
  Lib.Peptidoform.Q = 0.01,
  simplify_snames = TRUE,
  rm_contaminants = TRUE,
  impute = FALSE,
  plot = FALSE,
  pca = plot,
  pls = plot,
  fit = if (plot) "limma" else NULL,
  formula = ~subgroup,
  block = NULL,
  coefs = NULL,
  contrasts = NULL,
  palette = NULL,
  verbose = TRUE
)

read_diann(...)

```

Arguments

<code>file</code>	DIA-NN report file (tsv or parquet)
<code>Global.Q</code>	Global.Q cutoff
<code>Q</code>	Q cutoff
<code>Global.PG.Q</code>	Global.PG.Q cutoff
<code>PG.Q</code>	PG.Q cutoff
<code>Global.Peptidoform.Q</code>	Global.Peptidoform.Q cutoff
<code>Peptidoform.Q</code>	Peptidoform.Q cutoff
<code>Lib.Q</code>	Lib.Q cutoff
<code>Lib.PG.Q</code>	Lib.PG.Q cutoff
<code>Lib.Peptidoform.Q</code>	Lib.Peptidoform.Q cutoff
<code>verbose</code>	TRUE or FALSE
<code>simplify_snames</code>	TRUE or FALSE: simplify (drop common parts in) samplenames ?

<code>rm_contaminants</code>	TRUE or FALSE: rm contaminants ?
<code>impute</code>	TRUE or FALSE: impute group-specific NA values ?
<code>plot</code>	TRUE or FALSE
<code>pca</code>	TRUE or FALSE: run pca ?
<code>pls</code>	TRUE or FALSE: run pls ?
<code>fit</code>	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
<code>formula</code>	model formula
<code>block</code>	model blockvar: string or NULL
<code>coefs</code>	model coefficients of interest: character vector or NULL
<code>contrasts</code>	coefficient contrasts of interest: character vector or NULL
<code>palette</code>	color palette: named string vector
<code>...</code>	used to maintain deprecated functions

Details

Defaults for various Q value cutoffs correspond to recommendations by the DIA-NN team for DIA-NN v.2 (as of 03.2025). Of these, the reader of the legacy file format (flat tab separated values, pre-DIA-NN v.2) only utilizes Lib.PG.Q.

Value

data.table or SummarizedExperiment

Examples

```
# Read
file <- download_data('dilution.report.tsv')
.read_diann_precursors(file)      # precursors longdt
.read_diann_proteingroups(file)   # proteingroups longdt
fdt(read_diann_proteingroups(file)) # proteingroups sumexp

# Compare
PR <- .read_diann_precursors(file)
PG <- .read_diann_proteingroups(file)
PG[intensity==top1] # matches      : 24975 (85%) proteingroups
PG[intensity!=top1] # doesnt match : 4531 (15%) proteingroups
RUN <- 'IPT_HeLa_1_DIAstd_Slot1-40_1_9997'
PR[uniprot=='Q96JP5;Q96JP5-2' & run == RUN, 1:6] # match:      8884 ==      8884
PR[uniprot=='P36578' & run == RUN, 1:6] # no match: 650887 != 407978
PR[intensity != top1][feature_id == unique(feature_id)[1]][run == unique(run)[1]][1:2, 1:6]
PR[intensity != top1][feature_id == unique(feature_id)[2]][run == unique(run)[1]][1:2, 1:6]
PR[intensity != top1][feature_id == unique(feature_id)[3]][run == unique(run)[1]][1:3, 1:6]
```

`.read_maxquant_proteingroups`*Read proteingroups/phosphosites as-is*

Description

Read proteingroups/phosphosites as-is

Usage

```
.read_maxquant_proteingroups(  
  file,  
  quantity = guess_maxquant_quantity(file),  
  verbose = TRUE  
)  
  
.read_maxquant_phosphosites(  
  file,  
  profile,  
  quantity = guess_maxquant_quantity(file),  
  verbose = TRUE  
)
```

Arguments

<code>file</code>	proteingroups / phosphosites file
<code>quantity</code>	string
<code>verbose</code>	TRUE / FALSE
<code>profile</code>	proteingroups file

Value

data.table

Examples

```
profile <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')  
fosfile <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics')  
prodt <- .read_maxquant_proteingroups(file = profile)  
fosdt <- .read_maxquant_phosphosites( file = fosfile, profile = profile)
```

<code>.read_metabolon</code>	<i>Read metabolon xlsxfile</i>
------------------------------	--------------------------------

Description

Read metabolon xlsxfile

Usage

```
.read_metabolon(  
  file,  
  sheet = "OrigScale",  
  fidvar = "BIOCHEMICAL",  
  sidvar = "(CLIENT_IDENTIFIER|Client ID)",  
  sfile = NULL,  
  by.x = "sample_id",  
  by.y = NULL,  
  groupvar = NULL,  
  verbose = TRUE  
)
```

```
read_metabolon(  
  file,  
  sheet = "OrigScale",  
  fidvar = "BIOCHEMICAL",  
  sidvar = "(CLIENT_IDENTIFIER|Client ID)",  
  sfile = NULL,  
  by.x = "sample_id",  
  by.y = NULL,  
  groupvar = NULL,  
  fnamevar = "BIOCHEMICAL",  
  kegg_pathways = FALSE,  
  smiles = FALSE,  
  impute = TRUE,  
  plot = FALSE,  
  pca = plot,  
  pls = plot,  
  label = "feature_id",  
  fit = if (plot) "limma" else NULL,  
  formula = as.formula("~ subgroup"),  
  block = NULL,  
  coefs = NULL,  
  contrasts = NULL,  
  palette = NULL,  
  verbose = TRUE  
)
```

Arguments

<code>file</code>	metabolon xlsx file
<code>sheet</code>	excel sheet (number or string)
<code>fidvar</code>	featureid var
<code>sidvar</code>	samplid var
<code>sfile</code>	sample file
<code>by.x</code>	'file' mergeby column
<code>by.y</code>	'sfile' mergeby column
<code>groupvar</code>	string
<code>verbose</code>	TRUE or FALSE
<code>fnamevar</code>	featurename fvar
<code>kegg_pathways</code>	TRUE or FALSE: add kegg pathways?
<code>smiles</code>	TRUE or FALSE: add smiles?
<code>impute</code>	TRUE or FALSE: impute group-specific NA values?
<code>plot</code>	TRUE or FALSE
<code>pca</code>	TRUE or FALSE
<code>pls</code>	TRUE or FALSE
<code>label</code>	fvar
<code>fit</code>	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
<code>formula</code>	model formula
<code>block</code>	model blockvar: string or NULL
<code>coefs</code>	model coefficients of interest: character vector or NULL
<code>contrasts</code>	coefficient contrasts of interest: character vector or NULL
<code>palette</code>	NULL or colorvector

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
read_metabolon(file, plot = TRUE, block = 'Subject')
```

<code>.read_rectangles</code>	<i>Read omics data from rectangular file</i>
-------------------------------	--

Description

Read omics data from rectangular file

Usage

```
.read_rectangles(  
  file,  
  sheet = 1,  
  fid_rows,  
  fid_cols,  
  sid_rows,  
  sid_cols,  
  expr_rows,  
  expr_cols,  
  fvar_rows = NULL,  
  fvar_cols = NULL,  
  svar_rows = NULL,  
  svar_cols = NULL,  
  fdata_rows = NULL,  
  fdata_cols = NULL,  
  sdata_rows = NULL,  
  sdata_cols = NULL,  
  transpose = FALSE,  
  verbose = TRUE  
)
```

```
read_rectangles(  
  file,  
  sheet = 1,  
  fid_rows,  
  fid_cols,  
  sid_rows,  
  sid_cols,  
  expr_rows,  
  expr_cols,  
  fvar_rows = NULL,  
  fvar_cols = NULL,  
  svar_rows = NULL,  
  svar_cols = NULL,  
  fdata_rows = NULL,  
  fdata_cols = NULL,  
  sdata_rows = NULL,  
  sdata_cols = NULL,
```

```

    transpose = FALSE,
    sfile = NULL,
    sfileby = NULL,
    subgroupvar = character(0),
    verbose = TRUE
  )

```

Arguments

<code>file</code>	string: name of text (txt, csv, tsv, adat) or excel (xls, xlsx) file
<code>sheet</code>	integer/string: only relevant for excel files
<code>fid_rows</code>	numeric vector: featureid rows
<code>fid_cols</code>	numeric vector: featureid cols
<code>sid_rows</code>	numeric vector: sampleid rows
<code>sid_cols</code>	numeric vector: sampleid cols
<code>expr_rows</code>	numeric vector: expr rows
<code>expr_cols</code>	numeric vector: expr cols
<code>fvar_rows</code>	numeric vector: fvar rows
<code>fvar_cols</code>	numeric vector: fvar cols
<code>svar_rows</code>	numeric vector: svar rows
<code>svar_cols</code>	numeric vector: svar cols
<code>fdata_rows</code>	numeric vector: fdata rows
<code>fdata_cols</code>	numeric vector: fdata cols
<code>sdata_rows</code>	numeric vector: sdata rows
<code>sdata_cols</code>	numeric vector: sdata cols
<code>transpose</code>	TRUE or FALSE (default)
<code>verbose</code>	TRUE (default) or FALSE
<code>sfile</code>	sample file
<code>sfileby</code>	sample file mergeby column
<code>subgroupvar</code>	subgroupvar in sfile

Value

SummarizedExperiment

Examples

```

# RNASEQ
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
read_rectangles( file, fid_rows = 2:25,    fid_cols = 2,
                  sid_rows = 1,           sid_cols = 5:28,
                  expr_rows = 2:25 ,      expr_cols = 5:28,
                  fvar_rows = 1,          fvar_cols = 1:4,
                  fdata_rows = 2:25 ,     fdata_cols = 1:4,    transpose = FALSE)

```

```

# LCMSMS PROTEINGROUPS
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
read_rectangles( file,
                  fid_rows = 2:21,    fid_cols = 383,
                  sid_rows = 1,       sid_cols = seq(124, 316, by = 6),
                  expr_rows = 2:21,   expr_cols = seq(124, 316, by = 6),
                  fvar_rows = 1,       fvar_cols = c(2, 6, 7, 383),
                  fdata_rows = 2:21,  fdata_cols = c(2, 6, 7, 383),
                  transpose = FALSE )

# SOMASCAN
file <- system.file('extdata/atkin.somascan.adat', package = 'autonomics')
read_rectangles(file, fid_rows = 30,    fid_cols = 23:42,
                  sid_rows = 42:108,    sid_cols = 4,
                  expr_rows = 42:108,   expr_cols = 23:42,
                  fvar_rows = 28:40,     fvar_cols = 22,
                  svar_rows = 41,        svar_cols = 1:21,
                  fdata_rows = 28:40,    fdata_cols = 23:42,
                  sdata_rows = 42:108,   sdata_cols = 1:21, transpose = TRUE)

# METABOLON
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
read_rectangles(file, sheet = 2,
                  fid_rows = 11:30,     fid_cols = 2,
                  sid_rows = 4,         sid_cols = 15:81,
                  expr_rows = 11:30,    expr_cols = 15:81,
                  fvar_rows = 10,       fvar_cols = 1:14,
                  svar_rows = 1:10,     svar_cols = 14,
                  fdata_rows = 11:30,   fdata_cols = 1:14,
                  sdata_rows = 1:10,    sdata_cols = 15:81,
                  transpose = FALSE )

```

<i>.read_rnaseq_bams</i>	<i>Read rnaseq counts/bams</i>
--------------------------	--------------------------------

Description

Read rnaseq counts/bams

Usage

```

.read_rnaseq_bams(
  dir,
  paired,
  genome,
  nthreads = detectCores(),
  sfile = NULL,
  by.y = NULL,
  ensdb = NULL,
  verbose = TRUE
)

```

```
.read_rnaseq_counts(  
  file,  
  fid_col = 1,  
  sfile = NULL,  
  by.y = NULL,  
  ensdb = NULL,  
  verbose = TRUE  
)  
  
read_rnaseq_bams(  
  dir,  
  paired,  
  genome,  
  nthreads = detectCores(),  
  sfile = NULL,  
  by.y = NULL,  
  block = NULL,  
  formula = as.formula("~ subgroup"),  
  min_count = 10,  
  pseudo = 0.5,  
  ensdb = NULL,  
  tpm = FALSE,  
  cpm = TRUE,  
  log2 = TRUE,  
  plot = FALSE,  
  label = "feature_id",  
  pca = plot,  
  pls = plot,  
  fit = if (plot) "limma" else NULL,  
  voom = cpm,  
  coefs = NULL,  
  contrasts = NULL,  
  palette = NULL,  
  verbose = TRUE  
)  
  
read_rnaseq_counts(  
  file,  
  fid_col = 1,  
  sfile = NULL,  
  by.y = NULL,  
  formula = as.formula("~ subgroup"),  
  block = NULL,  
  min_count = 10,  
  pseudo = 0.5,  
  tpm = FALSE,  
  ensdb = NULL,
```

```

cpm = !tpm,
log2 = TRUE,
plot = FALSE,
label = "feature_id",
pca = plot,
pls = plot,
fit = if (plot) "limma" else NULL,
voom = cpm,
coefs = NULL,
contrasts = NULL,
palette = NULL,
verbose = TRUE
)

```

Arguments

<code>dir</code>	<code>read_rnaseq_bams</code> : bam/sam dir
<code>paired</code>	<code>read_rnaseq_bams</code> : TRUE/FALSE : paired end reads ?
<code>genome</code>	<code>read_rnaseq_bams</code> : 'mm10', 'hg38', etc. or GTF file
<code>nthreads</code>	<code>read_rnaseq_bams</code> : nthreads used by Rsubread::featureCounts()
<code>sfile</code>	sample file
<code>by.y</code>	sample file mergeby column
<code>ensdb</code>	EnsDb with genesizes : e.g. AnnotationHub::AnnotationHub[['AH64923']]
<code>verbose</code>	TRUE or FALSE: message?
<code>file</code>	count file
<code>fid_col</code>	featureid column (number or string)
<code>block</code>	model blockvar: string or NULL
<code>formula</code>	model formula
<code>min_count</code>	min feature count required in some samples
<code>pseudo</code>	pseudocount added to prevent -Inf log2 values
<code>tpm</code>	TRUE or FALSE : add tpm to assays (counts / libsize / genelength) ?
<code>cpm</code>	TRUE or FALSE: add cpm to assays (counts / effectivelibsize) ?
<code>log2</code>	TRUE or FALSE: log2 transform ?
<code>plot</code>	TRUE or FALSE: plot?
<code>label</code>	fvar
<code>pca</code>	TRUE or FALSE: perform and plot pca?
<code>pls</code>	TRUE or FALSE: run pls ?
<code>fit</code>	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
<code>voom</code>	model weights to be computed? TRUE/FALSE
<code>coefs</code>	model coefficients of interest: string vector or NULL
<code>contrasts</code>	model coefficient contrasts of interest: string vector or NULL
<code>palette</code>	color palette : named string vector

Value

SummarizedExperiment

Author(s)

Aditya Bhagwat, Shahina Hayat

Examples

```
# read_rnaseq_bams
if (installed('Rsubread')){
  dir <- download_data('billing16.bam.zip')
  object <- read_rnaseq_bams(dir, paired = TRUE, genome = 'hg38')
  object <- read_rnaseq_bams(dir, paired = TRUE, genome = 'hg38', plot = TRUE)
}

# read_rnaseq_counts
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file, fit = 'limma', coefs = 'E15-E00')
object <- read_rnaseq_counts(file, fit = 'limma', coefs = 'E15-E00', voom = FALSE)
object <- read_rnaseq_counts(file, fit = 'limma', coefs = 'E15-E00', voom = FALSE, cpm = FALSE)
object <- read_rnaseq_counts(file, fit = 'limma', coefs = 'E15-E00', voom = FALSE, cpm = FALSE,
                             log2 = FALSE)
object <- read_rnaseq_counts(file, plot = TRUE)

# read_rnaseq_counts(tpm = TRUE)
## Not run:
ah <- AnnotationHub::AnnotationHub()
ensdb <- ah[['AH64923']]
object <- read_rnaseq_counts(file, fit = 'limma', coefs = 'E02-E00', tpm = TRUE, ensdb = ensdb)

## End(Not run)
```

.read_somascan

Read somascan adatfile

Description

Read somascan adatfile

Usage

```
.read_somascan(
  file,
  fidvar = "Target",
  sidvar = "SampleId",
  sfile = NULL,
  by.x = NULL,
  by.y = NULL,
  groupvar = "SampleGroup",
```

```

    verbose = TRUE
  )

read_somascan(
  file,
  fidvar = "Target",
  sidvar = "SampleId",
  sfile = NULL,
  by.x = NULL,
  by.y = NULL,
  groupvar = "SampleGroup",
  fname_var = "EntrezGeneSymbol",
  sample_type = "Sample",
  feature_type = "Protein",
  sample_quality = c("FLAG", "PASS"),
  feature_quality = c("FLAG", "PASS"),
  rm_na_svars = FALSE,
  rm_single_value_svars = FALSE,
  plot = FALSE,
  label = "feature_id",
  pca = plot,
  pls = plot,
  fit = if (plot) "limma" else NULL,
  formula = as.formula(sprintf("~ %s", groupvar)),
  block = NULL,
  coefs = NULL,
  contrasts = NULL,
  palette = NULL,
  verbose = TRUE
)

```

Arguments

<code>file</code>	somascan (adat) file
<code>fidvar</code>	featureid var
<code>sidvar</code>	sampleid var
<code>sfile</code>	sample file
<code>by.x</code>	'file' mergeby column
<code>by.y</code>	'sfile' mergeby column
<code>groupvar</code>	string
<code>verbose</code>	TRUE or FALSE: message?
<code>fname_var</code>	featurename var: string
<code>sample_type</code>	subset of c('Sample', 'QC', 'Buffer', 'Calibrator')
<code>feature_type</code>	subset of c('Protein', 'Hybridization Control Elution', 'Rat Protein')
<code>sample_quality</code>	subset of c('PASS', 'FLAG', 'FAIL')

feature_quality	subset of c('PASS', 'FLAG', 'FAIL')
rm_na_svars	TRUE or FALSE: rm NA svars?
rm_single_value_svars	TRUE or FALSE: rm single value svars?
plot	TRUE or FALSE: plot ?
label	fvar
pca	TRUE or FALSE: run pca?
pls	TRUE or FALSE: run pls?
fit	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
formula	model formula
block	model blockvar
coefs	model coefficients of interest: character vector or NULL
contrasts	coefficient contrasts of interest: character vector or NULL
palette	character vector or NULL

Value

Summarizedexperiment

Examples

```
file <- system.file('extdata/atkin.somascan.adat', package = 'autonomics')
read_somascan(file, plot = TRUE, block = 'Subject')
```

abstract_fit

Abstract model fit

Description

Abstract model fit

Usage

```
abstract_fit(
  object,
  sep = guess_fitsep(fdt(object)),
  fit = fits(object),
  coef = coefs(object, fit = fit),
  significancevar = "p",
  significance = 0.05
)
```

Arguments

object	SummarizedExperiment
sep	string
fit	character vector
coef	character vector
significancevar	'p' or 'fdr'
significance	fraction : pvalue cutoff

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file, fit = 'limma', coef = 't3-t0')
fdt(object)
fdt(abstract_fit(object))
```

add_adjusted_pvalues *Add adjusted pvalues*

Description

Add adjusted pvalues

Usage

```
add_adjusted_pvalues(object, ...)
```

```
## S3 method for class 'data.table'
add_adjusted_pvalues(
  object,
  method = "fdr",
  fit = fits(object),
  coefs = autonomics::coefs(object, fit = fit),
  verbose = TRUE,
  ...
)
```

```
## S3 method for class 'SummarizedExperiment'
add_adjusted_pvalues(
  object,
  method = "fdr",
  fit = fits(object),
```

```

    coefs = autonomics::coefs(object, fit = fit),
    verbose = TRUE,
    ...
)

## S3 method for class '`NULL`'
add_adjusted_pvalues(object, ...)

```

Arguments

object	SummarizedExperiment or (feature) data.table
...	for s3 dispatch
method	'fdr', 'bonferroni', ... (see 'p.adjust.methods')
fit	'limma', 'lm', 'lme', 'lmer'
coefs	coefficient (string)
verbose	TRUE or FALSE

Value

SummarizedExperiment

Examples

```

file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
fdt(object) %<>% extract(, 1:2)
object %<>% linmod_limma()
object %<>% extract(order(fdt(.)$`p~Adult-X30dpt~limma`), )
  fdt(object)
(fdt(object) %<>% add_adjusted_pvalues('fdr'))
(fdt(object) %<>% add_adjusted_pvalues('fdr'))      # smart enough not to add second column
(fdt(object) %>% add_adjusted_pvalues('bonferroni'))

```

add_assay_means	<i>Add assay means</i>
-----------------	------------------------

Description

Add assay means

Usage

```
add_assay_means(object, assay = assayNames(object)[1], bin = TRUE)
```

Arguments

object	SummarizedExperiment or NULL
assay	string
bin	TRUE or FALSE

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
fdt(object) %<>% extract(, 1:2)
fdt(object)
object %<>% add_assay_means(SummarizedExperiment::assayNames(.))
fdt(object)
```

add_facetvars	<i>Add facetvars</i>
---------------	----------------------

Description

Add facetvars

Usage

```
add_facetvars(
  object,
  fit = fits(object)[1],
  coefs = autonomics::coefs(object, fit = fit)
)
```

Arguments

object	SummarizedExperiment
fit	string
coefs	string vector

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file, fit = 'limma')
object %<>% add_adjusted_pvalues()
fdt(object)
fdt(add_facetvars(object))
```

```
add_opentargets_by_uniprot
      Add opentargets annotations
```

Description

Add opentargets annotations

Usage

```
add_opentargets_by_uniprot(
  object,
  cols = c("genesymbol", "genename", "function"),
  verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
cols	character vector
verbose	TRUE or FALSE

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
object %<>% add_opentargets_by_uniprot()
```

`add_psp`*Add psp*

Description

Add PhosphoSitePlus literature counts

Usage

```
add_psp(  
  object,  
  pspfile = file.path(R_user_dir("autonomics", "cache"), "phosphositeplus",  
    "Phosphorylation_site_dataset.gz")  
)
```

Arguments

<code>object</code>	SummarizedExperiment
<code>pspfile</code>	phosphositeplus file

Details

Go to www.phosphosite.org
Register and Login.
Download `Phosphorylation_site_dataset.gz`.
Save into: `file.path(R_user_dir('autonomics','cache'),'phosphositeplus')`

Value

SummarizedExperiment

Examples

```
fosfile <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics')  
profile <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')  
object <- read_maxquant_phosphosites(fosfile = fosfile, profile = profile)  
fdt(object)  
object %<>% add_psp()  
fdt(object)
```

`add_smiles`*Add smiles*

Description

Add smiles

Usage

```
add_smiles(object)
```

Arguments

`object` character/factor vector with pubchem ids

Value

character/factor vector

References

<https://pubchemdocs.ncbi.nlm.nih.gov/pug-rest-tutorial>

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
# add_smiles(object[1:10, ]) # seems down
```

`altenrich`*Alternative Enrichment Analysis*

Description

Alternative Enrichment Analysis

Usage

```
altenrich(
  object,
  pathwaydt,
  genevar = "gene",
  genesep = "[ ,;]",
  coef = autonomics::coefs(object)[1],
  fit = fits(object)[1],
  significancevar = "p",
```

```
significance = 0.05,
effectsize = 0,
n = 3,
genes = FALSE,
verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
pathwaydt	data.table, e.g. read_msigt
genevar	gene fvar
genesep	string or NULL
coef	string in <code>coefs(object)</code>
fit	'limma', 'lm', 'lme', 'lmer', 'wilcoxon'
significancevar	'p' or 'fdr'
significance	significance cutoff
effectsize	effectsize cutoff
n	no of detected genes required (for geneset to be examined)
genes	whether to record genes
verbose	whether to msg

Details

This is an alternative enrichent analysis implementation. It is more modular: uses four times `.enrichment(VERBOSE)?` as backend. But also four times slower than `enrichment`, so not recommended. It is retained for testing purposes.

This alternative enrichment implementation

See Also

`[enrichment()]`

analysis	<i>Get/set analysis</i>
----------	-------------------------

Description

Get/set analysis

Usage

```
analysis(object)

## S4 method for signature 'SummarizedExperiment'
analysis(object)

analysis(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,list'
analysis(object) <- value
```

Arguments

object	SummarizedExperiment
value	list

Value

analysis details (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
analysis(object)
```

analyze	<i>Analyze</i>
---------	----------------

Description

Analyze

Usage

```
analyze(
  object,
  pca = TRUE,
  pls = TRUE,
  fit = "limma",
  formula = ~subgroup,
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  contrasts = NULL,
  coefs = contrast_coefs(object, formula = formula, drop = drop, coding = coding),
  block = NULL,
  weightvar = if ("weights" %in% assayNames(object)) "weights" else NULL,
```

```

plot = pca & !is.null(fit),
label = "feature_id",
palette = NULL,
verbose = TRUE
)

```

Arguments

object	SummarizedExperiment
pca	TRUE / FALSE: perform pca ?
pls	TRUE / FALSE: perform pls ?
fit	linmod engine: 'limma', 'lm', 'lme(r)', 'lmer', 'wilcoxon'
formula	model formula
drop	TRUE / FALSE : drop varname in designmat ?
coding	string: codingfunname • contr.treatment: intercept = y_0, coefi = $y_i - y_0$ • contr.treatment.explicit: intercept = y_0, coefi = $y_i - y_0$ • code_control: intercept = ymean, coefi = $y_i - y_0$ • contr.diff: intercept = y_0, coefi = $y_i - y_{(i-1)}$ • code_diff: intercept = ymean, coefi = $y_i - y_{(i-1)}$ • code_diff_forward: intercept = ymean, coefi = $y_i - y_{(i+)}$ • code_deviation: intercept = ymean, coefi = $y_i - y_{\text{mean}}$ (drop last) • code_deviation_first: intercept = ymean, coefi = $y_i - y_{\text{mean}}$ (drop first) • code_helmert: intercept = ymean, coefi = $y_i - \text{mean}(y_0:(y_{i-1}))$ • code_helmert_forward: intercept = ymean, coefi = $y_i - \text{mean}(y_{(i+1)}:y_p)$
contrasts	model coefficient contrasts of interest: string vector or NULL
coefs	model coefficients of interest: string vector or NULL
block	model blockvar
weightvar	NULL or name of weight matrix in assays(object)
plot	TRUE / FALSE
label	fvar
palette	NULL or colorvector
verbose	TRUE / FALSE: message?

Value

SummarizedExperiment

Examples

```

file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% analyze()

```

annotate_compounddiscoverer
Read compound discoverer output

Description

Read compound discoverer output

Usage

```
annotate_compounddiscoverer(
  x,
  dir = getwd(),
  files = list.files(path = dir, pattern = ".*masslist.*\\.xlsx$", ignore.case = TRUE,
    full.names = TRUE),
  verbose = TRUE
)
```

Arguments

x	SummarizedExperiment (read_compounddiscoverer)
dir	compound discoverer output directory
files	compound discoverer masslist files
verbose	TRUE or FALSE : message ?

Value

SummarizedExperiment

annotate_maxquant *Annotate maxquant*

Description

Annotate maxquant data.table

Usage

```
annotate_maxquant(
  dt,
  uniprothdrs,
  contaminanthdrs,
  maxquanthdrs,
  restapi = FALSE,
  verbose = TRUE
)
```

Arguments

dt	data.table : output of read_maxquant_(proteingroups phosphosites)
uniprothdrs	data.table : output of read_uniprot
contaminanthdrs	data.table : output of read_uniprot
maxquanthdrs	data.table : output of read_uniprot
restapi	logical(1) : use uniprot restapi to complete missing annotations ?
verbose	logical(1) : message ?

Details

Uncollapse, annotate, curate, recollapse, name

Value

data.table

Examples

```
# Fukuda 2020: contaminants + maxquanthdrs
#-----
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
dt <- .read_maxquant_proteingroups(file)
dt[, 1:2]
uniprothdrs <- NULL
contaminanthdrs <- read_contaminantdt()
maxquanthdrs <- parse_maxquant_hdrs(dt$`Fasta headers`); dt$`Fasta headers` <- NULL
dt %<>% annotate_maxquant(uniprothdrs, contaminanthdrs, maxquanthdrs)
dt[, 1:9]
dt[reverse== '+', 1:9]
dt[contaminant== '+', 1:9]

# Billing 2019: uniprothdrs + contaminants + maxquanthdrs
#-----
profile <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
fosfile <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics')
upfile <- system.file('extdata/uniprot_hsa_20140515.fasta', package = 'autonomics')
prodt <- .read_maxquant_proteingroups(profile); prodt[, 1:2]
fosdt <- .read_maxquant_phosphosites(fosfile, profile); fosdt[, 1:3]
uniprothdrs <- read_uniprot(upfile)
contaminanthdrs <- read_contaminantdt()
maxquanthdrs <- parse_maxquant_hdrs(prodt$`Fasta headers`)
annotate_maxquant(prodt, uniprothdrs, contaminanthdrs, maxquanthdrs)[, 1:8]
annotate_maxquant(fosdt, uniprothdrs, contaminanthdrs, maxquanthdrs)[, 1:8]
```

annotate_uniprot_rest *Annotate uniprot/ensp*

Description

Annotate uniprot/ensp

Usage

```
annotate_uniprot_rest(x, columns = UNIPROTCOLS, verbose = TRUE)
```

Arguments

x	character vector
columns	character vector
verbose	TRUE or FALSE

Value

data.table(dbid, uniprot, reviewed, protein, gene, canonical, isoform, fragment, existence, organism, full)

Examples

```
# works, but sometimes fails during check
annotate_uniprot_rest( x = c('P00761', 'Q32MB2') )
annotate_uniprot_rest( x = c('ENSBTAP00000006074', 'ENSP00000377550') )
```

assert_is_valid_sumexp

Assert that x is a valid SummarizedExperiment

Description

Assert that x is a valid SummarizedExperiment

Usage

```
assert_is_valid_sumexp(x, .xname = get_name_in_parent(x))
```

Arguments

x	SummarizedExperiment
.xname	see get_name_in_parent

Value

TRUE or FALSE

Examples

```
# VALID
  file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
  x <- read_metabolon(file)
  assert_is_valid_sumexp(x)
# NOT VALID
  rownames(SummarizedExperiment::colData(x)) <- NULL
  # assert_is_valid_sumexp(x)
```

AUTONOMICS_DATASETS	<i>Data used in examples/vignette/tests/longtests</i>
---------------------	---

Description

Data used in examples/vignette/tests/longtests

Usage

AUTONOMICS_DATASETS

Format

An object of class character of length 19.

Examples

AUTONOMICS_DATASETS

awblinmod	<i>General Linear Modeling (across-within-between interface)</i>
-----------	--

Description

General Linear Modeling (across-within-between interface)

Usage

```

awblinmod(
  object,
  engine,
  modelvars,
  across = TRUE,
  within = if (length(modelvars) == 1) FALSE else TRUE,
  between = if (length(modelvars) == 1) FALSE else TRUE,
  coding = c("code_control", "code_diff"),
  drop = TRUE,
  verbose = TRUE,
  ...
)

awblinmod_limma(object, ...)

awblinmod_lm(object, ...)

awblinmod_lme(object, ...)

awblinmod_lmer(object, ...)

```

Arguments

object	SummarizedExperiment
engine	'limma', 'lm', 'lme', or 'lmer'
modelvars	svars
across	TRUE/FALSE: fit across model (additive) ?
within	TRUE/FALSE: fit within model (nested) ?
between	TRUE/FALSE: fit between model (interaction) ?
coding	character: codingfunname
drop	TRUE or FALSE
verbose	TRUE or FALSE
...	passed to linmod

Examples

```

file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
svars(object)
awblinmod_limma(object, modelvars = c('Diabetes', 'Time'), block = 'Subject')
awblinmod_lme( object, modelvars = c('Diabetes', 'Time'), block = 'Subject')
awblinmod_lmer( object, modelvars = c('Diabetes', 'Time'), block = 'Subject')
awblinmod_lm(  object, modelvars = c('Diabetes', 'Time'))
awblinmod(object, engine = 'limma', modelvars = 'Time')
awblinmod(object, engine = 'limma', modelvars = c('Diabetes', 'Time'))

```

biplot

*Biplot***Description**

Biplot

Usage

```

biplot(
  object,
  method = biplot_methods(object)[1],
  by = biplot_by(object, method)[1],
  dims = biplot_dims(object, method, by)[1:2],
  color = if (method %in% DIMREDSUPER) by else "subgroup",
  labelcolors = FALSE,
  shape = NULL,
  size = NULL,
  alpha = NULL,
  group = NULL,
  linetype = NULL,
  label = NULL,
  feature_label = "feature_id",
  fixed = list(shape = 15, size = 3),
  nx = 0,
  ny = 0,
  colorpalette = make_svar_palette(object, color),
  alphapalette = make_alpha_palette(object, alpha),
  title = paste0(method, "~", by),
  theme = ggplot2::theme(plot.title = element_text(hjust = 0.5), panel.grid =
    element_blank())
)

```

Arguments

object	SummarizedExperiment
method	'pca', 'pls', 'lda', 'spl', 'opls', 'sma'
by	svar
dims	numeric vector: e.g. 1:2
color	svar
labelcolors	TRUE or FALSE
shape	svar
size	svar
alpha	svar

group	svar
linetype	svar
label	svar
feature_label	fvar
fixed	fixed plot aesthetics
nx	number of x features to plot
ny	number of y features to plot
colorpalette	character vector
alphapalette	character vector
title	string
theme	ggplot2::theme output

Value

ggplot object

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% pca(ndim = 4)
object %<>% pls(ndim = 4)
biplot(object)
biplot(object, nx = 1)
biplot(object, dims = 3:4, nx = 1)
biplot(object, method = 'pls')
biplot(object, method = 'pls', dims = 3:4)
biplot(object, method = 'pls', dims = 3:4, group = 'Subject')
```

biplot_corrections	<i>Biplot batch corrections</i>
--------------------	---------------------------------

Description

Biplot batch corrections

Usage

```
biplot_corrections(
  object,
  method = "pca",
  by = "sample_id",
  color = "subgroup",
  covariates = character(0),
  varcols = ceiling(sqrt(1 + length(covariates))),
  plot = TRUE
)
```

Arguments

object	SummarizedExperiment
method	'pca', 'pls', 'lda', or 'sma'
by	svar
color	variable mapped to color (symbol)
covariates	covariates to be batch-corrected
varcols	number of covariate columns
plot	TRUE/FALSE: plot?

Value

grid object

See Also

biplot_covariates

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file, pca = TRUE, plot = FALSE)
biplot_corrections(object, color = 'subgroup', covariates = c('Sex', 'Diabetes', 'Subject', 'Time'))
```

biplot_covariates	<i>Biplot covariates</i>
-------------------	--------------------------

Description

Biplot covariates

Usage

```
biplot_covariates(
  object,
  method = "pca",
  by = "sample_id",
  block = NULL,
  covariates = "subgroup",
  ndim = 6,
  dimcols = 1,
  varcols = length(covariates),
  plot = TRUE
)
```

Arguments

object	SummarizedExperiment
method	'pca', 'pls', 'lda', or 'sma'
by	svar
block	svar
covariates	covariates: mapped to color or batch-corrected
ndim	number of dimensions to plot
dimcols	number of dimension columns
varcols	number of covariate columns
plot	TRUE or FALSE: whether to plot

Value

ggplot object

See Also

biplot_corrections

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file, pca = TRUE)
biplot_covariates(object, covariates = 'subgroup', ndim = 12, dimcols = 3)
biplot_covariates(object, covariates = c('Sex', 'Diabetes', 'Subject', 'Time'))
biplot_covariates(object, covariates = c('Sex', 'Diabetes', 'Subject', 'Time'), ndim = 2)
biplot_covariates(object, covariates = c('subgroup'), dimcols = 3)
```

block2limma	<i>block2limma</i>
-------------	--------------------

Description

block2limma

Usage

```
block2limma(block, ...)

## S3 method for class '`NULL`'
block2limma(block, ...)

## S3 method for class 'character'
block2limma(block, ...)
```

```
## S3 method for class 'list'
block2limma(block, ...)

## S3 method for class 'formula'
block2limma(block, ...)
```

Arguments

block block: charactervector or formula
... required for s3 dispatch

Examples

```
block2limma( block = c(        'subject',        'batch'        ))
block2limma( block = c(`1`= 'subject',        `1`= 'batch'        ))
block2limma( block = list(    subject = ~1,        batch = ~1 ))
block2limma( block =        ~(1|subject)        + (1|batch)    )
```

block2lm	<i>block2lm</i>
----------	-----------------

Description

block2lm

Usage

```
block2lm(block, formula, ...)

## S3 method for class '`NULL`'
block2lm(block, formula, ...)

## S3 method for class 'character'
block2lm(block, formula, ...)

## S3 method for class 'list'
block2lm(block, formula, ...)

## S3 method for class 'formula'
block2lm(block, formula, ...)
```

Arguments

block block: charactervector or formula
formula model formula
... required for s3 dispatch

Examples

```

block2lm( block = NULL,                      formula = ~ subgroup)
block2lm( block = c('subject', 'batch'),    formula = ~ subgroup)
block2lm( block = c(`1`= 'subject', `1`= 'batch'), formula = ~ subgroup)
block2lm( block = ~(1|subject) + (1|batch),  formula = ~ subgroup)
block2lm( block = list(subject = ~1, batch = ~1 ), formula = ~ subgroup)

```

block2lme

block2lme

Description

block2lme

Usage

```

block2lme(block, ...)

## S3 method for class 'list'
block2lme(block, ...)

## S3 method for class 'formula'
block2lme(block, ...)

## S3 method for class 'character'
block2lme(block, ...)

```

Arguments

block	block: character vector or formula
...	required for s3 dispatch

Examples

```

block2lme( block = c(      'subject',      'batch'))
block2lme( block = c(`1`= 'subject', `1`= 'batch'))
block2lme( block =   ~(1|subject) + (1|batch)      )
block2lme( block = list(subject = ~1, batch = ~1 ))

```

block2lmer

block2lmer

Description

block2lmer

Usage

```
block2lmer(block, formula, ...)

## S3 method for class 'formula'
block2lmer(block, formula = NULL, ...)

## S3 method for class 'character'
block2lmer(block, formula = NULL, ...)

## S3 method for class 'list'
block2lmer(block, formula = NULL, ...)
```

Arguments

block	block: character vector or formula
formula	model formula
...	required for s3 dispatch

Examples

```
block2lmer( block = c('subject', 'batch'))
block2lmer( block = c('subject', 'batch'), formula = ~ subgroup)

block2lmer( block = c(`1` = 'subject', `1` = 'batch'))
block2lmer( block = c(`1` = 'subject', `1` = 'batch'), formula = ~ subgroup)

block2lmer( block = ~(1|subject) + (1|batch))
block2lmer( block = ~(1|subject) + (1|batch), formula = ~ subgroup)

block2lmer( block = list(subject = ~1, batch = ~1 ))
block2lmer( block = list(subject = ~1, batch = ~1 ), formula = ~ subgroup)
```

block_has_two_levels	<i>Block has two levels</i>
----------------------	-----------------------------

Description

Block has two levels

Usage

```
block_has_two_levels(block, data)
```

Arguments

block	string
data	data.table

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
data <- sumexp_to_longdt(object, svars = 'Subject')
data %<>% extract(feature_id == feature_id[1])
block_has_two_levels(block = 'Subject', data)
```

center	<i>Center samples</i>
--------	-----------------------

Description

Center samples

Usage

```
center(
  object,
  selector = rep(TRUE, nrow(object)) == TRUE,
  fun = "median",
  verbose = TRUE
)

center_mean(object, ...)

center_median(object, ...)
```

Arguments

object	SummarizedExperiment
selector	logical vector (length = nrow(object))
fun	aggregation function (string)
verbose	TRUE/FALSE
...	parameters handed through to center()

Value

SummarizedExperiment

Examples

```
require(matrixStats)
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
fdt(object)$housekeeping <- FALSE
fdt(object)$housekeeping[order(rowVars(values(object)))[1:5]] <- TRUE
values(object)[, object$subgroup=='Adult'] %<>% magrittr::add(5)
plot_sample_densities(object)
plot_sample_densities(center(object))
plot_sample_densities(center(object, housekeeping))
```

code

Contrast Code Factor

Description

Contrast Code Factor for General Linear Model

Usage

```
code(object, ...)

## S3 method for class 'factor'
code(object, coding, verbose = TRUE, ...)

## S3 method for class 'character'
code(object, coding, verbose = TRUE, ...)

## S3 method for class 'logical'
code(object, coding, verbose = TRUE, ...)

## S3 method for class 'numeric'
code(object, coding, verbose = TRUE, ...)
```

```
## S3 method for class 'data.table'
code(object, coding, vars = names(object), verbose = TRUE, ...)

contr.treatment.explicit(n)

code_control(n)

contr.diff(n)

code_diff(n)

code_diff_forward(n)

code_deviation(n)

code_deviation_first(n)

code_helmert(n)

code_helmert_forward(n)
```

Arguments

object	factor vector
...	used for s3 dispatch
coding	string: codingfunname • <code>contr.treatment</code>: $\text{intercept} = y_0$, $\text{coefi} = y_i - y_0$ • <code>contr.treatment.explicit</code>: $\text{intercept} = y_0$, $\text{coefi} = y_i - y_0$ • <code>code_control</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - y_0$ • <code>contr.diff</code>: $\text{intercept} = y_0$, $\text{coefi} = y_i - y_{(i-1)}$ • <code>code_diff</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - y_{(i-1)}$ • <code>code_diff_forward</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - y_{(i+)}$ • <code>code_deviation</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - y_{\text{mean}}$ (drop last) • <code>code_deviation_first</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - y_{\text{mean}}$ (drop first) • <code>code_helmert</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - \text{mean}(y_0:(y_i-1))$ • <code>code_helmert_forward</code>: $\text{intercept} = y_{\text{mean}}$, $\text{coefi} = y_i - \text{mean}(y_{(i+1):yp})$
verbose	TRUE or FALSE
vars	svars
n	character vector

Details

A General Linear Model contains:

- * An Intercept Coefficient: expressing some form of sample average
- * For each numeric variable: a slope coefficient
- * For each k-leveled factor: (k-1) Contrast Coefficients.

The interpretation of (intercept and contrast) coefficients depends on the contrast coding function used. Several contrast coding functions are available in 'stats' and 'codingMatrices'. But their (function and coefficient) namings are a bit confusing and unsystematic. Instead, the functions below offer an intuitive interface (to the otherwise powerful stats/codingMatrices packages). The names of these functions reflect the contrast coding used (treatment, backward, sum, or helmert contrasts). They also reflect the intercept interpretation (either first factor's first level or grand mean). They all produce intuitive coefficient names (e.g. 't1-t0' rather than just 't1'). They all have unit scaling (a coefficient of 1 means a backward of 1).

Value

(explicitly coded) factor vector

Examples

```
# Coding functions
x <- factor(paste0('t', 0:3))
xlevels <- levels(x)
contr.treatment(      xlevels)
contr.treatment.explicit(xlevels)
contr.diff(           xlevels)
code_control(         xlevels)
code_diff(            xlevels)
code_diff_forward(    xlevels)
code_deviation(       xlevels)
code_deviation_first( xlevels)
code_helmert(         xlevels)
code_helmert_forward( xlevels)

# Code
x %<>% code('contr.treatment')
x %<>% code('contr.treatment.explicit')
x %<>% code('contr.diff')
x %<>% code('code_control')
x %<>% code('code_diff')
x %<>% code('code_diff_forward')
x %<>% code('code_deviation')
x %<>% code('code_deviation_first')
x %<>% code('code_helmert')
x %<>% code('code_helmert_forward')

# Model
file <- system.file('extdata/atkin.metabolon.xls', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma(coding = 'contr.treatment') # default
object %<>% linmod_limma(coding = 'contr.treatment.explicit')
object %<>% linmod_limma(coding = 'contr.diff')
object %<>% linmod_limma(coding = 'code_control')
object %<>% linmod_limma(coding = 'code_diff')
object %<>% linmod_limma(coding = 'code_diff_forward')
object %<>% linmod_limma(coding = 'code_deviation')
object %<>% linmod_limma(coding = 'code_deviation_first')
```

```
object %<>% linmod_limma(coding = 'code_helmert')
object %<>% linmod_limma(coding = 'code_helmert_forward')
```

collapsed_entrezg_to_symbol

Collapsed entrezg to genesymbol

Description

Collapsed entrezg to genesymbol

Usage

```
collapsed_entrezg_to_symbol(x, sep, orgdb)
```

Arguments

x	charactervector
sep	string
orgdb	OrgDb

Value

character vector

Examples

```
if (installed('org.Hs.eg.db')){
  x <- c('7448/3818/727', '5034/9601/64374')
  orgdb <- org.Hs.eg.db::org.Hs.eg.db
  collapsed_entrezg_to_symbol(x, sep = '/', orgdb = orgdb)
}
```

COMPOUNDDISCOVERER_PATTERNS

compound discoverer quantity patterns

Description

compound discoverer quantity patterns

Usage

```
COMPOUNDDISCOVERER_PATTERNS
```

Format

An object of class character of length 2.

Examples

COMPOUNDDISCOVERER_PATTERNS

contrastdt	<i>Get contrastdt</i>
------------	-----------------------

Description

Get contrastdt

Usage

```
contrastdt(  
  object,  
  fitcoef,  
  annocols = fvars(object) %>% extract(!stri_detect_fixed(., "~")),  
  assays = assayNames(object)[0],  
  verbose = TRUE  
)
```

Arguments

object	SummarizedExperiment
fitcoef	e.g. 't2-t1~limma'
annocols	annotation fvars
assays	subset of assayNames(object)
verbose	TRUE or FALSE

Value

data.table

Examples

```
object <- survobj()  
object %<>% linmod_limma(~sex/age)  
contrastdt(object, fitcoef = 'm:senior-junior~limma')  
contrastdt(object[, 1:2], fitcoef = 'm:senior-junior~limma', assays = SummarizedExperiment::assayNames(object)[1])  
contrastdt(object[, 1:2], fitcoef = 'm:senior-junior~limma', assays = SummarizedExperiment::assayNames(object)[1])
```

contrast_coefs	<i>Get model coefs</i>
----------------	------------------------

Description

Get model coefs

Usage

```
contrast_coefs(  
  object,  
  formula = default_formula(object),  
  drop = varlevels_dont_clash(object, all.vars(formula)),  
  coding = "code_control",  
  design = create_design(object, formula = formula, drop = drop, coding = coding, verbose  
    = FALSE)  
)  
  
model_coefs(  
  object,  
  formula = default_formula(object),  
  drop = varlevels_dont_clash(object, all.vars(formula)),  
  coding = "code_control",  
  design = create_design(object, formula = formula, drop = drop, coding = coding, verbose  
    = FALSE)  
)
```

Arguments

object	SummarizedExperiment
formula	formula
drop	TRUE or FALSE
coding	string: codingfunname
design	design matrix

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')  
object <- read_metabolon(file)  
object %<>% linmod_limma()  
  model_coefs(object)  
contrast_coefs(object)
```

contrast_subgroup_cols	<i>Row/Col contrasts</i>
------------------------	--------------------------

Description

Row/Col contrasts

Usage

```
contrast_subgroup_cols(object, subgroupvar)

contrast_subgroup_rows(object, subgroupvar)
```

Arguments

object	SummarizedExperiment
subgroupvar	subgroup svar

Value

matrix

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object$subgroup <- paste0(object$Diabetes, '.', object$Time)
subgroup_matrix(object, subgroupvar = 'subgroup')
contrast_subgroup_cols(object, subgroupvar = 'subgroup')
contrast_subgroup_rows(object, subgroupvar = 'subgroup')
```

counts	<i>Get/Set counts</i>
--------	-----------------------

Description

Get / Set counts matrix

Usage

```

counts(object)

## S4 method for signature 'SummarizedExperiment'
counts(object)

counts(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
counts(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
counts(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,NULL'
counts(object) <- value

```

Arguments

object	SummarizedExperiment
value	count matrix (features x samples)

Value

count matrix (get) or updated object (set)

Examples

```

file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
counts(object)[1:3, 1:3]
counts(object) <- values(object)

```

counts2cpm

Convert between counts and cpm/tpm

Description

Convert between counts and cpm/tpm

Usage

```

counts2cpm(x, libsize = scaledlibsizes(x))

cpm2counts(x, libsize)

```

Arguments

x count/cpm matrix
libsize (scaled) libsize vector

Value

cpm/tpm/count matrix

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
libsize <- scaledlibsizes(counts(object))
tpm <- counts2tpm(counts(object), genesize = 1)
cpm <- counts2cpm(counts(object), libsize)
counts <- cpm2counts(cpm, libsize)
sum(counts(object) - counts)
```

counts2tpm	<i>counts to tpm</i>
------------	----------------------

Description

counts to tpm

Usage

```
counts2tpm(x, genesize)
```

Arguments

x count matrix
genesize genesize vector (kilobase)

Value

tpm matrix

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
counts(object)[1:3, 1:3]
counts2tpm(counts(object), genesize = 1)[1:3, 1:3]
```

count_in	<i>Count/Collapse in/outside intersection</i>
----------	---

Description

Count/Collapse in/outside intersection

Usage

```
count_in(x, ...)

## S3 method for class 'character'
count_in(x, y, ...)

## S3 method for class 'factor'
count_in(x, y, ...)

## S3 method for class 'list'
count_in(x, y, ...)

collapse_in(x, ...)

## S3 method for class 'character'
collapse_in(x, y, sep, ...)

## S3 method for class 'factor'
collapse_in(x, y, sep, ...)

## S3 method for class 'list'
collapse_in(x, y, sep, ...)

count_out(x, ...)

## S3 method for class 'character'
count_out(x, y, ...)

## S3 method for class 'factor'
count_out(x, y, ...)

## S3 method for class 'list'
count_out(x, y, ...)
```

Arguments

x	character OR list
...	used for S3 dispatch

y character
sep string

Value

number OR numeric

Examples

```
# Sets
contrast1 <- c('a', 'b', 'c', 'd')
pathway <- c('c', 'd', 'e', 'f')
contrast2 <- c('e', 'f', 'g', 'h')

# Count outside
count_out(contrast1, pathway)
count_out(list(contrast1 = contrast1, contrast2 = contrast2), pathway)

# Count inside
count_in(contrast1, pathway)
count_in(list(contrast1 = contrast1, contrast2 = contrast2), pathway)

# Collapse inside
collapse_in(contrast1, pathway, sep = ' ')
collapse_in(list(contrast1 = contrast1, contrast2 = contrast2), pathway, sep = ' ')
```

cpm

Get/Set cpm

Description

Get / Set cpm matrix

Usage

```
cpm(object)

## S4 method for signature 'SummarizedExperiment'
cpm(object)

cpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
cpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
cpm(object) <- value
```

Arguments

object	SummarizedExperiment
value	cpm matrix (features x samples)

Value

cpm matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
cpm(object)[1:3, 1:3]
cpm(object) <- values(object)
```

create_design	<i>Create design matrix</i>
---------------	-----------------------------

Description

Create design matrix for statistical analysis

Usage

```
create_design(object, ...)

## S3 method for class 'SummarizedExperiment'
create_design(
  object,
  formula = default_formula(object),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  verbose = TRUE,
  ...
)

## S3 method for class 'data.table'
create_design(
  object,
  formula = default_formula(object),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  verbose = TRUE,
  ...
)
```

Arguments

object	SummarizedExperiment or data.frame
...	required to s3ify
formula	formula with svars
drop	whether to drop predictor names
coding	string: codingfunname • contr.treatment: intercept = y0, coefi = yi - y0 • contr.treatment.explicit: intercept = y0, coefi = yi - y0 • code_control: intercept = ymean, coefi = yi - y0 • contr.diff: intercept = y0, coefi = yi - y(i-1) • code_diff: intercept = ymean, coefi = yi - y(i-1) • code_diff_forward: intercept = ymean, coefi = yi - y(i+) • code_deviation: intercept = ymean, coefi = yi - ymean (drop last) • code_deviation_first: intercept = ymean, coefi = yi - ymean (drop first) • code_helmert: intercept = ymean, coefi = yi - mean(y0:(yi-1)) • code_helmert_forward: intercept = ymean, coefi = yi - mean(y(i+1):yp)
verbose	whether to message

Value

design matrix

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
unique(create_design(object))
unique(create_design(object, ~ Time))
unique(create_design(object, ~ Time, coding = 'code_control'))
unique(create_design(object, ~ Time, coding = 'code_diff'))
unique(create_design(object, ~ Time + Diabetes))
unique(create_design(object, ~ Time / Diabetes))
unique(create_design(object, ~ Time * Diabetes))
```

DATADIR	<i>Download autonomics example data</i>
---------	---

Description

Download autonomics example data

Usage

DATADIR

```
download_data(
  filename = NULL,
  localdir = file.path(DATADIR, split_extract_fixed(filename, ".", 1)),
  verbose = TRUE,
  force = FALSE
)
```

Arguments

filename	file name		
	'atkin.somascan.adat'	Halama, 2018	effects of hypoglycemia
	'atkin.metabolon.xlsx'		
	'billing16.bam.zip'	Billing, 2016	stemcell comparison
	'billing16.rnaccounts.txt'		
	'billing16.somascan.adat'		
	'billing16.proteingroups.txt'		
	'billing19.rnaccounts.txt'	Billing, 2016	stemcell differentiation
	'billing19.proteingroups.txt'		
	'billing19.phosphosites.txt'		
	'ddglucose.proteingroups.txt'	Omics Module	glycolysis inhibitor
	'fukuda20.proteingroups.txt'	Fukuda, 2020	zebrafish development
	'halama18.metabolon.xlsx'	Halama, 2018	glutaminase inhibitor
localdir	local dir to save file to		
verbose	TRUE / FALSE		
force	TRUE / FALSE		

Format

An object of class character of length 1.

Value

local file path

Examples

```
# Show available datasets
download_data()

# atkin 2018 - hypoglycemia - pubmed 30525282
# download_data('atkin.somascan.adat')           # somascan intensities
# download_data('atkin.metabolon.xlsx')           # metabolon intensities

# billing16 - stemcell characterization - pubmed 26857143
```

```
# download_data('billing16.proteingroups.txt')      # proteingroup ratios
# download_data('billing16.somascan.adat')           # somascan intensities
# download_data('billing16.rnaseq.counts.txt')       # rnaseq counts
# download_data('billing16.rnaseq.alignments.zip')   # rnaseq alignments

# billing19 - stemcell differentiation - pubmed 31332097
# download_data('billing19.proteingroups.txt')      # proteingroup ratios
# download_data('billing19.phosphosite.ratios.txt') # phosphosite ratios
# download_data('billing19.rnaseq.counts.txt')       # rnaseq counts

# fukuda20 - heart regeneration - pubmed PXD016235
# download_data('fukuda20.proteingroups.txt')       # proteingroup LFQ

# halama18 - glutaminase inhibition - pubmed 30525282
# download_data('halama18.metabolon.xls')           # metabolon intensities
```

defaultmsigfile	Default msigdb file
-----------------	---------------------

Description

Default msigdb file

Usage

defaultmsigfile()

Value

file

default_formula	Create default formula
-----------------	------------------------

Description

Create default formula

Usage

default_formula(object)

Arguments

object SummarizedExperiment

Value

formula

Examples

```
# Abundances
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
default_formula(object)
file <- download_data('billing16.proteingroups.txt')
object <- read_maxquant_proteingroups(file)
default_formula(object)
```

default_geom	<i>Default geom</i>
--------------	---------------------

Description

Default geom

Usage

```
default_geom(object, x, block = NULL)
```

Arguments

object	SummarizedExperiment
x	svar
block	svar or NULL

Value

character vector

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object$Age <- runif(min = 20, max = 60, n = ncol(object))
svars(object)
default_geom(object, x = 'Age')
default_geom(object, x = c('Age', 'Diabetes'))
default_geom(object, x = c('Age', 'Diabetes'), block = 'Subject')
```

default_sfile	<i>Default sfile</i>
---------------	----------------------

Description

Default sfile

Usage

default_sfile(file)

Arguments

file data file

Value

sample file

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
default_sfile(file)
```

demultiplex	<i>Demultiplex snames</i>
-------------	---------------------------

Description

Demultiplex maxquant samplenames

Usage

demultiplex(x, verbose = FALSE)

Arguments

x character vector
verbose TRUE or FALSE

Details

WT(L).KD(H).R1{H/L} -> KD_WT.R1 WT(1).KD(2).R1{1} -> WT.R1 WT.R1 -> WT.R1

Value

character

Examples

```
# uniplexed / intensity / ratio
  demultiplex(c('KD.R1','OE.R1'))
  demultiplex(c('WT(L).KD(M).OE(H).R1{M}', 'WT(L).KD(M).OE(H).R1{H}'))
  demultiplex(c('WT(L).KD(M).OE(H).R1{M/L}', 'WT(L).KD(M).OE(H).R1{H/L}'))
# run / replicate
  demultiplex(c('WT(L).OE(H).R1{L}', 'WT(L).OE(H).R1{H}')) # run
  demultiplex(c('WT.R1(L).OE.R1(H){L}', 'WT.R1(L).OE.R1(H){H}')) # repl
# label / index
  demultiplex(c('WT(L).OE(H).R1{L}', 'WT(L).OE(H).R1{H}')) # label
  demultiplex(c('WT(1).OE(2).R1{1}', 'WT(1).OE(2).R1{2}')) # index
# with unused channels
  demultiplex('WT(1).KD(2).OE(3).R1{6}')
```

dequantify	<i>Dequantify maxquant snames</i>
------------	-----------------------------------

Description

Drop quantity ('Reporter intensity').
Encode {channel} as suffix.

Usage

```
dequantify(x, quantity = guess_maxquant_quantity(x), verbose = FALSE)
```

Arguments

x	character
quantity	'ratio', 'normalizedratio', 'LFQ intensity', 'intensity', 'labeledintensity' 'reporterintensity', 'correctedreporterintensit
verbose	TRUE or FALSE

Details

Ratio H/L WT(L).KD(H).R1 -> WT(L).KD(H).R1{H/L} LFQ intensity WT.R1 -> WT.R1
Reporter intensity 0 WT(126).KD(127).R1 -> WT(1).KD(2).R1{1}

Value

character

Examples

```
dequantify(c('Ratio H/L WT(L).KD(M).OE(H).R1',          # Ratios
             'Ratio M/L WT(L).KD(M).OE(H).R1'))
dequantify(c('Ratio H/L normalized WT(L).KD(M).OE(H).R1', # Norm. Ratios
             'Ratio M/L normalized WT(L).KD(M).OE(H).R1'))
dequantify(c('LFQ intensity WT.R1',                      # LFQ intensity
             'LFQ intensity KD.R1'))
dequantify(c('Reporter intensity 1 WT(126).KD(127).R1',  # Rep.intensities
             'Reporter intensity 2 WT(126).KD(127).R1'))
```

dequantify_compounddiscoverer
<i>dequantify_compounddiscoverer compound discoverer snames</i>

Description

Drop quantity.

Usage

```
dequantify_compounddiscoverer(  
  x,  
  quantity = guess_compounddiscoverer_quantity(x),  
  verbose = FALSE  
)
```

Arguments

x	character
quantity	'area', 'normalizedarea'
verbose	TRUE or FALSE

Details

Norm. Area: 20230908_F143_HILICNEG.raw (F11) -> 20230908_F143_HILICNEG.raw (F11)
Area: 20230908_F143_HILICNEG.raw (F11) -> 20230908_F143_HILICNEG.raw (F11)

Value

character

Examples

```
dequantify_compounddiscoverer("Norm. Area: 20230908_F143_HILICNEG.raw (F11)") # Norm. Area  
dequantify_compounddiscoverer("Area: 20230908_F143_HILICNEG.raw (F11)")      # Area
```

 DIMREDUN

Dimension Reduction Methods

Description

Dimension Reduction Methods

Usage

DIMREDUN

DIMREDSUPER

DIMREDENGINES

Format

An object of class character of length 2.

An object of class character of length 4.

An object of class character of length 6.

Details

- DIMREDUN: c('pca', 'sma')
- DIMREDSUPER: c('lda', 'pls', 'opls', 'spls')
- DIMREDENGINES: c('pca', 'sma', 'lda', 'pls', 'opls', 'spls')

 download_gtf

Download GTF file

Description

Download GTF file with feature annotations

Usage

```
download_gtf(
  organism,
  release = 100,
  gtffile = sprintf("%s/gtf/%s", R_user_dir("autonomics", "cache"),
    basename(make_gtf_url(organism, release) %>% substr(1, nchar(.) - 3)))
)
```

Arguments

organism	'Homo sapiens', 'Mus musculus' or 'Rattus norvegicus'
release	GTF release (number)
gtffile	string: path to local GTF file

Value

gtffile path

Examples

```
organism <- 'Homo sapiens'
# download_gtf(organism)
```

download_mcclain21	<i>Download mcclain21 data</i>
--------------------	--------------------------------

Description

Download mcclain21 data

Usage

```
download_mcclain21(
  counts_or_samples = "counts",
  localdir = file.path(DATADIR, "mcclain21"),
  force = FALSE
)
```

Arguments

counts_or_samples	'counts' or 'samples'
localdir	dirname
force	TRUE or FALSE

Details

Mc clain 2021: COVID19 transcriptomics:

Examples

```
download_mcclain21('counts')
download_mcclain21('samples')
```

dt2mat	<i>'data.table' to 'matrix'</i>
--------	---------------------------------

Description

Convert between 'data.table' and 'matrix'

Usage

```
dt2mat(x)

mat2dt(x, idvar)
```

Arguments

x data.table / matrix
idvar idvar string

Value

matrix / data.table

Examples

```
x <- data.table::data.table(
  gene   = c('ENSG001', 'ENSG002', 'ENSG003'),
  sampleA = c(1787, 10, 432),
  sampleB = c(1143, 3, 268))
dt2mat(x)
mat2dt(dt2mat(x), 'gene')
```

enrichment	<i>Enrichment analysis</i>
------------	----------------------------

Description

Are selected genes enriched in pathway?

Usage

```
enrichment(
  object,
  pathwaydt,
  fit = fits(object)[1],
  coef = coefs(object, fit = fit)[1],
  var = abstractvar(object, fit = fit, coef = coef),
```

```

    levels = fdt(object)[[var]] %>% base::levels() %>% extract(-1),
    genevar = "gene",
    genesep = "[ ,;]",
    n = 3,
    verbose = TRUE,
    genes = FALSE
  )

```

Arguments

<code>object</code>	SummarizedExperiment
<code>pathwaydt</code>	pathway data.table
<code>fit</code>	string
<code>coef</code>	string
<code>var</code>	selection fvar
<code>levels</code>	selection levels
<code>genevar</code>	gene fvar
<code>genesep</code>	gene separator (string)
<code>n</code>	number
<code>verbose</code>	whether to msg
<code>genes</code>	whether to report genes

Details

Four enrichment analyses per geneset using the Fisher Exact Test (see four pvalues). Results are returned in a data.table

<code>in</code>	: genes in pathway
<code>in.det</code>	: detected genes in pathway
<code>in.sel</code>	: up/downregulated genes in pathway
<code>in.up(.genes)</code>	: upregulated genes in pathway
<code>in.down(.genes)</code>	: downregulated genes in pathway
<code>out</code>	: genes outside pathway
<code>det</code>	: detected genes (in + out)
<code>sel</code>	: up/downregulated genes (in + out)
<code>up</code>	: upregulated genes (in + out)
<code>down</code>	: downregulated genes (in + out)
<code>p.coef.upDET</code>	: prob to randomly select this many (or more) upregulated genes (among detected genes)
<code>p.coef.downDET</code>	: prob to randomly select this many (or more) downregulated genes (among detected genes)
<code>p.coef.selDET</code>	: prob to randomly select this many (or more) up OR downregulated genes (among detected genes)
<code>p.coef.selGEN</code>	: prob to randomly select this many (or more) up OR downregulated genes (among genome genes)
<code>p.detGEN</code>	: prob to randomly select this many (or more) detected genes (among genome genes)

Examples

```
# Read
pathwaydt <- read_msigt(collections = 'C5:GO:BP')
file <- system.file('extdata/atkin.somascan.adat', package = 'autonomics')
object <- read_somascan(file, fit = 'limma', coefs = 't1-t0')
fvars(object) %<>% gsub('EntrezGeneSymbol', 'gene', .)
object %<>% abstract_fit()
varlevels <- c('flat', 'down', 'up')
enrichdt1 <- enrichment(object, pathwaydt, var = 't1-t0~limma') # 2:n factor
enrichdt2 <- enrichment(object, pathwaydt, var = 't1-t0~limma', levels = varlevels) # 1:n factor
enrichdt3 <- altenrich(object, pathwaydt) # alternative implementation
cols <- intersect(names(enrichdt1), names(enrichdt3))
all(enrichdt1[, cols, with = FALSE] == enrichdt3[, cols, with = FALSE]) # identical
```

ens2org	<i>taxon/ens to organism</i>
---------	------------------------------

Description

taxon/ens to organism

Usage

```
ens2org(x)

taxon2org(x)
```

Arguments

x character vector

Value

character vector

Examples

```
taxon2org( x = c('9606', '9913') )
ens2org( x = c('ENSP00000377550', 'ENSBTAP00000038329') )
```

entrezg_to_symbol	<i>Entrezg to genesymbol</i>
-------------------	------------------------------

Description

Entrezg to genesymbol

Usage

```
entrezg_to_symbol(x, orgdb)
```

Arguments

x	charactervector
orgdb	OrgDb

Value

character vector

Examples

```
if (installed('org.Hs.eg.db')){
  orgdb <- org.Hs.eg.db::org.Hs.eg.db
  entrezg_to_symbol(x = c('7448', '3818', '727'), orgdb)
}
```

extract_rectangle	<i>Extract rectangle from omics file, data.table, or matrix</i>
-------------------	---

Description

Extract rectangle from omics file, data.table, or matrix

Usage

```
extract_rectangle(x, ...)

## S3 method for class 'character'
extract_rectangle(
  x,
  rows = seq_len(nrows(x, sheet = sheet)),
  cols = seq_len(ncols(x, sheet = sheet)),
  verbose = FALSE,
  transpose = FALSE,
  drop = FALSE,
```

```

    sheet = 1,
    ...
)

## S3 method for class 'data.table'
extract_rectangle(
  x,
  rows = seq_len(nrow(x)),
  cols = seq_len(ncol(x)),
  transpose = FALSE,
  drop = FALSE,
  ...
)

## S3 method for class 'matrix'
extract_rectangle(
  x,
  rows = seq_len(nrow(x)),
  cols = seq_len(ncol(x)),
  transpose = FALSE,
  drop = FALSE,
  ...
)

```

Arguments

x	omics datafile or datatable
...	allow for S3 method dispatch
rows	numeric vector
cols	numeric vector
verbose	logical
transpose	logical
drop	logical
sheet	numeric or string

Value

matrix

Examples

```

# FROM FILE: extract_rectangle.character
#=====
x <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
extract_rectangle(x, rows = 11:30, cols = 15:81, sheet = 2)[ 1:3, 1:3 ] # exprs
extract_rectangle(x, rows = 11:30, cols = 2, sheet = 2)[ 1:3, ] # fids
extract_rectangle(x, rows = 4, cols = 15:81, sheet = 2)[ , 1:3 ] # sids
extract_rectangle(x, rows = 10:30, cols = 1:14, sheet = 2)[ 1:3, 1:3 ] # fdt

```

```

extract_rectangle(x, rows = 1:10, cols = 14:81, sheet = 2, transpose = TRUE)[1:3, 1:3] # sdt

# FROM MATRIX: extract_rectangle.matrix
#=====
x <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
x %<>% extract_rectangle(sheet = 2)
extract_rectangle(x, rows = 11:30, cols = 15:81, sheet = 2)[ 1:3, 1:3 ] # exprs
extract_rectangle(x, rows = 11:30, cols = 2, sheet = 2)[ 1:3, ] # fids
extract_rectangle(x, rows = 4, cols = 15:81, sheet = 2)[ , 1:3 ] # sids
extract_rectangle(x, rows = 10:30, cols = 1:14, sheet = 2)[ 1:3, 1:3 ] # fdt
extract_rectangle(x, rows = 1:10, cols = 14:81, sheet = 2, transpose = TRUE)[1:3, 1:3] # sdt

```

factorize

Factorize/Bin

Description

Factorize/Bin

Usage

```
factorize(x, ...)
```

```
## S3 method for class 'logical'
factorize(x, ...)
```

```
## S3 method for class 'character'
factorize(x, ...)
```

```
## S3 method for class 'factor'
factorize(x, ...)
```

```
## S3 method for class 'numeric'
factorize(
  x,
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  numericlevels = TRUE,
  ...
)
```

```
## S3 method for class 'matrix'
factorize(
  x,
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  numericlevels = TRUE,
  ...
)
```

```
)

## S3 method for class 'SummarizedExperiment'
factorize(
  x,
  assay = assayNames(x)[1],
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  numericlevels = TRUE,
  drop = TRUE,
  verbose = TRUE,
  ...
)

factorize_assay(
  x,
  assay = assayNames(x)[1],
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  verbose = TRUE,
  ...
)

bin(x, ...)

## S3 method for class 'logical'
bin(x, ...)

## S3 method for class 'character'
bin(x, ...)

## S3 method for class 'factor'
bin(x, ...)

## S3 method for class 'numeric'
bin(
  x,
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  numericlevels = TRUE,
  ...
)

## S3 method for class 'matrix'
bin(
  x,
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
```

```

    numericlevels = TRUE,
    ...
)

## S3 method for class 'SummarizedExperiment'
bin(
  x,
  assay = assayNames(x)[1],
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  verbose = TRUE,
  ...
)

bin_assay(
  x,
  assay = assayNames(x)[1],
  method = "quantile",
  k = switch(method, quantile = 3, mclust = NULL, mixtools = 3),
  verbose = TRUE
)

```

Arguments

<code>x</code>	vector, matrix or SummarizedExperiment
<code>...</code>	(S3 dispatch)
<code>method</code>	'quantile', 'mclust', or 'mixtools'
<code>k</code>	number of bins/levels
<code>numericlevels</code>	TRUE (levels: 1,2, ...) or FALSE (levels: 2.1+, 3.2+, ...)
<code>assay</code>	string
<code>drop</code>	whether to drop assayname in levels ('1','2') or not ('exprs1', 'exprs2') when factorizing
<code>verbose</code>	TRUE or FALSE

Details

'bin' transform into numeric bins : `c(1,2,3,4,5,6) -> c( 1, 1, 2, 2, 3, 3 )` 'factorize' transform into factor levels: `c(1,2,3,4,5,6) -> c('1','1','2','2','3','3')`

Value

vector, matrix or SummarizedExperiment

Examples

```

# data
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')

```

```

object <- read_maxquant_proteingroups(file, impute = TRUE)
fdt(object)

# logical
fdt(object)$imputed
fdt(object)$imputed %>% factorize()
fdt(object)$imputed %>% bin()

# character
as.character(fdt(object)$imputed)
as.character(fdt(object)$imputed) %>% factorize()
as.character(fdt(object)$imputed) %>% bin()

# factor
factor(fdt(object)$imputed)
factor(fdt(object)$imputed) %>% factorize()
factor(fdt(object)$imputed) %>% bin()

# numeric
fdt(object)$pepcounts
fdt(object)$pepcounts %>% factorize()
fdt(object)$pepcounts %>% bin()

# Matrix/SummarizedExperiment
values(object)
values(object) %>% factorize()
object %>% factorize()
values(object) %>% bin()
object %>% bin()

```

fcluster

*Cluster features***Description**

Cluster features

Usage

```

fcluster(
  object,
  distmat = NULL,
  method = "cmeans",
  k = 2:10,
  verbose = TRUE,
  plot = TRUE,
  label = if ("gene" %in% fvars(object)) "gene" else "feature_id",
  alpha = 1,
  nrow = if (length(method) > 1) length(method) else NULL,
  ncol = NULL
)

```

Arguments

object	SummarizedExperiment
distmat	distance matrix
method	'cmeans'
k	number of clusters
verbose	TRUE or FALSE
plot	TRUE or FALSE
label	fvar
alpha	fraction
nrow	number
ncol	number

Value

SummarizedExperiment
SummarizedExperiment

Examples

```
object <- twofactor_sumexp()
distmat <- fdist(object)
fcluster(object) # membership-based colors
fcluster(object, distmat) # silhouette-based colors
fcluster(object, distmat, method = c('cmeans', 'hclust', 'pamk')) # more methods
```

fdata	<i>Get/Set sample/feature data</i>
-------	------------------------------------

Description

Get/Set sample/feature data

Usage

```
fdata(object)

sdata(object)

fdt(object)

sdt(object)

## S4 method for signature 'SummarizedExperiment'
fdata(object)
```

```
## S4 method for signature 'SummarizedExperiment'
sdata(object)

## S4 method for signature 'SummarizedExperiment'
fdt(object)

## S4 method for signature 'SummarizedExperiment'
sdt(object)

fdata(object) <- value

sdata(object) <- value

fdt(object) <- value

sdt(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,data.frame'
fdata(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,data.frame'
sdata(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,DataFrame'
sdata(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,data.table'
fdt(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,data.table'
sdt(object) <- value
```

Arguments

object	SummarizedExperiment
value	data.frame/data.table

Value

data.frame/data.table (get) or updated object (set)

Examples

```
# Read data
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
# sdt/fdt
sdt(object)[1:3, ]
```

```

    fdt(object)[1:3, ]
    sdt(object) %<>% cbind(b=1)
    fdt(object) %<>% cbind(b=1)
    sdt(object)
    fdt(object)
# sdata/fdata
    sdata(object)[1:3, ]
    fdata(object)[1:3, ]
    sdata(object) %<>% cbind(a=1)
    fdata(object) %<>% cbind(a=1)
    sdata(object)[1:3, ]
    fdata(object)[1:3, ]

```

fdr2p

fdr to p

Description

fdr to p

Usage

```
fdr2p(fdr)
```

Arguments

fdr fdr values

Examples

```

# Read/Fit
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma()
pcol <- pvar(fdt(object), fit = 'limma', coef = 't3-t0')
object %<>% extract(order(fdt(.)[[pcol]]), )
object %<>% extract(1:10, )
fdt(object) %<>% extract(, 1)
object %<>% linmod_limma()
# fdr2p
fdt(object)[[pcol]]
fdt(object)[[pcol]] %>% p.adjust(method = 'fdr')
fdt(object)[[pcol]] %>% p.adjust(method = 'fdr') %>% fdr2p()

```

filter_exprs_replicated_in_some_subgroup

Filter features with replicated expression in some subgroup

Description

Filter features with replicated expression in some subgroup

Usage

```
filter_exprs_replicated_in_some_subgroup(
  object,
  subgroupvar = "subgroup",
  assay = assayNames(object)[1],
  comparator = if (contains_ratios(object)) "!=" else ">",
  lod = 0,
  nsample = 2,
  nsubgroup = 1,
  verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
subgroupvar	subgroup svar
assay	string
comparator	'>' or '!='
lod	number: limit of detection
nsample	number
nsubgroup	number
verbose	TRUE or FALSE

Value

Filtered SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% filter_exprs_replicated_in_some_subgroup()
filter_exprs_replicated_in_some_subgroup(object, character(0))
filter_exprs_replicated_in_some_subgroup(object, NULL)
```

filter_features	<i>Filter features on condition</i>
-----------------	-------------------------------------

Description

Filter features on condition

Usage

```
filter_features(object, condition, verbose = TRUE)
```

Arguments

object	SummarizedExperiment
condition	filter condition
verbose	logical

Value

filtered eSet

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
filter_features(object, SUPER_PATHWAY == 'Lipid')
```

filter_medoid	<i>Filter medoid sample</i>
---------------	-----------------------------

Description

Filter medoid sample

Usage

```
filter_medoid(object, by = NULL, verbose = FALSE)
```

Arguments

object	SummarizedExperiment
by	svar
verbose	whether to message

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file, plot = FALSE)
object %<>% filter_medoid(by = 'subgroup', verbose=TRUE)
```

filter_samples	<i>Filter samples on condition</i>
----------------	------------------------------------

Description

Filter samples on condition

Usage

```
filter_samples(object, condition, verbose = TRUE, record = TRUE, drop = TRUE)
```

Arguments

object	SummarizedExperiment
condition	filter condition
verbose	TRUE/FALSE
record	TRUE/FALSE
drop	TRUE/FALSE : whether to drop levels

Value

filtered SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
filter_samples(object, subgroup != 't0', verbose = TRUE)
```

fits

*Get fit models***Description**

Get fit models

Usage

```
fits(object, ...)

## S3 method for class 'data.table'
fits(object, ...)

## S3 method for class 'SummarizedExperiment'
fits(object, ...)

## S3 method for class '`NULL`'
fits(object, ...)

coefs(object, ...)

## S3 method for class 'factor'
coefs(object, intercept = FALSE, ...)

## S3 method for class 'data.table'
coefs(object, fit = fits(object), intercept = FALSE, ...)

## S3 method for class 'SummarizedExperiment'
coefs(object, fit = fits(object), intercept = FALSE, ...)

## S3 method for class '`NULL`'
coefs(object, ...)

fitcoefs(object)
```

Arguments

object	SummarizedExperiment or data.table
...	S3 dispatch
intercept	TRUE or FALSE : whether to include the intercept
fit	'limma', 'lm', 'lme', 'lmer', 'wilcoxon'

Value

character vector

Examples

```
object <- survobj()
object %<>% linmod_limma(~sex+age)
fits(object)
coefs(object) # sumexp
coefs(fdt(object)) # data.table
coefs(code(factor(object$age), 'code_control')) # factor
fitcoefs(object)
```

fix_xlgenes	<i>Fix excel genes</i>
-------------	------------------------

Description

Fix excel genes

Usage

```
fix_xlgenes(x)
```

Arguments

x character

Value

character

Examples

```
x <- c('FAM46B', '15-Sep', '2-Mar', 'MARCHF6')
x
fix_xlgenes(x)
```

flevels	<i>Get fvar levels</i>
---------	------------------------

Description

Get fvar levels

Usage

```
flevels(object, fvar)
```

Arguments

object	SummarizedExperiment
fvar	feature variable

Value

fvar values

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
head(flevels(object, 'feature_id'))
```

fnames

Get/Set fnames

Description

Get/Set feature names

Usage

```
fnames(object)

## S4 method for signature 'SummarizedExperiment'
fnames(object)

fnames(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,character'
fnames(object) <- value
```

Arguments

object	SummarizedExperiment, eSet, or EList
value	character vector with feature names

Value

feature name vector (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
fnames(object) %<>% paste0('protein_', .)
object
```

formula2str	<i>formula to string</i>
-------------	--------------------------

Description

formula to string

Usage

```
formula2str(formula)
```

Arguments

formula formula

Value

string

Examples

```
formula2str(~0+subgroup)
```

ftype	<i>Feature type</i>
-------	---------------------

Description

Feature type

Usage

```
ftype(  
  object,  
  formula = default_formula(object),  
  drop = varlevels_dont_clash(object, all.vars(formula)),  
  fit = fits(object)[1],  
  coding = "code_control"  
)
```

Arguments

object	SummarizedExperiment
formula	model formula
drop	TRUE or FALSE
fit	'limma', 'lm', 'lme', 'wilcoxon'
coding	coding function

Value

SummarizedExperiment

Examples

```
file <- download_data('atkin.metabolon.xlsx')
object <- read_metabolon(file)
object %<>% linmod_limma(block = 'Subject', coefs = model_coefs(object)) # model_coefs !
object %<>% ftype() # model_coefs not contrast_coefs !
fdt(object) # because intercept is required to recreate predictions
```

fvalues	<i>Get fvalues</i>
---------	--------------------

Description

Get fvar values

Usage

```
fvalues(object, fvar)
```

Arguments

object	SummarizedExperiment
fvar	feature variable

Value

fvar values

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
head(fvalues(object, 'feature_id'))
fvalues(object, NULL)
```

fvars	<i>Get/Set fvars</i>
-------	----------------------

Description

Get/Set feature variables

Usage

```
fvars(object)

## S4 method for signature 'SummarizedExperiment'
fvars(object)

fvars(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,character'
fvars(object) <- value
```

Arguments

object	SummarizedExperiment
value	character vector with feature variables

Value

feature variables vector (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
fvars(object)[1] %<>% paste0('1')
fvars(object)[1]
```

genome_to_orgdb	<i>Get corresponding orgdb</i>
-----------------	--------------------------------

Description

Get corresponding orgdb

Usage

```
genome_to_orgdb(genome)
```

Arguments

genome 'hg38', 'hg19', 'mm10', or 'mm9'

Value

OrgDb

Examples

```
if (requireNamespace('org.Hs.eg.db', quiet = TRUE)){
  class(genome_to_orgdb('hg38'))
}
```

group_by_level	<i>group by level</i>
----------------	-----------------------

Description

group by level

Usage

```
group_by_level(x, ...)

## S3 method for class 'character'
group_by_level(x, ...)

## S3 method for class 'factor'
group_by_level(x, ...)

## S3 method for class 'data.table'
group_by_level(x, var, idvar, ...)
```

Arguments

x named logical/character/factor

... S3 dispatch

var string

idvar string

Value

unnamed character

Examples

```
t1 <- c( KLF5 = 'up', F11 = 'up', RIG = 'flat', ABT1 = 'down')
dt <- data.table( gene = c( 'KL5', 'F11', 'RIG', 'ABT1' ),
                  t1 = c( 'up', 'up', 'flat', 'down' ) )
group_by_level(t1)           # character
group_by_level(factor(t1))   # factor
group_by_level(dt, 't1', 'gene') # data.table
```

```
guess_compounddiscoverer_quantity
```

Guess compound discoverer quantity from snames

Description

Guess compound discoverer quantity from snames

Usage

```
guess_compounddiscoverer_quantity(x)
```

Arguments

x character vector

Value

string: value from names(COMPOUNDDISCOVERER_PATTERNS)

Examples

```
## Not run:
# file
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
guess_compounddiscoverer_quantity(file)

## End(Not run)

# character vector
x <- "Area: 20230908_F143_HILICNEG.raw (F11)"
guess_compounddiscoverer_quantity(x)

x <- "Norm. Area: 20230908_F143_HILICNEG.raw (F11)"
guess_compounddiscoverer_quantity(x)
```

guess_fitsep	<i>guess_fitsep</i>
--------------	---------------------

Description

guess_fitsep

Usage

```
guess_fitsep(object, ...)

## S3 method for class 'data.table'
guess_fitsep(object, ...)

## S3 method for class 'SummarizedExperiment'
guess_fitsep(object, ...)
```

Arguments

object	data.table or SummarizedExperiment
...	S3 dispatch

Value

string

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
object %<>% linmod_limma()
guess_fitsep(object)
```

guess_maxquant_quantity	<i>Guess maxquant quantity from snames</i>
-------------------------	--

Description

Guess maxquant quantity from snames

Usage

```
guess_maxquant_quantity(x)
```

Arguments

x character vector

Value

string: value from names(MAXQUANT_PATTERNS)

Examples

```
# file
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
guess_maxquant_quantity(file)

# character vector
x <- "Ratio M/L normalized STD(L)_E00(M)_E01(H)_R1"
guess_maxquant_quantity(x)

x <- "Ratio M/L STD(L)_E00(M)_E01(H)_R1"
guess_maxquant_quantity(x)

x <- "LFQ intensity E00.R1"
guess_maxquant_quantity(x)

x <- "Reporter intensity corrected 0 STD(0)E00(1)E01(2)_R1"
guess_maxquant_quantity(x)

x <- "Reporter intensity 0 STD(0)E00(1)E01(2)_R1"
guess_maxquant_quantity(x)

x <- "Intensity H STD(L)_E00(M)_E01(H)_R1"
guess_maxquant_quantity(x)
```

guess_sep

Guess separator

Description

Guess separator

Usage

```
guess_sep(x, ...)
```

S3 method for class 'numeric'

```
guess_sep(x, ...)
```

S3 method for class 'character'

```
guess_sep(x, separators = c(".", "_"), verbose = FALSE, ...)
```

```
## S3 method for class 'factor'
guess_sep(x, ...)

## S3 method for class 'SummarizedExperiment'
guess_sep(x, var = "sample_id", separators = c(".", "_"), verbose = FALSE, ...)
```

Arguments

x	character vector or SummarizedExperiment
...	used for proper S3 method dispatch
separators	character vector: possible separators to look for
verbose	TRUE or FALSE
var	svar or fvar

Value

separator (string) or NULL (if no separator could be identified)

Examples

```
# character vector
guess_sep(c('PERM_NON.R1[H/L]', 'PERM_NON.R2[H/L]'))
guess_sep(c('WT_untreated_1', 'WT_untreated_2'))
guess_sep(c('group1', 'group2.R1'))
# SummarizedExperiment
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
guess_sep(object)
```

has_multiple_levels	<i>Variable has multiple levels?</i>
---------------------	--------------------------------------

Description

Variable has multiple levels?

Usage

```
has_multiple_levels(x, ...)

## S3 method for class 'character'
has_multiple_levels(x, .xname = get_name_in_parent(x), ...)

## S3 method for class 'factor'
has_multiple_levels(x, .xname = get_name_in_parent(x), ...)
```

```
## S3 method for class 'numeric'
has_multiple_levels(x, .xname = get_name_in_parent(x), ...)

## S3 method for class 'data.table'
has_multiple_levels(
  x,
  y,
  .xname = get_name_in_parent(x),
  .yname = get_name_in_parent(y),
  ...
)

## S3 method for class 'SummarizedExperiment'
has_multiple_levels(
  x,
  y,
  .xname = get_name_in_parent(x),
  .yname = get_name_in_parent(y),
  ...
)
```

Arguments

x	vector, data.table or SummarizedExperiment
...	required for s3 dispatch
.xname	string
y	string
.yname	string

Value

TRUE or false

Examples

```
# numeric
a <- numeric(); has_multiple_levels(a)
a <- c(1, 1); has_multiple_levels(a)
a <- c(1, 2); has_multiple_levels(a)

# character
a <- character(); has_multiple_levels(a)
a <- c('A', 'A'); has_multiple_levels(a)
a <- c('A', 'B'); has_multiple_levels(a)

# factor
a <- factor(); has_multiple_levels(a)
a <- factor(c('A', 'A')); has_multiple_levels(a)
a <- factor(c('A', 'B')); has_multiple_levels(a)

# data.table
dt <- data.table(a = factor()); has_multiple_levels(dt, 'b')
```

```
dt <- data.table(a = factor());          has_multiple_levels(dt, 'a')
dt <- data.table(a = factor());          has_multiple_levels(dt, 'a')
dt <- data.table(a = factor(c('A', 'A'))); has_multiple_levels(dt, 'a')
dt <- data.table(a = factor(c('A', 'B'))); has_multiple_levels(dt, 'a')
# sumexp
object <- matrix(1:9, nrow = 3)
rownames(object) <- sprintf('%d', 1:3)
colnames(object) <- sprintf('%d', 1:3)
object <- list(exprs = object)
object %<>% SummarizedExperiment::SummarizedExperiment()
object$subgroup <- c('A', 'A', 'A');      has_multiple_levels(object, 'group')
object$subgroup <- c('A', 'A', 'A');      has_multiple_levels(object, 'subgroup')
object$subgroup <- c('A', 'B', 'A');      has_multiple_levels(object, 'subgroup')
```

hdlproteins	<i>hdl proteomewatch proteins</i>
-------------	-----------------------------------

Description

hdl proteomewatch proteins

Usage

hdlproteins()

Value

string vector: HDLProteomeWatch protein entries

Examples

hdlproteins()

impute	<i>Impute</i>
--------	---------------

Description

Impute NA values

Usage

```

impute(object, ...)

## S3 method for class 'numeric'
impute(object, shift = 2.5, width = 0.3, verbose = TRUE, plot = FALSE, ...)

## S3 method for class 'matrix'
impute(
  object,
  shift = 2.5,
  width = 0.3,
  verbose = TRUE,
  plot = FALSE,
  n = min(9, ncol(object)),
  palette = make_colors(colnames(object)),
  ...
)

## S3 method for class 'SummarizedExperiment'
impute(
  object,
  assay = assayNames(object)[1],
  by = "subgroup",
  shift = 2.5,
  width = 0.3,
  frac = 0.5,
  verbose = TRUE,
  plot = FALSE,
  palette = make_colors(colnames(object)),
  n = min(9, ncol(object)),
  ...
)

```

Arguments

<code>object</code>	numeric vector, SumExp
<code>...</code>	required for s3 dispatch
<code>shift</code>	number: sd units
<code>width</code>	number: sd units
<code>verbose</code>	TRUE or FALSE
<code>plot</code>	TRUE or FALSE
<code>n</code>	number of samples to plot
<code>palette</code>	color vector
<code>assay</code>	string
<code>by</code>	svar

frac fraction: fraction of available samples should be greater than this value for a subgroup to be called available

Details

Imputes NA values from N(mean - 2.5 sd, 0.3 sd)

Value

numeric vector, matrix or SumExp

Examples

```
# Simple Design
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
impute(values(object)[, 1], plot = TRUE)[1:3]                    # vector
impute(values(object),        plot = TRUE)[1:3, 1:3]            # matrix
impute(object, plot = TRUE)                                      # sumexp

# Complex Design
subgroups <- sprintf('%s_STD', c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00'))
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, subgroups = subgroups)
impute(values(object)[1:3, 1    ])                              # vector
impute(values(object)[1:3, 1:5 ])                              # matrix
impute( object )                                                  # sumexp
```

installed	<i>Is package installed?</i>
-----------	------------------------------

Description

Is package installed?

Usage

installed(pkg)

Arguments

pkg package (string)

Value

TRUE or FALSE

invert_subgroups	<i>Invert subgroups</i>
------------------	-------------------------

Description

Invert expressions , subgroups, and sample ids

Usage

```
invert_subgroups(  
  object,  
  subgroups = slevels(object, "subgroup"),  
  sep = guess_sep(object, "subgroup")  
)
```

Arguments

object	SummarizedExperiment
subgroups	character vector: subgroup levels to be inversed
sep	string: collapsed string separator

Value

character vector or SummarizedExperiment

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')  
object <- read_maxquant_proteingroups(file)  
invert_subgroups(object)
```

is_character_matrix	<i>Is character matrix</i>
---------------------	----------------------------

Description

Is character matrix

Usage

```
is_character_matrix(x, .xname = get_name_in_parent(x))  
  
assert_character_matrix(x, .xname = get_name_in_parent(x))
```

Arguments

x	matrix
.xname	string

Value

TRUE or false

Examples

```
object <- survobj()
is_character_matrix(SummarizedExperiment::assays(object)$exprs)
is_character_matrix(SummarizedExperiment::assays(object)$exprs2bins)
is_character_matrix(SummarizedExperiment::assays(object)$exprs2levels)
```

is_collapsed_subset	<i>Is collapsed subset</i>
---------------------	----------------------------

Description

Is collapsed subset

Usage

```
is_collapsed_subset(x, y, sep = ";")
```

Arguments

x	character vector
y	character vector
sep	string

Value

character vector

Examples

```
x <- c('H3BNX8;H3BRM5', 'G5E9Y3')
y <- c('P20674;H3BNX8;H3BV69;H3BRM5', 'G5E9Y3;Q8WWN8;B4DIT1')
is_collapsed_subset(x, y)
```

`is_compounddiscoverer_output`*Is compounddiscoverer output?*

Description

Is compounddiscoverer output?

Usage

```
is_compounddiscoverer_output(x, .xname = get_name_in_parent(x))
```

Arguments

<code>x</code>	file
<code>.xname</code>	name of x

Examples

```
file <- NULL;                                     is_compounddiscoverer_output(file)
file <- 3;                                         is_compounddiscoverer_output(file)
file <- 'blabla.tsv';                             is_compounddiscoverer_output(file)
file <- download_data('dilution.report.tsv');    is_compounddiscoverer_output(file)
file <- download_data('multiorganism.combined_protein.tsv'); is_compounddiscoverer_output(file)
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics'); is_compounddiscoverer_output(file)
file <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics'); is_compounddiscoverer_output(file)
```

`is_correlation_matrix` *Assert correlation matrix*

Description

Assert correlation matrix

Usage

```
is_correlation_matrix(
  x,
  .xname = get_name_in_parent(x),
  severity = getOption("assertive.severity", "stop")
)

assert_correlation_matrix(x, .xname = get_name_in_parent(x))
```

Arguments

x	correlation matrix
.xname	string
severity	'warning' or 'stop'

Value

TRUE or false

Examples

```
x <- matrix(c(1,0.7, 0.3, 1), nrow = 2)
rownames(x) <- c('gene1', 'gene2')
colnames(x) <- c('gene1', 'gene2')
is_correlation_matrix(x)
is_correlation_matrix({x[1,1] <- -2; x})
```

is_diann_report	<i>Is diann report ?</i>
-----------------	--------------------------

Description

Is diann report ?

Usage

```
is_diann_report(x, .xname = get_name_in_parent(x))
assert_diann_report(x, .xname = get_name_in_parent(x))
assert_fragpipe_tsv(x, .xname = get_name_in_parent(x))
assert_maxquant_proteingroups(x, .xname = get_name_in_parent(x))
assert_maxquant_phosphosites(x, .xname = get_name_in_parent(x))
assert_compounddiscoverer_output(x, .xname = get_name_in_parent(x))
```

Arguments

x	file
.xname	name of x

Examples

```
file <- NULL;                                     is_diann_report(file)
file <- 3;                                       is_diann_report(file)
file <- 'blabla.tsv';                           is_diann_report(file)
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics'); is_diann_report(file)
file <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics'); is_diann_report(file)
file <- download_data('multiorganism.combined_protein.tsv');           is_diann_report(file)
file <- download_data('dilution.report.tsv');           is_diann_report(file)
```

is_fastadt	<i>Is fastadt</i>
------------	-------------------

Description

Is fastadt

Usage

```
is_fastadt(x, .xname = get_name_in_parent(x))

assert_fastadt(x, .xname = get_name_in_parent(x))
```

Arguments

x	fasta data.table
.xname	string

Examples

```
fastafile <- system.file('extdata/uniprot_hsa_20140515.fasta', package = 'autonomics')
x <- read_uniprotDT(fastafile)
# is_fastadt(x) # slow
```

is_file	<i>Is a file?</i>
---------	-------------------

Description

Is a file (and not a dir)

Usage

```
is_file(file)
```

Arguments

file	filepath
------	----------

Details

This function distinguishes between dir and file. Others dont: is.file, fs::file_exists, assertive::is_existing_file

Examples

```
dir <- tempdir(); dir.create(dir, showWarnings = FALSE)
file <- tempfile(); invisible(file.create(file))
is_file(dir)
is_file(file)
```

is_fraction	<i>Is fraction</i>
-------------	--------------------

Description

Is fraction

Usage

```
is_fraction(x, .xname = get_name_in_parent(x))

assert_is_fraction(x, .xname = get_name_in_parent(x))
```

Arguments

x	number
.xname	string

Value

TRUE or false

Examples

```
is_fraction(0.1)      # YES
is_fraction(1)        # YES
is_fraction(1.2)      # NO - more than 1
is_fraction(c(0.1, 0.2)) # NO - vector
```

is_fragpipe_tsv	<i>Is fragpipe file?</i>
-----------------	--------------------------

Description

Is fragpipe file?

Usage

```
is_fragpipe_tsv(x, .xname = get_name_in_parent(x))
```

Arguments

x	file
.xname	name of x

Examples

```
file <- NULL;                                is_fragpipe_tsv(file)
file <- 3;                                    is_fragpipe_tsv(file)
file <- 'blabla.tsv';                         is_fragpipe_tsv(file)
file <- download_data('dilution.report.tsv'); is_fragpipe_tsv(file)
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics'); is_fragpipe_tsv(file)
file <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics'); is_fragpipe_tsv(file)
file <- download_data('multiorganism.combined_protein.tsv'); is_fragpipe_tsv(file)
```

is_imputed	<i>Get/set is_imputed</i>
------------	---------------------------

Description

Get/Set is_imputed

Usage

```
is_imputed(object)

## S4 method for signature 'SummarizedExperiment'
is_imputed(object)

is_imputed(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
is_imputed(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,NULL'
is_imputed(object) <- value
```

Arguments

object	SummarizedExperiment
value	matrix

Value

matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, impute = TRUE)
sum(is_imputed(object))
```

is_maxquant_phosphosites
<i>Is maxquant phosphosites file?</i>

Description

Is maxquant phosphosites file?

Usage

```
is_maxquant_phosphosites(x, .xname = get_name_in_parent(x))
```

Arguments

x	file
.xname	name of x

Examples

```
file <- NULL; is_maxquant_phosphosites(file)
file <- 3; is_maxquant_phosphosites(file)
file <- 'blabla.tsv'; is_maxquant_phosphosites(file)
file <- download_data('dilution.report.tsv'); is_maxquant_phosphosites(file)
file <- download_data('multiorganism.combined_protein.tsv'); is_maxquant_phosphosites(file)
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics'); is_maxquant_phosphosites(file)
file <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics'); is_maxquant_phosphosites(file)
```

is_maxquant_proteingroups
<i>Is maxquant proteingroups file?</i>

Description

Is maxquant proteingroups file?

Usage

is_maxquant_proteingroups(x, .xname = get_name_in_parent(x))

Arguments

x	file
.xname	name of x

Examples

file <- NULL;	is_maxquant_proteingroups(file)
file <- 3;	is_maxquant_proteingroups(file)
file <- 'blabla.tsv';	is_maxquant_proteingroups(file)
file <- download_data('dilution.report.tsv');	is_maxquant_proteingroups(file)
file <- download_data('multiorganism.combined_protein.tsv');	is_maxquant_proteingroups(file)
file <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics');	is_maxquant_proteingroups(file)
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics');	is_maxquant_proteingroups(file)

is_non_numeric	<i>Are all variables non-numeric ?</i>
----------------	--

Description

Are all variables non-numeric ?

Usage

is_non_numeric(x)

all_non_numeric(object, formula)

Arguments

x	vector
object	SummarizedExperiment
formula	formula

Value

TRUE or FALSE

Examples

```
all_non_numeric(survobj(), ~ age)
all_non_numeric(survobj(), ~ exprs2levels)
all_non_numeric(survobj(), ~ age/exprs2levels)
all_non_numeric(survobj(), ~ age/exprs)
```

is_positive_number	<i>Is positive number</i>
--------------------	---------------------------

Description

Is positive number

Usage

```
is_positive_number(x, .xname = get_name_in_parent(x))

assert_positive_number(x, .xname = get_name_in_parent(x))

is_weakly_positive_number(x, .xname = get_name_in_parent(x))

assert_weakly_positive_number(x, .xname = get_name_in_parent(x))
```

Arguments

x	number
.xname	name of x

Value

TRUE or false

Examples

```
is_positive_number( 3)
is_positive_number(-3)
is_positive_number( 0)
is_weakly_positive_number(0)
assert_positive_number(3)
```

is_scalar_subset	<i>Is scalar subset</i>
------------------	-------------------------

Description

Is scalar subset

Usage

```
is_scalar_subset(  
  x,  
  y,  
  .xname = get_name_in_parent(x),  
  .yname = get_name_in_parent(y)  
)  
  
assert_scalar_subset(  
  x,  
  y,  
  .xname = get_name_in_parent(x),  
  .yname = get_name_in_parent(y)  
)
```

Arguments

x	scalar
y	SummarizedExperiment
.xname	name of x
.yname	name of y

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')  
object <- read_maxquant_proteingroups(file)  
is_scalar_subset('subgroup', svars(object))  
is_scalar_subset('subject', svars(object))  
assert_scalar_subset('subgroup', svars(object))
```

is_sig	<i>Is significant?</i>
--------	------------------------

Description

Is significant?

Usage

```
is_sig(  
  object,  
  fit = fits(object)[1],  
  contrast = coefs(object),  
  quantity = "fdr"  
)
```

Arguments

object	SummarizedExperiment
fit	subset of autonomics::TESTS
contrast	subset of colnames(metadata(object)[[fit]])
quantity	value in dimnames(metadata(object)[[fit]])[3]

Value

matrix: -1 (downregulated), +1 (upregulatd), 0 (not fdr significant)

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')  
object <- read_maxquant_proteingroups(file)  
object %<>% linmod_lm()  
object %<>% linmod_limma()  
issig <- is_sig(object, fit = c('lm','limma'), contrast = 'Adult-X30dpt')  
plot_contrast_venn(issig)
```

is_valid_formula	<i>Is valid formula</i>
------------------	-------------------------

Description

Is valid formula

Usage

```

is_valid_formula(
  x,
  y,
  .xname = get_name_in_parent(x),
  .yname = get_name_in_parent(y)
)

assert_valid_formula(
  x,
  y,
  .xname = get_name_in_parent(x),
  .yname = get_name_in_parent(y)
)

```

Arguments

x	formula
y	SummarizedExperiment
.xname	string
.yname	string

Value

TRUE or false

Examples

```

object <- matrix(1:9, nrow = 3)
rownames(object) <- sprintf('f%d', 1:3)
colnames(object) <- sprintf('s%d', 1:3)
object <- list(exprs = object)
object %<>% SummarizedExperiment::SummarizedExperiment()
object$group <- 'group0'
object$subgroup <- c('A', 'B', 'C')
svars(object)
  is_valid_formula( 'condition', object) # not formula
  is_valid_formula( ~condition, object) # not svar
  is_valid_formula( ~group, object) # not multilevel
  is_valid_formula( ~subgroup, object) # TRUE
  is_valid_formula( ~0+subgroup, object) # TRUE
  is_valid_formula( ~1, object) # TRUE
assert_valid_formula( ~subgroup, object)

```

keep_estimable_features
<i>Keep estimable features</i>

Description

Keep estimable features

Usage

```
keep_estimable_features(  
  object,  
  formula = ~1,  
  block = NULL,  
  coding = "code_control",  
  verbose = TRUE  
)
```

Arguments

object	SummarizedExperiment
formula	model formula
block	blockvar specification as string/character, list or formula
coding	coding function name (string)
verbose	TRUE or FALSE

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')  
object <- read_metabolon(file)  
keep_estimable_features(object, formula = ~ subgroup, block = 'Subject')
```

label2index	<i>Convert labels into indices</i>
-------------	------------------------------------

Description

Convert labels into indices

Usage

```
label2index(x)
```

Arguments

x	‘character’
---	-------------

Examples

```

label2index(x = 'Reporter intensity 0 WT(0).KD(1).OE(2).R1')
label2index(x = 'Reporter intensity 1 WT(1).KD(2).OE(3).R1')
label2index(x = 'Reporter intensity 0 WT(126).KD(127).OE(128).R1')
label2index(x = 'Reporter intensity 1 WT(126).KD(127).OE(128).R1')
label2index(x = 'Reporter intensity 1 Mix1')

```

left.vars

Get factor variables

Description

Get factor variables

Usage

```

left.vars(formula)

right.vars(formula)

factor.vars(formula, object)

## S4 method for signature 'formula,SummarizedExperiment'
factor.vars(formula, object)

## S4 method for signature 'formula,data.table'
factor.vars(formula, object)

```

Arguments

formula	formula
object	SummarizedExperiment or data.table

Value

character vector

Examples

```

object <- survobj()
formula <- survival::Surv(timetoevent, event) ~ age/exprs2levels
  all.vars(formula)
  left.vars(formula)
  right.vars(formula)
  factor.vars(formula, object)

```

 LINMOD

General Linear Model

Description

General Linear Model

Usage

```

LINMOD(
  object,
  formula = as.formula("~ subgroup"),
  engine = "limma",
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  design = create_design(object, formula = formula, drop = drop, coding = coding, verbose
    = FALSE),
  block = NULL,
  coefs = contrast_coefs(object, design = design),
  contrasts = NULL,
  weightvar = if ("weights" %in% assayNames(object)) "weights" else NULL,
  suffix = paste0("~", engine),
  verbose = TRUE,
  outdir = NULL,
  writefun = "write_xl",
  plotvolcano = FALSE,
  plotexprs = FALSE,
  argsvolcano = list(),
  argsexprs = list(),
  ...
)

linmod_limma(
  object,
  formula = as.formula("~ subgroup"),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  design = create_design(object, formula = formula, drop = drop, coding = coding, verbose
    = FALSE),
  contrasts = NULL,
  coefs = if (is.null(contrasts)) contrast_coefs(design = design) else NULL,
  block = NULL,
  weightvar = if ("weights" %in% assayNames(object)) "weights" else NULL,
  reset = TRUE,
  suffix = "~limma",
  verbose = TRUE
)

```

```

fit_limma(...)

linmod_lm(
  object,
  formula = as.formula("~ subgroup"),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  design = NULL,
  block = NULL,
  coefs = contrast_coefs(object, formula = formula, coding = coding, drop = drop),
  weightvar = if ("weights" %in% assayNames(object)) "weights" else NULL,
  reset = TRUE,
  suffix = "~lm",
  contrasts = NULL,
  verbose = TRUE
)

fit_lm(...)

linmod_lme(
  object,
  formula = as.formula("~ subgroup"),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  design = NULL,
  block = NULL,
  coefs = contrast_coefs(object, formula = formula, coding = coding, drop = drop),
  weightvar = if ("weights" %in% assayNames(object)) "weights" else NULL,
  reset = TRUE,
  opt = "optim",
  suffix = "~lme",
  contrasts = NULL,
  verbose = TRUE
)

fit_lme(...)

linmod_lmer(
  object,
  formula = as.formula("~ subgroup"),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control",
  design = NULL,
  block = NULL,
  coefs = contrast_coefs(object, formula = formula, coding = coding, drop = drop),
  weightvar = if ("weights" %in% assayNames(object)) "weights" else NULL,
  reset = TRUE,

```

```

    suffix = "~lmer",
    contrasts = NULL,
    verbose = TRUE
)

fit_lmer(...)

linmod_wilcoxon(
  object,
  formula = as.formula("~ subgroup"),
  drop = NULL,
  coding = "code_control",
  design = NULL,
  block = NULL,
  coefs = NULL,
  contrasts = NULL,
  weightvar = NULL,
  reset = TRUE,
  suffix = "~wilcoxon",
  verbose = TRUE
)

fit_wilcoxon(...)

```

Arguments

object	SummarizedExperiment
formula	model formula
engine	'limma', 'lm', 'lme', 'lmer', or 'wilcoxon'
drop	TRUE or FALSE
coding	string: codingfunname • 'contr.treatment': intercept = y_0, coefi = $y_i - y_0$ • 'contr.treatment.explicit': intercept = y_0, coefi = $y_i - y_0$ • 'code_control': intercept = y_{mean}, coefi = $y_i - y_0$ • 'contr.diff': intercept = y_0, coefi = $y_i - y_{(i-1)}$ • 'code_diff': intercept = y_{mean}, coefi = $y_i - y_{(i-1)}$ • 'code_diff_forward': intercept = y_{mean}, coefi = $y_i - y_{(i+)}$ • 'code_deviation': intercept = y_{mean}, coefi = $y_i - y_{\text{mean}}$ (drop last) • 'code_deviation_first': intercept = y_{mean}, coefi = $y_i - y_{\text{mean}}$ (drop first) • 'code_helmert': intercept = y_{mean}, coefi = $y_i - \text{mean}(y_0:(y_i-1))$ • 'code_helmert_forward': intercept = y_{mean}, coefi = $y_i - \text{mean}(y_{(i+1):yp})$
design	design matrix
block	block svar. Formated as string ('Subject') - all engines), list(Subject = ~ 1) -lme, or formula () ~ (1 Subject)) - lmer.
coefs	NULL or character vector: model coefs to record

contrasts	NULL or character vector: posthoc contrasts to record
weightvar	NULL or name of weight matrix in assays(object)
suffix	string: pvar suffix ("limma" in "p~t2~limma")
verbose	whether to msg
outdir	NULL or dir
writefun	'write_xl' or 'write_ods'
plotvolcano	TRUE or FALSE
plotexprs	TRUE or FALSE
argsvolcano	list: volcano args
argsexprs	list: expr args
...	used for s3 dispatch
reset	TRUE/FALSE whether to wipe earlier modeling results
opt	lme options

Value

Updated SummarizedExperiment

Examples

```
# Standard usage
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
LINMOD(object) # Default
LINMOD(object, ~subgroup) # Custom formula
LINMOD(object, ~subgroup, block = 'Subject') # Block effect

# Alternative engines: argument 'engine' or dedicated function
linmod_limma(object, ~subgroup, block = 'Subject') # Default engine
linmod_lm(object, ~subgroup, block = 'Subject') # Traditional
linmod_lme(object, ~subgroup, block = 'Subject') # Powerful random effects
linmod_lme(object, ~subgroup, block = list(Subject = ~1)) # using lme formula
linmod_lmer(object, ~subgroup, block = 'Subject') # Yet more powerful random effects
linmod_lmer(object, ~subgroup, block = ~(1|Subject)) # using lmer formula
linmod_wilcoxon(object, ~subgroup, block = 'Subject') # Non-parametric

# Alternative coding: backward diffs instead of baseline
linmod_limma(object, ~ subgroup, block = 'Subject', coding = 'code_diff')
linmod_lme(object, ~ subgroup, block = 'Subject', coding = 'code_diff')
linmod_lmer(object, ~ subgroup, block = 'Subject', coding = 'code_diff')

# Posthoc contrasts: limma-only, flexible, but sometimes approximate
linmod_limma(object, ~ subgroup, block = 'Subject', coding = 'code_control')
linmod_limma(object, ~ 0 + subgroup, block = 'Subject', contrasts = 't1-t0')
# flexible, but only approximate
# stat.ethz.ch/pipermail/bioconductor/2014-February/057682.html
```

```
# Top-level function also plots and writes
LINMOD(object, block = 'Subject', coefs = 't1-t0')
LINMOD(object, block = 'Subject', coefs = 't1-t0', plotvolcano = TRUE)
LINMOD(object, block = 'Subject', coefs = 't1-t0', plotexprs = TRUE)
LINMOD(object, block = 'Subject', coefs = 't1-t0', plotvolcano = TRUE, plotexprs = TRUE)
LINMOD(object, block = 'Subject', coefs = 't1-t0', plotvolcano = TRUE, plotexprs = TRUE, outdir = tempdir())
```

LINMODENGINES	<i>Linear Modeling Engines</i>
---------------	--------------------------------

Description

Linear Modeling Engines

Usage

LINMODENGINES

Format

An object of class character of length 5.

Examples

LINMODENGINES

list2mat	<i>list to matrix</i>
----------	-----------------------

Description

list to matrix

Usage

list2mat(x)

Arguments

x list

Value

matrix

Examples

```
x <- list(roundfruit = c('apple', 'orange'), redfruit = c('apple', 'strawberry'))
list2mat(x)
```

list_files	<i>list_files</i>
------------	-------------------

Description

list.files for programming

Usage

```
list_files(dir, full.names)
```

Arguments

dir	directory
full.names	TRUE or FALSE

Details

Adds a small layer on list.files. Returning NULL rather than character(0) when no files. Making it better suited for programming.

log2counts	<i>Get/Set log2counts</i>
------------	---------------------------

Description

Get / Set log2counts matrix

Usage

```
log2counts(object)

## S4 method for signature 'SummarizedExperiment'
log2counts(object)

log2counts(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
log2counts(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
log2counts(object) <- value
```

Arguments

object SummarizedExperiment
value log2count matrix (features x samples)

Value

log2count matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
log2counts(object)[1:3, 1:3]
log2counts(object) <- values(object)
```

log2cpm	<i>Get/Set log2cpm</i>
---------	------------------------

Description

Get / Set log2cpm matrix

Usage

```
log2cpm(object)

## S4 method for signature 'SummarizedExperiment'
log2cpm(object)

log2cpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
log2cpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
log2cpm(object) <- value
```

Arguments

object SummarizedExperiment
value log2cpm matrix (features x samples)

Value

log2cpm matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
log2cpm(object)[1:3, 1:3]
log2cpm(object) <- values(object)
```

log2diffs	<i>Get/Set log2diffs</i>
-----------	--------------------------

Description

Get/Set log2diffs

Usage

```
log2diffs(object)

## S4 method for signature 'SummarizedExperiment'
log2diffs(object)

log2diffs(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
log2diffs(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
log2diffs(object) <- value
```

Arguments

object	SummarizedExperiment
value	occupancy matrix (features x samples)

Value

occupancy matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
log2diffs(object)[1:3, 1:3]
```

log2proteins	<i>Get/Set log2proteins</i>
--------------	-----------------------------

Description

Get/Set log2proteins

Usage

```
log2proteins(object)

## S4 method for signature 'SummarizedExperiment'
log2proteins(object)

log2proteins(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
log2proteins(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
log2proteins(object) <- value
```

Arguments

object	SummarizedExperiment
value	occupancy matrix (features x samples)

Value

occupancy matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
log2proteins(object)[1:3, 1:3]
```

log2sites	<i>Get/Set log2sites</i>
-----------	--------------------------

Description

Get/Set log2sites

Usage

```
log2sites(object)

## S4 method for signature 'SummarizedExperiment'
log2sites(object)

log2sites(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
log2sites(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
log2sites(object) <- value
```

Arguments

object	SummarizedExperiment
value	occupancy matrix (features x samples)

Value

occupancy matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
log2sites(object)[1:3, 1:3]
```

log2tpm

Get/Set log2tpm

Description

Get / Set log2tpm matrix

Usage

```
log2tpm(object)

## S4 method for signature 'SummarizedExperiment'
log2tpm(object)

log2tpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
log2tpm(object) <- value
```

```
## S4 replacement method for signature 'SummarizedExperiment,numeric'  
log2tpm(object) <- value
```

Arguments

object SummarizedExperiment
value log2tpm matrix (features x samples)

Value

log2tpm matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')  
object <- read_rnaseq_counts(file)  
log2tpm(object) <- values(object)  
log2tpm(object)[1:3, 1:3]
```

log2transform	<i>Transform values</i>
---------------	-------------------------

Description

Transform values

Usage

```
log2transform(  
  object,  
  assay = assayNames(object)[1],  
  pseudo = 0,  
  verbose = FALSE  
)  
  
exp2transform(object, assay = assayNames(object)[1], verbose = FALSE)  
  
zscore(object, verbose = FALSE)  
  
sscale(mat, verbose = FALSE)  
  
fscale(mat, verbose = FALSE)  
  
quantnorm(object, verbose = FALSE)  
  
invnorm(object, verbose = FALSE)  
  
vsn(object, delog = TRUE, relog = delog, verbose = FALSE)
```

Arguments

object	SummarizedExperiment
assay	character vector : assays for which to perform transformation
pseudo	number : pseudo value to be added prior to transformation
verbose	TRUE or FALSE : whether to msg
mat	matrix
delog	TRUE or FALSE (vsn)
relog	TRUE or FALSE (vsn)

Value

Transformed sumexp

Examples

```

file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)

object                                %>% plot_sample_densities()
invnorm(object)                       %>% plot_sample_densities()

object                                %>% plot_sample_densities()
quantnorm(object)                     %>% plot_sample_densities()

object                                %>% plot_sample_densities()
#vsn(object)                          %>% plot_sample_densities() # dataset too small

object                                %>% plot_sample_densities()
zscore(object)                        %>% plot_sample_densities()

object                                %>% plot_sample_densities()
exp2transform(object)                 %>% plot_sample_densities()
log2transform(exp2transform(object)) %>% plot_sample_densities()

```

logical2factor	<i>logical to factor</i>
----------------	--------------------------

Description

logical to factor

Usage

```

logical2factor(x, true = get_name_in_parent(x), false = paste0("not", true))

factor2logical(x)

```

Arguments

x	logical vector
true	string : truelevel
false	string : falselevel

Value

factor

Examples

```
t1up <- c( TRUE,  FALSE,  TRUE)
t1  <- c('flat', 'down', 'up' ) %>% factor(., .)
t1up
logical2factor(t1up)
factor2logical(t1)
```

make_alpha_palette	<i>Make alpha palette</i>
--------------------	---------------------------

Description

Make alpha palette

Usage

```
make_alpha_palette(object, alpha)
```

Arguments

object	SummarizedExperiment
alpha	string

Value

character vector

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
make_alpha_palette(object, 'Time')
```

make_colors	<i>Make colors</i>
-------------	--------------------

Description

Make colors

Usage

```
make_colors(
  varlevels,
  sep = guess_sep(varlevels),
  show = FALSE,
  verbose = FALSE
)
```

Arguments

varlevels	character vector
sep	string
show	TRUE or FALSE: whether to plot
verbose	TRUE or FALSE: whether to msg

Examples

```
make_colors(c('A', 'B', 'C', 'D' ), show = TRUE)
make_colors(c('A.1', 'B.1', 'A.2', 'B.2'), show = TRUE)
```

make_volcano_dt	<i>Create volcano datatable</i>
-----------------	---------------------------------

Description

Create volcano datatable

Usage

```
make_volcano_dt(
  object,
  fit = fits(object)[1],
  coefs = coefs(object, fit = fit)[1],
  shape = "imputed",
  size = NULL,
  alpha = NULL,
  label = if ("gene" %in% fvars(object)) "gene" else "feature_id"
)
```

Arguments

object	SummarizedExperiment
fit	'limma', 'lme', 'lm', 'wilcoxon'
coefs	character vector: coefs for which to plot volcanoes
shape	fvar or NULL
size	fvar or NULL
alpha	fvar or NULL
label	fvar or NULL

Value

data.table

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, impute = TRUE, fit = 'limma')
make_volcano_dt(object, fit = 'limma', coefs = 'Adult-X30dpt')
```

map_fvalues	<i>Map fvalues</i>
-------------	--------------------

Description

Map fvalues

Usage

```
map_fvalues(object, fvalues, from = "uniprot", to = "feature_id", sep = ";")
```

Arguments

object	SummarizedExperiment
fvalues	uncollapsed string vector
from	string (fvar)
to	string (svar)
sep	collapse separator

Value

string vector

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
fdt(object)
map_fvalues(object, c('Q6DHL5', 'Q6PFS7'), from = 'uniprot', to = 'feature_id', sep = ';')
```

matrix2sumexp	<i>Convert matrix into SummarizedExperiment</i>
---------------	---

Description

Convert matrix into SummarizedExperiment

Usage

```
matrix2sumexp(x, verbose = TRUE)
```

Arguments

x	matrix
verbose	TRUE/FALSE

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
x <- values(read_metabolon(file))
object <- matrix2sumexp(x)
object %<>% pca()
biplot(object, color = 'subgroup')
```

MAXQUANT_PATTERNS	<i>maxquant quantity patterns</i>
-------------------	-----------------------------------

Description

maxquant quantity patterns

Usage

```
MAXQUANT_PATTERNS
```

Format

An object of class character of length 7.

Examples

```
MAXQUANT_PATTERNS
```

mclust_breaks	<i>Mixture/Quantile breaks</i>
---------------	--------------------------------

Description

Mixture/Quantile breaks

Usage

```
mclust_breaks(x, k = NULL)

mixtools_breaks(x, k = 2)

quantile_breaks(x, k = 3, probs = seq_len(k - 1)/k)
```

Arguments

x	numeric
k	number
probs	probabilities

Examples

```
set.seed(1)
x <- c(rnorm(20, 3), rnorm(20,7), rnorm(20, 11))
mclust_breaks(x)
mixtools_breaks(x, k = 3)
quantile_breaks(x)
```

mdsplot

*Feature correlations/distances***Description**

Feature correlations/distances

Usage`mdsplot(distmat, title = NULL)``fcor(object, verbose = TRUE)``scor(object, verbose = TRUE)``fdist(object, method = "cor")``sdist(object, method = "cor")`**Arguments**

<code>distmat</code>	distance matrix
<code>title</code>	NULL or string
<code>object</code>	SummarizedExperiment
<code>verbose</code>	TRUE or FALSE
<code>method</code>	'cor', 'euclidian', etc

Value

matrix

Examples

```
# Correlations
object <- twofactor_sumexp()
scor(object)           %>% pheatmap::pheatmap()
fcor(object)           %>% pheatmap::pheatmap()

# Distances
sdist(object, 'cor')    %>% mdsplot('samples: cor')
sdist(object, 'euclidian') %>% mdsplot('samples: euclidian')
fdist(object, 'cor')    %>% mdsplot('features: cor')
fdist(object, 'euclidian') %>% mdsplot('features: euclidian')
```

merge_compounddiscoverer	<i>merge compound discoverer files</i>
--------------------------	--

Description

merge compound discoverer files

Usage

merge_compounddiscoverer(x, quantity = NULL, verbose = TRUE)

Arguments

x	'list'
quantity	'area', 'normalizedarea'
verbose	'TRUE' or 'FALSE'

Value

'data.table'

merge_sample_excel	<i>Merge sample excel</i>
--------------------	---------------------------

Description

Merge sample excel

Usage

```
merge_sample_excel(  
  object,  
  sfile,  
  range = NULL,  
  by.x = "sample_id",  
  by.y = "sample_id"  
)
```

Arguments

object	SummarizedExperiment
sfile	sample file
range	string
by.x	string
by.y	string

Value

SummarizedExperiment

merge_sample_file	<i>Merge sample / feature file</i>
-------------------	------------------------------------

Description

Merge sample / feature file

Usage

```
merge_sample_file(
  object,
  sfile = NULL,
  by.x = "sample_id",
  by.y = "sample_id",
  all.x = TRUE,
  select = NULL,
  stringsAsFactors = FALSE,
  verbose = TRUE
)

merge_ffile(
  object,
  ffile = NULL,
  by.x = "feature_id",
  by.y = "feature_id",
  all.x = TRUE,
  select = NULL,
  stringsAsFactors = FALSE,
  verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
sfile	string : sample file path
by.x	string : object mergevar
by.y	string : file mergevvar
all.x	TRUE / FALSE : whether to keep samples / feature without annotation
select	character : [sf]file columns to select
stringsAsFactors	TRUE / FALSE
verbose	TRUE / FALSE
ffile	string : ffile path

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
subgroups <- c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00')
subgroups %<>% paste0('_STD')
object <- read_maxquant_proteingroups(file, subgroups = subgroups)
sfile <- paste0(tempdir(), '/', basename(tools::file_path_sans_ext(file)))
sfile %<>% paste0('.samples.txt')
dt <- data.table(sample_id = object$sample_id,
                 day = split_extract_fixed(object$subgroup, '_', 1))
data.table::fwrite(dt, sfile)
sdt(object)
sdt(merge_sample_file(object, sfile))
```

merge_sdata	<i>Merge sample/feature dt</i>
-------------	--------------------------------

Description

Merge sample/feature dt

Usage

```
merge_sdata(
  object,
  dt,
  by.x = "sample_id",
  by.y = names(dt)[1],
  all.x = TRUE,
  verbose = TRUE
)

merge_sdt(
  object,
  dt,
  by.x = "sample_id",
  by.y = "sample_id",
  all.x = TRUE,
  verbose = TRUE
)

merge_fdata(
  object,
  dt,
  by.x = "feature_id",
```

```
by.y = names(dt)[1],
all.x = TRUE,
verbose = TRUE
)

merge_fdt(
  object,
  dt,
  by.x = "feature_id",
  by.y = "feature_id",
  all.x = TRUE,
  verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
dt	data.frame, data.table, DataFrame
by.x	string : object mergevar
by.y	string : df mergevar
all.x	TRUE / FALSE : whether to keep samples / features without annotation
verbose	TRUE / FALSE : whether to msg

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
sdt(object)
sdt(merge_sdt(object, data.table(sample_id = object$sample_id,
                                number = seq_along(object$sample_id))))
```

message_df	<i>message dataframe</i>
------------	--------------------------

Description

message dataframe using sprintf syntax. Use place holder ' '

Usage

```
message_df(format_string, x)
```

Arguments

format_string sprintf style format string
x data.frame

Value

nothing returned

Examples

```
x <- data.frame(feature_id = c('F001', 'F002'), symbol = c('FEAT1', 'FEAT2'))
message_df('\t%s', x)

x <- c(rep('PASS', 25), rep('FAIL', 25))
message_df(format_string = '%s', table(x))
```

modelvar	<i>Get model variable</i>
----------	---------------------------

Description

Get model variable

Usage

```
modelvar(object, ...)
```

```
## S3 method for class 'data.table'
modelvar(
  object,
  quantity,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit),
  ...
)
```

```
## S3 method for class 'SummarizedExperiment'
modelvar(
  object,
  quantity,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit),
  ...
)
```

```
## S3 method for class '`NULL`'
modelvar(object, ...)
```

```

effectvar(
  object,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit)
)

tvar(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

pvar(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

fdrvar(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

abstractvar(object, ...)

## S3 method for class 'data.table'
abstractvar(
  object,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit),
  ...
)

## S3 method for class 'SummarizedExperiment'
abstractvar(
  object,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit),
  ...
)

modelvec(object, ...)

## S3 method for class 'data.table'
modelvec(
  object,
  quantity,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  fvar = "feature_id",
  ...
)

## S3 method for class 'SummarizedExperiment'
modelvec(
  object,
  quantity,
  fit = fits(object)[1],

```

```

    coef = autonomics::coefs(object, fit = fit)[1],
    fvar = "feature_id",
    ...
)

effectvec(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object)[1],
  fvar = "feature_id"
)

tvec(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  fvar = "feature_id"
)

pvec(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  fvar = "feature_id"
)

fdrvec(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  fvar = "feature_id"
)

abstractvec(object, ...)

## S3 method for class 'data.table'
abstractvec(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  fvar = "feature_id",
  ...
)

## S3 method for class 'SummarizedExperiment'
abstractvec(
  object,
  fit = fits(object)[1],

```

```

    coef = autonomics::coefs(object, fit = fit)[1],
    fvar = "feature_id",
    ...
)

modeldt(object, ...)

## S3 method for class 'data.table'
modeldt(
  object,
  quantity,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit),
  ...
)

## S3 method for class 'SummarizedExperiment'
modeldt(
  object,
  quantity,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit),
  ...
)

## S3 method for class '`NULL`'
modeldt(object, ...)

effectdt(
  object,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit)
)

tdt(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

pdt(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

modelmat(
  object,
  quantity,
  fit = fits(object),
  coef = autonomics::coefs(object, fit = fit)
)

modelmat(
  object,
  quantity,

```

```

    fit = fits(object),
    coef = autonomics::coefs(object, fit = fit)
  )

  effectmat(
    object,
    fit = fits(object),
    coef = autonomics::coefs(object, fit = fit)
  )

  effectsizemat(
    object,
    fit = fits(object),
    coef = autonomics::coefs(object, fit = fit)
  )

  tmat(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

  pmat(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

  fdrmat(object, fit = fits(object), coef = autonomics::coefs(object, fit = fit))

  modelfeatures(object, ...)

  ## S3 method for class 'data.table'
  modelfeatures(
    object,
    fit = fits(object)[1],
    coef = autonomics::coefs(object, fit = fit)[1],
    fvar = "feature_id",
    significancevar = "p",
    significance = 0.05,
    effectdirection = "<>",
    effectsize = 0,
    ...
  )

  ## S3 method for class 'SummarizedExperiment'
  modelfeatures(object, ...)

  upfeatures(
    object,
    fit = fits(object)[1],
    coef = autonomics::coefs(object, fit = fit)[1],
    fvar = "feature_id",
    significancevar = "p",
    significance = 0.05,
    effectsize = 0
  )

```

```

)

downfeatures(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  fvar = "feature_id",
  significancevar = "p",
  significance = 0.05,
  effectsize = 0
)

```

Arguments

object	data.table or SummarizedExperiment
...	S3 dispatch
quantity	'p', 'effect', 'fdr', 't', or 'se'
fit	string (vector)
coef	string (vector)
fvar	'feature_id' or other fvar for values (pvec) or names (upfeatures)
significancevar	'p' or 'fdr'
significance	p or fdr cutoff (fractional number)
effectdirection	'<>', '<' or '>'
effectsize	effectsize cutoff (positive number)

Value

string (tvar), matrix (tmat), numeric vector (tvec), character vector (tfeatures)

Examples

```

file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma()
object %<>% linmod_lm()

effectvar(object)
effectvec(object)[1:3]
effecttdt(object)[1:3, ]
effectmat(object)[1:3, ]

tvar(object)
tvec(object)[1:3]
tdt(object)[1:3, ]
tmat(object)[1:3, ]

```

```
pvar(object)
pvec(object)[1:3]
pdt(object)[1:3, ]
pmat(object)[1:3, ]

modelfeatures(object)
downfeatures(object)
upfeatures(object)
```

MSIGCOLLECTIONSHUMAN	<i>Human/Mouse Msigdb Collections</i>
----------------------	---------------------------------------

Description

Human/Mouse Msigdb Collections

Usage

```
MSIGCOLLECTIONSHUMAN
MSIGCOLLECTIONSMOUSE
```

Format

- An object of class character of length 25.
- An object of class character of length 13.

MSIGDIR	<i>local msigdb dir</i>
---------	-------------------------

Description

local msigdb dir

Usage

```
MSIGDIR
```

Format

- An object of class character of length 1.

nfactors	<i>stri_split and extract</i>
----------	-------------------------------

Description

stri_split and extract

Usage

```
nfactors(x, sep = guess_sep(x))  
  
split_extract_fixed(x, sep, i)  
  
split_extract_regex(x, sep, i)  
  
split_extract(x, i, sep = guess_sep(x))
```

Arguments

x	character vector
sep	string
i	integer

Value

character vector

Examples

```
# Read  
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')  
object <- read_metabolon(file)  
x <- object$sample_id[1:5]  
nfactors(x)  
  
# Split  
split_extract_fixed(x, '.', 1:2)  
split_extract_fixed(x, '.', seq_len(nfactors(x)-1))  
split_extract_fixed(x, '.', nfactors(x))  
split_extract_fixed(fdt(object)$PUBCHEM, ';', 1) # with NA values
```

object1	<i>Example objects for binding</i>
---------	------------------------------------

Description

Example objects for binding

Usage

object1()

object2()

Value

SummarizedExperiment

Examples

object1()
object2()

OPENTARGETSDIR	<i>opentargets dir</i>
----------------	------------------------

Description

opentargets dir

Usage

OPENTARGETSDIR

Format

An object of class character of length 1.

order_on_p	<i>Order on p</i>
------------	-------------------

Description

Order on p

Usage

```
order_on_p(  
  object,  
  fit = autonomics::fits(object),  
  coefs = autonomics::coefs(object, fit = fit),  
  combiner = "|",  
  decreasing = FALSE,  
  verbose = TRUE  
)  
  
order_on_t(  
  object,  
  fit = autonomics::fits(object),  
  coefs = autonomics::coefs(object, fit = fit),  
  combiner = "|",  
  decreasing = FALSE,  
  verbose = TRUE  
)  
  
order_on_effect(  
  object,  
  fit = autonomics::fits(object),  
  coefs = autonomics::coefs(object, fit = fit),  
  combiner = "|",  
  verbose = TRUE  
)
```

Arguments

object	SummarizedExperiment
fit	string vector: subset of 'fits(object)'
coefs	string vector: subset of 'coefs(object)'
combiner	' ' or '&'
decreasing	TRUE or FALSE
verbose	TRUE or FALSE

Value

SummarizedExperiment

Examples

```
# Linmod
  file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
  object <- read_metabolon(file)
  order_on_p(object)
  object %<>% linmod_limma()
  order_on_p(object)
# Survival
  object <- survobj()
  object %<>% fit_survival()
  order_on_p(object)
```

overall_parameters	<i>Distribution parameters</i>
--------------------	--------------------------------

Description

Mean, sd, weight of overall/mixture distribution

Usage

```
overall_parameters(x)

mclust_parameters(x, k = NULL)

mixtools_parameters(x, k = 2)
```

Arguments

- x numeric vector
- k number of components

Value

data.table (mean, sd, weight)

Examples

```
set.seed(1)
x <- c(rnorm(20, 3), rnorm(20,7), rnorm(20, 11))
overall_parameters(x)
mclust_parameters(x)
mixtools_parameters(x)
```

pca

PCA, SMA, LDA, PLS, SPLS, OPLS

Description

Perform a dimension reduction. Store sample scores, feature loadings, and dimension variances.

Usage

```
pca(  
  object,  
  by = "sample_id",  
  assay = assayNames(object)[1],  
  ndim = 2,  
  minvar = 0,  
  center_samples = TRUE,  
  verbose = TRUE,  
  plot = FALSE,  
  ...  
)
```

```
pls(  
  object,  
  by = "subgroup",  
  assay = assayNames(object)[1],  
  ndim = 2,  
  minvar = 0,  
  verbose = FALSE,  
  plot = FALSE,  
  ...  
)
```

```
sma(  
  object,  
  by = "sample_id",  
  assay = assayNames(object)[1],  
  ndim = 2,  
  minvar = 0,  
  verbose = TRUE,  
  plot = FALSE,  
  ...  
)
```

```
lda(  
  object,  
  assay = assayNames(object)[1],  
  by = "subgroup",
```

```

    ndim = 2,
    minvar = 0,
    verbose = TRUE,
    plot = FALSE,
    ...
)

spls(
  object,
  assay = assayNames(object)[1],
  by = "subgroup",
  ndim = 2,
  minvar = 0,
  plot = FALSE,
  ...
)

opls(
  object,
  by = "subgroup",
  assay = assayNames(object)[1],
  ndim = 2,
  minvar = 0,
  verbose = FALSE,
  plot = FALSE,
  ...
)

```

Arguments

object	SummarizedExperiment
by	svar or NULL
assay	string
ndim	number
minvar	number
center_samples	TRUE/FALSE: center samples prior to pca ?
verbose	TRUE/FALSE: message ?
plot	TRUE/FALSE: plot ?
...	passed to biplot

Value

SummarizedExperiment

Author(s)

Aditya Bhagwat, Laure Cougnaud (LDA)

Examples

```

file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
pca(object, plot = TRUE)    # Principal Component Analysis
pls(object, plot = TRUE)    # Partial Least Squares
lda(object, plot = TRUE)    # Linear Discriminant Analysis
sma(object, plot = TRUE)    # Spectral Map Analysis
spls(object, plot = TRUE)   # Sparse PLS
# opls(object, plot = TRUE) # OPLS # outcommented because it produces a file named FALSE

```

pg_to_canonical	<i>proteingroup to isoforms</i>
-----------------	---------------------------------

Description

proteingroup to isoforms

Usage

```

pg_to_canonical(x, unique = TRUE)

pg_to_isoforms(x, unique = TRUE)

```

Arguments

x	proteingroups string vector
unique	whether to remove duplicates

Value

string vector

Examples

```

(x <- c('Q96JP5;Q96JP5-2', 'Q96JP5', 'Q96JP5-2;P86791'))
pg_to_isoforms(x)
pg_to_canonical(x)
pg_to_isoforms(x, unique = FALSE)
pg_to_canonical(x, unique = FALSE)
# .pg_to_isoforms(x[1]) # unexported dot functions
# .pg_to_canonical(x[1]) # operate on scalars

```

plot_coef_densities	<i>Plot contrast densities</i>
---------------------	--------------------------------

Description

Plot contrast densities

Usage

```
plot_coef_densities(  
  object,  
  fit = fits(object)[1],  
  coefs = autonomics::coefs(object, fit = fit),  
  label = "feature_id"  
)
```

Arguments

object	SummarizedExperiment
fit	'limma', 'lm', 'lme', 'lmer', or 'wilcoxon'
coefs	character vector
label	svar

Value

ggplot

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')  
object <- read_metabolon(file)  
object %<>% linmod_limma(~subgroup, block = 'Subject')  
plot_coef_densities(object)
```

plot_contrastogram	<i>Plot contrastogram</i>
--------------------	---------------------------

Description

Plot contrastogram

Usage

```
plot_contrastogram(  
  object,  
  subgroupvar,  
  formula = as.formula(paste0("~ 0 +", subgroupvar)),  
  colors = make_colors(slevels(object, subgroupvar), guess_sep(object)),  
  curve = 0.1  
)
```

Arguments

object	SummarizedExperiment
subgroupvar	subgroup svar
formula	formula
colors	named color vector (names = subgroups)
curve	arrow curvature

Value

list returned by [plotmat](#)

Examples

```
if (installed('diagram')){  
  file <- download_data('halama18.metabolon.xlsx')  
  object <- read_metabolon(file)  
  plot_contrastogram(object, subgroupvar = 'subgroup')  
}
```

plot_contrast_venn	<i>Plot contrast venn</i>
--------------------	---------------------------

Description

Plot contrast venn

Usage

```
plot_contrast_venn(issig, colors = NULL)
```

Arguments

issig	matrix(nrow, ncontrast): -1 (down), +1 (up)
colors	NULL or colorvector

Value

nothing returned

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_wilcoxon(~ subgroup, block = 'Subject')
object %<>% linmod_limma( ~ subgroup, block = 'Subject')
isfdr <- is_sig(object, contrast = 't3-t0', quantity = 'p', fit = fits(object))
plot_contrast_venn(isfdr)
```

plot_data	<i>Plot data</i>
-----------	------------------

Description

Plot data

Usage

```
plot_data(
  data,
  geom = geom_point,
  color = NULL,
  fill = NULL,
  linetype = NULL,
  ...,
  palette = NULL,
  fixed = list(),
  theme = list()
)
```

Arguments

data	data.frame'
geom	geom_point, etc.
color	variable mapped to color (symbol)
fill	variable mapped to fill (symbol)
linetype	variable mapped to linetype (symbol)
...	mapped aesthetics
palette	color palette (named character vector)
fixed	fixed aesthetics (list)
theme	list with ggplot theme specifications

Value

ggplot object

Author(s)

Aditya Bhagwat, Johannes Graumann

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% pca()
data <- sdt(object)
plot_data(data, x = `t~sample_id~pca1`, y = `t~sample_id~pca2`)
plot_data(data, x = `t~sample_id~pca1`, y = `t~sample_id~pca2`, color = subgroup)
plot_data(data, x = `t~sample_id~pca1`, y = `t~sample_id~pca2`, color = NULL)
fixed <- list(shape = 15, size = 3)
plot_data(data, x = `t~sample_id~pca1`, y = `t~sample_id~pca2`, fixed = fixed)
```

plot_densities	<i>Plot sample/feature distributions</i>
----------------	--

Description

Plot sample/feature distributions

Usage

```
plot_densities(
  object,
  assay = assayNames(object)[1],
  group,
  fill,
  color = NULL,
  linetype = NULL,
  facet = NULL,
  nrow = NULL,
  ncol = NULL,
  dir = "h",
  scales = "free_y",
  labeller = label_value,
  palette = NULL,
  fixed = list(alpha = 0.8, na.rm = TRUE)
)

plot_sample_densities(
  object,
  assay = assayNames(object)[1],
```

```

    group = "sample_id",
    fill = if ("subgroup" %in% svars(object)) "subgroup" else "sample_id",
    color = NULL,
    linetype = NULL,
    n = 100,
    facet = NULL,
    nrow = NULL,
    ncol = NULL,
    dir = "h",
    scales = "free_y",
    labeller = label_value,
    palette = NULL,
    fixed = list(alpha = 0.8, na.rm = TRUE)
  )

plot_feature_densities(
  object,
  assay = assayNames(object)[1],
  fill = "feature_id",
  group = fill,
  color = NULL,
  linetype = NULL,
  n = 9,
  facet = NULL,
  nrow = NULL,
  ncol = NULL,
  dir = "h",
  scales = "free",
  labeller = label_value,
  palette = NULL,
  fixed = list(alpha = 0.8, na.rm = TRUE)
)

```

Arguments

object	SummarizedExperiment
assay	string
group	svar (string)
fill	svar (string)
color	svar (string)
linetype	svar (string)
facet	svar (character vector)
nrow	number of facet rows
ncol	number of facet cols
dir	'h' (horizontal) or 'v' (vertical)
scales	'free', 'fixed', 'free_y'

labeller	e.g. label_value
palette	named character vector
fixed	fixed aesthetics
n	number

Value

ggplot object

See Also

[plot_sample_violins](#), [plot_sample_boxplots](#)

Examples

```
# Data
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% extract(, order(.$subgroup))

# Sample distributions
plot_sample_densities(object)
plot_sample_violins( object, facet = 'Time')
plot_sample_boxplots(object)
plot_exprs(object)
plot_exprs(object, dim = 'samples', x = 'subgroup', facet = 'Time')

# Feature distributions
plot_feature_densities(object)
plot_feature_violins( object)
plot_feature_boxplots( object)
```

plot_densities_transforms

Visually evaluate transformation effects

Description

Visually evaluate transformation effects

Usage

```
plot_densities_transforms(
  object,
  assay = assayNames(object)[1],
  subgroupvar = "subgroup",
  transforms = c("center", "invnorm", "quantnorm", "vsn", "zscore"),
  ...,
```

```

    fixed = list(na.rm = TRUE, show.legend = FALSE, verbose = FALSE),
    verbose = TRUE
)

plot_violins_transforms(
  object,
  assay = assayNames(object)[1],
  subgroupvar = "subgroup",
  transforms = c("center", "invnorm", "quantnorm", "vsn", "zscore"),
  ...,
  fixed = list(na.rm = TRUE, trim = FALSE, draw_quantiles = c(0.25, 0.5, 0.75),
    show.legend = FALSE),
  verbose = TRUE
)

biplot_transforms(
  object,
  assay = assayNames(object)[1],
  subgroupvar = "subgroup",
  transforms = TRANSFORMSTRICT,
  method = DIMREDENGINE[1],
  dims = 1:2,
  color = subgroupvar,
  shape = NULL,
  size = NULL,
  alpha = NULL,
  group = NULL,
  label = NULL,
  ncol = NULL,
  nrow = NULL,
  ...,
  fixed = list(shape = 15, size = 3),
  verbose = FALSE
)

biplot_transforms_assays(
  object,
  assays = assayNames(object)[1],
  subgroupvar = "subgroup",
  transforms = TRANSFORMSTRICT,
  method = DIMREDENGINE[1],
  dims = 1:2,
  color = subgroupvar,
  shape = NULL,
  size = NULL,
  alpha = NULL,
  group = NULL,
  label = NULL,

```

```

    ...,
    verbose = FALSE,
    fixed = list(shape = 15, size = 3)
  )

```

Arguments

object	SummarizedExperiment
assay	string : assay name to operate on
subgroupvar	svar
transforms	character vector : transformations explored
...	: further plotting parameters
fixed	list : fixed aesthetics
verbose	TRUE/FALSE : message?
method	string : dimension reduction technique
dims	numbers : biplot dimensions
color	svar
shape	svar
size	svar
alpha	svar
group	svar
label	svar
ncol	integer : columns for facet wrapping
nrow	integer : rows for facet wrapping
assays	character vector : assay names to operate on

Value

ggplot2 object

Author(s)

Johannes Graumann

Examples

```

file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)

# `vsn` implemented, but example data set too small
transformations <- c(
  'center_mean', 'center_median', 'invnorm', 'quantnorm', 'zscore')

# object %>% plot_densities_transforms(transforms = transformations) # Requires package ggridges
object %>% plot_violins_transforms(transforms = transformations)

```

```

object %>% biplot_transforms(
  method = 'pca', transforms = transformations, nrow = 2)
object %>% biplot_transforms(
  method = 'pls', transforms = transformations, nrow = 2)

object[['replicate']] <- gsub('^.*\\.(.+)$', '\\1', object[['sample_id']])
object %>%
  biplot_transforms(
    transforms = transformations, label = 'replicate')

```

plot_design

Plot model

Description

Plot model

Usage

```
plot_design(object, coding = "code_control")
```

Arguments

object	SummarizedExperiment
coding	string: codingfunname

- contr.treatment: intercept = y_0 , coefi = $y_i - y_0$
- contr.treatment.explicit: intercept = y_0 , coefi = $y_i - y_0$
- code_control: intercept = ymean, coefi = $y_i - y_0$
- contr.diff: intercept = y_0 , coefi = $y_i - y(i-1)$
- code_diff: intercept = ymean, coefi = $y_i - y(i-1)$
- code_diff_forward: intercept = ymean, coefi = $y_i - y(i+)$
- code_deviation: intercept = ymean, coefi = $y_i - ymean$ (drop last)
- code_deviation_first: intercept = ymean, coefi = $y_i - ymean$ (drop first)
- code_helmert: intercept = ymean, coefi = $y_i - mean(y_0:(y_i-1))$
- code_helmert_forward: intercept = ymean, coefi = $y_i - mean(y(i+1):yp)$

Value

ggplot

Examples

```

file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
subgroups <- paste0(c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00'), '_STD')
object <- read_maxquant_proteingroups(file, subgroups = subgroups)
object$subgroup %<>% substr(1,3)
plot_design(object)

```

plot_exprs	<i>Plot exprs for coef</i>
------------	----------------------------

Description

Plot exprs for coef

Usage

```
plot_exprs(
  object,
  dim = "both",
  assay = assayNames(object)[1],
  features = NULL,
  fit = fits(object)[1],
  coefs = autonomics::coefs(object, fit = fit),
  block = NULL,
  x = default_x(object, dim),
  geom = default_geom(object, x = x, block = block),
  color = x,
  fill = x,
  shape = NULL,
  size = NULL,
  alpha = NULL,
  linetype = NULL,
  highlight = NULL,
  combiner = "|",
  p = 1,
  fdr = 1,
  facet = if (dim == "both") "feature_id" else NULL,
  file = NULL,
  width = 7,
  height = 7,
  n = if (is.null(file)) 4 else 12,
  ncol = if (is.null(file)) NULL else 3,
  nrow = if (is.null(file)) NULL else 4,
  scales = "free_y",
  labeller = "label_value",
  pointsize = if (is.null(block)) 0 else 0.5,
  jitter = if (is.null(block)) 0.1 else 0,
  fillpalette = make_var_palette(object, fill),
  colorpalette = make_var_palette(object, color),
  hlevels = NULL,
  title = switch(dim, both = x, features = "Feature Boxplots", samples =
    "Sample Boxplots"),
  subtitle = if (!is.null(fit)) coefs else "",
  xlab = x,
```

```

    ylab = "value",
    theme = ggplot2::theme(plot.title = element_text(hjust = 0.5)),
    guides = NULL,
    verbose = TRUE
  )

  plot_sample_boxplots(
    object,
    fill = if ("subgroup" %in% svars(object)) "subgroup" else "sample_id",
    n = min(ncol(object), 16),
    ...
  )

  plot_feature_boxplots(object, ...)

```

Arguments

object	SummarizedExperiment
dim	'samples' (per-sample distribution across features), 'features' (per-feature distribution across samples) or 'both' (subgroup distribution faceted per feature)
assay	string: value in assayNames(object)
features	features to plot no matter what (character vector)
fit	'limma', 'lm', 'lme', 'lmer', 'wilcoxon'
coefs	subset of coefs(object) to consider in selecting top
block	group svar
x	x svar
geom	'boxplot' or 'point'
color	color svar: points, lines
fill	fill svar: boxplots
shape	shape svar
size	size svar
alpha	alpha svar
linetype	linetype svar
highlight	highlight svar
combiner	'&' or ' '
p	fraction: p cutoff
fdr	fraction: fdr cutoff
facet	string: fvar mapped to facet
file	NULL or filepath
width	inches
height	inches

n	number of samples (dim = 'samples') or features (dim = 'features' or 'both') to plot
ncol	number of cols in faceted plot (if dim = 'both')
nrow	number of rows in faceted plot (if dim = 'both')
scales	'free_y', 'free_x', 'fixed'
labeller	string or function
pointsize	number
jitter	jitter width (number)
fillpalette	named character vector: fill palette
colorpalette	named character vector: color palette
hlevels	xlevels for which to plot hlines
title	string
subtitle	string
xlab	string
ylab	string
theme	ggplot2::theme(...) or NULL
guides	NULL or c(fill = 'none', color = 'none')
verbose	TRUE or FALSE
...	used to maintain deprecated functions

Value

ggplot object

See Also

[plot_sample_densities](#), [plot_sample_violins](#)

Examples

```
# Without limma
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
plot_exprs(object, block = 'Subject', title = 'Subgroup Boxplots')
plot_exprs(object, dim = 'samples')
plot_exprs(object, dim = 'features', block = 'sample_id')

# With limma
object %<>% linmod_limma(block = 'Subject')
plot_exprs(object, block = 'Subject')
plot_exprs(object, block = 'Subject', coefs = c('t1-t0', 't2-t0', 't3-t0'))
plot_exprs_per_coef(object, x = 'Time', block = 'Subject')

# Points
plot_exprs(object, geom = 'point', block = 'Subject')

# Add highlights
controlfeatures <- c('biotin', 'phosphate')
```

```

    fdt(object) %<>% cbind(control = .$feature_name %in% controlfeatures)
    plot_exprs(object, dim = 'samples', highlight = 'control')
# Multiple pages
    plot_exprs(object, block = 'Subject', n = 4, nrow = 1, ncol = 2)

```

plot_exprs_per_coef *Plot exprs per coef*

Description

Plot exprs per coef

Usage

```

plot_exprs_per_coef(
  object,
  fit = fits(object)[1],
  coefs = autonomics::coefs(object, fit = fit),
  x = default_x(object),
  block = NULL,
  geom = default_geom(object, x, block = block),
  orderbyp = FALSE,
  title = x,
  subtitle = default_subtitle(fit, x, coefs),
  n = 1,
  nrow = 1,
  ncol = NULL,
  theme = ggplot2::theme(legend.position = "bottom", legend.title = element_blank(),
    plot.title = element_text(hjust = 0.5), plot.subtitle = element_text(hjust = 0.5)),
  ...
)

```

Arguments

object	SummarizedExperiment
fit	'limma', 'lm', 'lme', 'lmer', 'wilcoxon'
coefs	subset of coefs(object) to consider in selecting top
x	x svar
block	group svar
geom	'boxplot' or 'point'
orderbyp	TRUE or FALSE
title	string
subtitle	string
n	number

nrow	number of rows in faceted plot
ncol	number of cols in faceted plot
theme	ggplot2::theme(...) or NULL
...	passed to plot_exprs

Value

ggplot object

See Also

[plot_sample_densities](#), [plot_sample_violins](#)

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma()
object %<>% pls(by = 'subgroup')
object %<>% pls(by = 'Diabetes')
object %<>% pls(by = 'Subject')
plot_exprs_per_coef(object)
plot_exprs_per_coef(object, orderbyp = TRUE)
plot_exprs_per_coef(object, fit = 'pls1', block = 'Subject')
```

plot_fit_summary	<i>Plot fit summary</i>
------------------	-------------------------

Description

Plot fit summary

Usage

```
plot_fit_summary(sumdt, nrow = NULL, ncol = NULL, order = FALSE)
```

Arguments

sumdt	data.table
nrow	number
ncol	number
order	TRUE or FALSE

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_lm()
object %<>% linmod_limma(block = 'Subject')
sumdt <- summarize_fit(object, coefs = c('t1-t0', 't2-t0', 't3-t0'))
plot_fit_summary(sumdt)
```

plot_heatmap	<i>Plot heatmap</i>
--------------	---------------------

Description

Plot heatmap

Usage

```
plot_heatmap(
  object,
  fit = fits(object)[1],
  coef = autonomics::coefs(object, fit = fit)[1],
  effectsize = 0,
  p = 1,
  fdr = 0.05,
  n = 100,
  assay = assayNames(object)[1],
  cluster_features = FALSE,
  cluster_samples = FALSE,
  flabel = intersect(c("gene", "feature_id"), fvars(object))[1],
  group = "subgroup",
  verbose = TRUE,
  title = NULL
)
```

Arguments

object	SummarizedExperiment
fit	'limma', 'lm', 'lme(r)', 'wilcoxon'
coef	string: one of coefs(object)
effectsize	number: effectsize filter
p	number: p filter
fdr	number: fdr filter
n	number: n filter
assay	string: one of assayNames(object)

cluster_features
TRUE or FALSE
cluster_samples
TRUE or FALSE
flabel
string: feature label
group
sample groupvar
verbose
TRUE or FALSE
title
string

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, fit = 'limma')
plot_heatmap(object)
```

plot_matrix	<i>Plot binary matrix</i>
-------------	---------------------------

Description

Plot binary matrix

Usage

```
plot_matrix(mat)
```

Arguments

mat matrix

Value

no return (base R plot)

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
mat <- sdt(object)[, .(Subject, subgroup)]
mat$present <- 1
mat %<>% data.table::dcast(Subject ~ subgroup, value.var = 'present', fill = 0)
mat %<>% dt2mat()
plot_matrix(mat)
```

plot_sample_nas	<i>Plot (summarized) detections</i>
-----------------	-------------------------------------

Description

plot_detections plots the detection structure at feature/sample resolution. It shows systematic/random NAs (white), full detection (bright color) and imputations (light color).

Usage

```
plot_sample_nas(...)

plot_subgroup_nas(...)

plot_detections(
  object,
  by = "subgroup",
  fill = by,
  palette = make_svar_palette(object, fill),
  axis.text.y = element_blank()
)

plot_summarized_detections(
  object,
  by = "subgroup",
  fill = by,
  palette = NULL,
  na_imputes = TRUE
)
```

Arguments

...	used to maintain deprecated functions
object	SummarizedExperiment
by	svar (string)
fill	svar (string)
palette	color vector (names = levels, values = colors)
axis.text.y	passed to ggplot2::theme
na_imputes	TRUE or FALSE

Details

plot_summarized_detections plots the detection structure at featuregroup/samplegroup resolution. It shows full detection and random NAs (bright color) and imputations (light color).

Value

ggplot object

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
plot_detections(object)
plot_detections(impute(object))
plot_summarized_detections(object)
plot_summarized_detections(impute(object))

subgroups <- sprintf('%s_STD', c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00'))
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, subgroups = subgroups)
plot_summarized_detections(object)
plot_summarized_detections(object, 'subgroup')
plot_detections(object)
plot_detections(object, 'subgroup')
```

plot_subgroup_points *Plot features*

Description

Plot features

Usage

```
plot_subgroup_points(
  object,
  subgroup = "subgroup",
  block = NULL,
  x = subgroup,
  color = subgroup,
  group = block,
  facet = "feature_id",
  nrow = NULL,
  scales = "free_y",
  ...,
  palette = NULL,
  fixed = list(na.rm = TRUE),
  theme = list(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust = 1))
)
```

Arguments

object	SummarizedExperiment
subgroup	subgroup svar
block	block svar
x	svar mapped to x
color	svar mapped to color
group	svar mapped to group
facet	svar mapped to facets
nrow	number of rows
scales	'free_y' etc.
...	mapped aesthetics
palette	color palette (named character vector)
fixed	fixed aesthetics
theme	ggplot theme specifications

Value

ggplot object

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file, fit = 'limma')
idx <- order(fdata(object)$`p~t1-t0~limma`)[1:9]
object %<>% extract(idx, )
plot_sample_boxplots( object)
plot_feature_boxplots( object)
plot_sample_boxplots(object, x = 'Time')
plot_subgroup_points( object, subgroup = 'Time')
plot_subgroup_points( object, subgroup = 'Time', block = 'Subject')
```

plot_summary	<i>Plot summary</i>
--------------	---------------------

Description

Plot summary

Usage

```
plot_summary(  
  object,  
  fit = "limma",  
  formula = default_formula(object),  
  block = NULL,  
  label = "feature_id",  
  palette = make_svar_palette(object, svar = svar)  
)
```

Arguments

object	SummarizedExperiment
fit	linmod engine : 'limma', 'lm', 'lme', 'lmer' or 'wilcoxon'
formula	model formula
block	NULL or svar
label	fvar
palette	NULL or colorvector

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')  
object <- read_metabolon(file)  
object %<>% pca()  
object %<>% pls(by = 'subgroup')  
object %<>% linmod_limma()  
plot_summary(object, block = 'Subject')
```

plot_venn	<i>Plot venn</i>
-----------	------------------

Description

Plot venn

Usage

```
plot_venn(x)
```

Arguments

x	list
---	------

Examples

```
x <- list(roundfruit = c('apple', 'orange'), redfruit = c('apple', 'strawberry'))  
plot_venn(x)
```

<code>plot_venn_heatmap</code>	<i>Plot venn heatmap</i>
--------------------------------	--------------------------

Description

Plot venn heatmap

Usage

```
plot_venn_heatmap(x)
```

Arguments

<code>x</code>	list
----------------	------

Examples

```
x <- list(roundfruit = c('apple', 'orange'), redfruit = c('apple', 'strawberry'))
plot_venn_heatmap(x)
```

<code>plot_violins</code>	<i>Plot sample/feature violins</i>
---------------------------	------------------------------------

Description

Plot sample/feature violins

Usage

```
plot_violins(
  object,
  assay = assayNames(object)[1],
  x,
  fill,
  color = NULL,
  group = NULL,
  facet = NULL,
  nrow = NULL,
  ncol = NULL,
  dir = "h",
  scales = "free",
  labeller = label_value,
  highlight = NULL,
  palette = NULL,
  fixed = list(na.rm = TRUE)
)
```

```
plot_feature_violins(  
  object,  
  assay = assayNames(object)[1],  
  x = "feature_id",  
  fill = "feature_id",  
  color = NULL,  
  n = 9,  
  facet = NULL,  
  nrow = NULL,  
  ncol = NULL,  
  dir = "h",  
  scales = "free",  
  labeller = label_value,  
  highlight = NULL,  
  fixed = list(na.rm = TRUE)  
)  
  
plot_sample_violins(  
  object,  
  assay = assayNames(object)[1],  
  x = "sample_id",  
  fill = if ("subgroup" %in% svars(object)) "subgroup" else "sample_id",  
  color = NULL,  
  n = 100,  
  facet = NULL,  
  nrow = NULL,  
  ncol = NULL,  
  dir = "h",  
  scales = "free",  
  labeller = label_value,  
  highlight = NULL,  
  fixed = list(na.rm = TRUE)  
)  
  
plot_subgroup_violins(  
  object,  
  assay = assayNames(object)[1],  
  subgroup,  
  x = "subgroup",  
  fill = "subgroup",  
  color = NULL,  
  highlight = NULL,  
  facet = "feature_id",  
  fixed = list(na.rm = TRUE)  
)
```

Arguments

object	SummarizedExperiment
assay	string
x	svar (string)
fill	svar (string)
color	svar (string)
group	svar (string)
facet	svar (character vector)
nrow	NULL or number
ncol	NULL or number
dir	'h' or 'v' : are facets filled horizontally or vertically ?
scales	'free', 'free_x', 'free_y', or 'fixed'
labeller	label_both or label_value
highlight	fvar expressing which feature should be highlighted (string)
palette	named color vector (character vector)
fixed	fixed aesthetics
n	number
subgroup	subgroup svar

Value

ggplot object

See Also

[plot_exprs](#), [plot_densities](#)

Examples

```
# data
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% extract(, order(.$subgroup))
control_features <- c('biotin', 'phosphate')
fdata(object) %<>% cbind(control = .$feature_name %in% control_features)

# plot
plot_violins(object[1:12, ], x = 'feature_id', fill = 'feature_id')
plot_feature_violins(object[1:12, ])
plot_sample_violins(object[, 1:12], highlight = 'control')
plot_subgroup_violins(object[1:4, ], subgroup = 'subgroup')
```

plot_volcano	<i>Plot volcano</i>
--------------	---------------------

Description

Plot volcano

Usage

```
plot_volcano(
  object,
  fit = fits(object)[1],
  coefs = autonomics::coefs(object, fit = fit)[1],
  facet = if (is_scalar(fit)) "coef" else c("fit", "coef"),
  scales = "fixed",
  shape = if ("imputed" %in% fvars(object)) "imputed" else NULL,
  size = NULL,
  alpha = NULL,
  label = if ("gene" %in% fvars(object)) "gene" else "feature_id",
  colors = c(down = "#ff5050", unchanged = "grey", up = "#009933"),
  max.overlaps = 10,
  features = NULL,
  nrow = length(fit),
  p = 0.05,
  fdr = 0.05,
  n = Inf,
  xndown = NULL,
  xnup = NULL,
  title = NULL,
  file = NULL,
  width = 7,
  height = 7,
  verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
fit	'limma', 'lme', 'lm', 'wilcoxon'
coefs	character vector
facet	character vector
scales	'free', 'fixed', etc.
shape	fvar (string)
size	fvar (string)
alpha	fvar (string)

label	fvar (string)
colors	character vector
max.overlaps	number: passed to ggrepel
features	feature ids (character vector): features to encircle
nrow	number: no of rows in plot
p	number: p cutoff for labeling
fdr	number: fdr cutoff for labeling
n	number: n cutoff for labeling
xndown	x position of ndown labels
xnup	x position of nup labels
title	string or NULL
file	filename
width	number
height	number
verbose	TRUE or FALSE

Value

ggplot object

Examples

```
# Regular Usage
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma()
object %<>% linmod_lm()
plot_volcano(object, coefs = 't3-t0', fit = 'limma') # single contrast
plot_volcano(object, coefs = c('t2-t0', 't3-t0'), fit = 'limma') # multip contrasts
plot_volcano(object, coefs = c('t2-t0', 't3-t0'), fit = c('limma', 'lm')) # multip contrs & methods

# When nothing passes FDR
fdr(object) %<>% add_adjusted_pvalues('fdr', fit = 'limma', coefs = 't3-t0')
object %<>% extract( fdrvec(object, fit = 'limma', coef = 't3-t0') > 0.05, )
plot_volcano(object, coefs = 't3-t0', fit = 'limma')

# Additional mappings
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, impute = TRUE)
object %<>% linmod_limma()
plot_volcano(object)
plot_volcano(object, label = 'gene')
plot_volcano(object, label = 'gene', size = 'log2maxlfq')
plot_volcano(object, label = 'gene', size = 'log2maxlfq', alpha = 'pepcounts')
plot_volcano(object, label = 'gene', features = c('Q503D2_DANRE'))
plot_volcano(object, label = 'gene', features = list(c('Q503D2_DANRE', 'Q6DGK4_DANRE'),
c('Q6DGK4_DANRE', 'F1Q7L0_DANRE'))))
```

plot_x_density	<i>Plot xy densities</i>
----------------	--------------------------

Description

Plot xy densities

Usage

```
plot_x_density(
  x,
  y = NULL,
  xbreaks = mclust_breaks(x),
  components = TRUE,
  title = NULL,
  color = "#F8766D",
  xlab = NULL,
  ylab = "Density",
  transcolor = "00000000",
  panel.border = element_rect(color = color),
  plot.margin = unit(c(5.5, 5.5, 5.5, 5.5), "points"),
  scale_x_position = "bottom",
  axis.ticks.x = element_line(color = color),
  axis.ticks.y = element_line(color = color),
  axis.text.x = element_text(color = color),
  axis.text.y = element_text(color = color),
  axis.title.y = element_text(color = color)
)
```

```
plot_y_density(
  y,
  x = NULL,
  ybreaks = mclust_breaks(y),
  title = NULL,
  color = "#F8766D",
  xlab = NULL,
  ylab = NULL,
  transcolor = "00000000"
)
```

```
plot_xy_scatter(
  x,
  y,
  xbreaks = mclust_breaks(x),
  ybreaks = mclust_breaks(y),
  color = c("#F8766D", "#00BFC4"),
  contour = FALSE,
```

```

    smooth = FALSE,
    xlab = NULL,
    ylab = NULL
  )

  plot_xy_density(
    x,
    y,
    xbreaks = mclust_breaks(x),
    ybreaks = mclust_breaks(y),
    xlab = get_name_in_parent(x),
    ylab = get_name_in_parent(y),
    color = c("#F8766D", "#00BFC4"),
    contour = FALSE,
    smooth = FALSE
  )

```

Arguments

<code>x</code>	numeric vector
<code>y</code>	numeric vector
<code>xbreaks</code>	numeric vector
<code>components</code>	TRUE or FALSE: whether to plot distributions of mixture components
<code>title</code>	NULL or string
<code>color</code>	vector or string
<code>xlab</code>	NULL or string
<code>ylab</code>	NULL or string
<code>transcolor</code>	string
<code>panel.border</code>	<code>element_rect(color = color)</code> etc.
<code>plot.margin</code>	<code>unit(c(5.5,5.5,5.5,5.5), 'points')</code> etc.
<code>scale_x_position</code>	'bottom' etc.
<code>axis.ticks.x</code>	<code>element_line(color = color)</code> etc.
<code>axis.ticks.y</code>	<code>element_line(color = color)</code> etc.
<code>axis.text.x</code>	<code>element_text(color = color)</code> etc.
<code>axis.text.y</code>	<code>element_text(color = color)</code> etc.
<code>axis.title.y</code>	<code>element_text(color = color)</code> etc.
<code>ybreaks</code>	numeric vector
<code>contour</code>	TRUE or FALSE: plot density contours ?
<code>smooth</code>	TRUE or FALSE: plot smooth line ?

Value

ggplot

Examples

```
# Bimodal
set.seed(1)
x <- c(rnorm(10, 3), rnorm(10,7))
y <- c(rnorm(10, 3), rnorm(10,7))
plot_xy_density(x,y)
plot_xy_density(x,y, contour = TRUE)
plot_xy_density(x,y, smooth = TRUE)
plot_xy_scatter(x,y)
plot_x_density(x)
plot_y_density(y)

# Unimodal
set.seed(1)
x <- c(rnorm(20, 3))
y <- c(rnorm(20, 3))
plot_xy_density(x,y)
plot_xy_scatter(x,y)
plot_x_density(x)
plot_y_density(y)
```

PRECURSOR_QUANTITY	<i>diann precursor quantity</i>
--------------------	---------------------------------

Description

diann precursor quantity

Usage

PRECURSOR_QUANTITY

Format

An object of class character of length 1.

preprocess_rnaseq_counts
<i>Preprocess RNAseq counts</i>

Description

Preprocess RNAseq counts

Usage

```
preprocess_rnaseq_counts(
  object,
  formula = ~subgroup,
  block = NULL,
  min_count = 10,
  pseudo = 0.5,
  tpm = FALSE,
  cpm = TRUE,
  voom = TRUE,
  log2 = TRUE,
  verbose = TRUE,
  plot = TRUE
)
```

Arguments

<code>object</code>	SummarizedExperiment
<code>formula</code>	designmat formula
<code>block</code>	block svar
<code>min_count</code>	min count required in some samples
<code>pseudo</code>	added pseudocount to avoid $\log(x)=-\text{Inf}$
<code>tpm</code>	TRUE or FALSE : tpm normalize?
<code>cpm</code>	TRUE or FALSE : cpm normalize? (counts per million (scaled) reads)
<code>voom</code>	TRUE or FALSE : voom weight?
<code>log2</code>	TRUE or FALSE : log2 transform?
<code>verbose</code>	TRUE or FALSE : msg?
<code>plot</code>	TRUE or FALSE : plot?

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- .read_rnaseq_counts(file)
object$subgroup
object %<>% preprocess_rnaseq_counts()
```

pull_columns	<i>Pull columns in a dataframe to the front</i>
--------------	---

Description

Pull columns in a dataframe to the front

Usage

```
pull_columns(df, first_cols, verbose = TRUE)
```

Arguments

df	data.frame
first_cols	character vector: columns to be pulled to the front
verbose	TRUE (default) or FALSE

Value

dataframe with re-ordered columns

Examples

```
df <- data.frame(
  symbol = c('A1BG', 'A2M'),
  id      = c('1',   '2'),
  name    = c('alpha-1-B glycoprotein', 'alpha-2-macroglobulin'),
  type    = c('proteinencoding', 'proteinencoding'))
first_cols <- c('id', 'symbol', 'location', 'uniprot')
pull_columns(df, first_cols)
```

pvalues_estimable	<i>Are coefs/pvalues estimable</i>
-------------------	------------------------------------

Description

Are coefs/pvalues estimable

Usage

```
pvalues_estimable(formula, data)

coefs_estimable(formula, data)
```

Arguments

formula	formula
data	data.table

Examples

```
# Onevar design
# -----
# Design not full rank, coefficients/pvalues not estimable
(dt <- data.table( time = factor(c('t0', 't1', 't2', 't3' )),
  value =      c( 0, 1, 2, NA  )),
  coefs_estimable(~time, data = dt)
  pvalues_estimable(~time, data = dt)
  summary(lm(value~time, data = dt))

# Design full rank, coefficients estimable.
# No residual dof, pvalues not estimable.
(dt <- data.table( time = factor(c('t0', 't1', 't2', 't3' )),
  value =      c( 0, 1, 2, 3  )),
  coefs_estimable(~time, data = dt)
  pvalues_estimable(~time, data = dt)
  summary(lm(value~time, data = dt))

# Design full rank, coefficients estimable
# Residual dof, pvalues estimable
(dt <- data.table( time = factor(c('t0', 't1', 't2', 't3', 't3' )),
  value =      c( 0, 1, 2, 3, 3.1 )),
  coefs_estimable(~time, data = dt)
  pvalues_estimable(~time, data = dt)
  summary(lm(value~time, data = dt))

# Twovar design
# -----
# Design not full rank, coefficients/pvalues not estimable.
(dt <- data.table( time = factor(c( 't0', 't1', 't2', 't2', 't3', 't3', 't0', 't1', 't2', 't3' )),
  diabetes = factor(c( 'C', 'C', 'C', 'C', 'C', 'C', 'D', 'D', 'D', 'D' )),
  value =      c( 0, 1, 2, 2.1, 3, 3.1, NA, NA, NA, NA )),
  coefs_estimable(~time+diabetes, data = dt)
  pvalues_estimable(~time+diabetes, data = dt)
  # summary(lm(value~time+diabetes, data = dt))

# Design full rank, coefficients estimable
# No residual dof, pvalues not estimable
(dt <- data.table( time = factor(c( 't0', 't1', 't2', 't3', 't0', 't1', 't2', 't3' )),
  diabetes = factor(c( 'C', 'C', 'C', 'C', 'D', 'D', 'D', 'D' )),
  value =      c( 0, 1, 2, 3, 0.5, NA, NA, NA )),
  coefs_estimable(~time+diabetes, data = dt)
  pvalues_estimable(~time+diabetes, data = dt)
  summary(lm(value~time+diabetes, data = dt))

# Design full rank, coefficients estimable
# Residual dof, pvalues estimable
```

```
(dt <- data.table( time = factor(c( 't0', 't1', 't2', 't3', 't0', 't1', 't2', 't3' )),
  diabetes = factor(c( 'C', 'C', 'C', 'C', 'D', 'D', 'D', 'D' )),
  value =      c(  0,  1,  2,  3,  0.5, 1.6, NA,  NA  )))
  coefs_estimable(~time+diabetes, data = dt)
  pvalues_estimable(~time+diabetes, data = dt)
  summary(lm(value~time+diabetes, data = dt))
```

read_affymetrix	<i>Read affymetrix microarray</i>
-----------------	-----------------------------------

Description

Read affymetrix microarray

Usage

```
read_affymetrix(celfiles)
```

Arguments

celfiles string vector: CEL file paths

Value

RangedSummarizedExperiment

Examples

```
# Downloading example dataset fails 600s limit - example outcommented.
# url <- paste0('http://www.bioconductor.org/help/publications/2003/Chiaretti/chiaretti2/T33.tgz')
# localdir <- file.path(tools::R_user_dir('autonomics', 'cache'), 'T33')
# dir.create(localdir, showWarnings = FALSE)
# localfile <- file.path(localdir, basename(url))
# if (!file.exists(localfile)){ download.file(url, destfile = localfile)
#                               untar(localfile, exdir = path.expand(localdir)) }
# localfile %<>% substr(1, nchar(.)-4)
# if (!installed("BiocManager")) install.packages('BiocManager')
# if (!installed("hgu95av2.db")) BiocManager::install('hgu95av2.db')
# read_affymetrix(celfiles = list.files(localfile, full.names = TRUE))
```

read_compounddiscoverer

Read compound discoverer output

Description

Read compound discoverer output

Usage

```
read_compounddiscoverer(
  dir = getwd(),
  files = list.files(path = dir, pattern = "(RP|HILIC).*\\.csv$", full.names = TRUE),
  colname_regex = "^.*(\\d{8}_+)(.*)+((HILIC|RP)(NEG|POS))\\.raw.*$",
  colname_format = function(x) stringi::stri_replace_first_regex(x, colname_regex,
    "$1$2", opts_regex = stringi::stri_opts_regex(case_insensitive = TRUE)),
  mod_extract = function(x) stringi::stri_subset_regex(x, colname_regex, opts_regex =
    stringi::stri_opts_regex(case_insensitive = TRUE)) %>%
    stringi::stri_replace_first_regex(colname_regex, "$3", opts_regex =
    stringi::stri_opts_regex(case_insensitive = TRUE)),
  quantity = NULL,
  nonames = FALSE,
  exclude_sname_pattern = "(blank|QC|RS)",
  subgroups = NULL,
  logbase = 2,
  impute = FALSE,
  plot = FALSE,
  label = "feature_id",
  pca = plot,
  pls = plot,
  fit = if (plot) "limma" else NULL,
  formula = ~subgroup,
  block = NULL,
  coefs = NULL,
  contrasts = NULL,
  palette = NULL,
  verbose = TRUE
)
```

Arguments

dir	compound discoverer output directory
files	compound discoverer output files
colname_regex	regular expression to parse sample names from column names
colname_format	function to reformat column names
mod_extract	function to extract MS modi from sample names

quantity	'area', 'normalizedarea' or NULL
nonames	TRUE or FALSE: retain compounds without Names?
exclude_sname_pattern	regular expression of sample names to exclude
subgroups	NULL or string vector : subgroups to retain
logbase	base for logarithmization of the data
impute	TRUE or FALSE: impute group-specific NA values?
plot	TRUE or FALSE: plot ?
label	fvar
pca	TRUE or FALSE: run pca ?
pls	TRUE or FALSE: run pls ?
fit	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
formula	model formula
block	model blockvar: string or NULL
coefs	model coefficients of interest: character vector or NULL
contrasts	coefficient contrasts of interest: character vector or NULL
palette	color palette : named character vector
verbose	TRUE or FALSE : message ?

Value

SummarizedExperiment

read_diann_pgmatrix *Read diann phosphosites*

Description

Read diann phosphosites

Usage

read_diann_pgmatrix(dir)

read_diann_phosphosites(dir)

read_diann_phosphodiffs(dir)

Arguments

dir directory with 'report_pgmatrix' and 'report.phosphosites_90.tsv'

Value

SummarizedExperiment

read_fragpipe	<i>Read fragpipe</i>
---------------	----------------------

Description

Read fragpipe

Usage

```
read_fragpipe(  
  dir = getwd(),  
  file = if (is_file(dir)) dir else file.path(dir, "combined_protein.tsv"),  
  contaminants = FALSE,  
  verbose = TRUE  
)
```

Arguments

dir	directory with 'combined_protein.tsv'
file	'combined_protein.tsv' (full path)
contaminants	whether to include contaminants
verbose	whether to msg

Value

SummarizedExperiment

Examples

```
file <- download_data('multiorganism.combined_protein.tsv')  
object <- read_fragpipe(file = file)  
object  
fdt(object)  
sdt(object)
```

read_maxquant_phosphosites	<i>Read maxquant phosphosites</i>
----------------------------	-----------------------------------

Description

Read maxquant phosphosites

Usage

```

read_maxquant_phosphosites(
  dir = getwd(),
  fosfile = if (is_file(dir)) dir else file.path(dir, "phospho (STY)Sites.txt"),
  profile = file.path(dirname(fosfile), "proteinGroups.txt"),
  fastafile = NULL,
  restapi = FALSE,
  quantity = NULL,
  subgroups = NULL,
  invert = character(0),
  rm_contaminants = TRUE,
  rm_reverse = TRUE,
  rm_missing_in_all_samples = TRUE,
  localization = 0.75,
  impute = FALSE,
  plot = FALSE,
  label = "feature_id",
  pca = plot,
  pls = plot,
  fit = if (plot) "limma" else NULL,
  formula = as.formula("~ subgroup"),
  block = NULL,
  coefs = NULL,
  contrasts = NULL,
  palette = NULL,
  verbose = TRUE
)

read_phosphosites(...)

```

Arguments

dir	proteingroups directory
fosfile	phosphosites file
profile	proteingroups file
fastafile	uniprot fastafile
restapi	TRUE or FALSE : annotate non-fastadt uniprot using uniprot restapi
quantity	'normalizedratio', 'ratio', 'correctedreporterintensity', 'reporterintensity', 'maxlfq', 'labeledintensity', 'intensity' or NULL
subgroups	NULL or string vector : subgroups to retain
invert	string vector: subgroups which require inversion
rm_contaminants	TRUE or FALSE: rm contaminants ?
rm_reverse	TRUE or FALSE: rm reverse proteins ?
rm_missing_in_all_samples	TRUE or FALSE

localization	number: min localization probability (for phosphosites)
impute	TRUE or FALSE: impute group-specific NA values?
plot	TRUE or FALSE
label	fvar
pca	TRUE or FALSE: run pca ?
pls	TRUE or FALSE: run pls ?
fit	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
formula	model formula
block	model blockvar: string or NULL
coefs	model coefficients of interest: string vector or NULL
contrasts	model coefficient contrasts of interest: string vector or NULL
palette	color palette: named string vector
verbose	TRUE or FALSE: message ?
...	maintain deprecated functions

Value

SummarizedExperiment

Examples

```
profile <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
fosfile <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics')
fastafile <- system.file('extdata/uniprot_hsa_20140515.fasta', package = 'autonomics')
subgroups <- sprintf('%s_STD', c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00'))
pro <- read_maxquant_proteingroups(file = profile, subgroups = subgroups)
fos <- read_maxquant_phosphosites(fosfile = fosfile, profile = profile, subgroups = subgroups)
fos <- read_maxquant_phosphosites(fosfile = fosfile, profile = profile, fastafile = fastafile, subgroups = subgroups)
```

read_maxquant_proteingroups
<i>Read maxquant proteingroups</i>

Description

Read maxquant proteingroups

Usage

```

read_maxquant_proteingroups(
  dir = getwd(),
  file = if (is_file(dir)) dir else file.path(dir, "proteinGroups.txt"),
  fastafile = NULL,
  restapi = FALSE,
  quantity = NULL,
  subgroups = NULL,
  invert = character(0),
  rm_contaminants = TRUE,
  rm_reverse = TRUE,
  rm_missing_in_all_samples = TRUE,
  impute = FALSE,
  plot = FALSE,
  label = "feature_id",
  pca = plot,
  pls = plot,
  fit = if (plot) "limma" else NULL,
  formula = as.formula("~ subgroup"),
  block = NULL,
  coefs = NULL,
  contrasts = NULL,
  palette = NULL,
  verbose = TRUE
)

read_proteingroups(...)

```

Arguments

dir	proteingroups directory
file	proteingroups file
fastafile	uniprot fastafile
restapi	TRUE or FALSE : use uniprot restapi to annotate uniprot not in fastadt ?
quantity	'normalizedratio', 'ratio', 'correctedreporterintensity', 'reporterintensity', 'maxlfq', 'labeledintensity', 'intensity' or NULL
subgroups	NULL or string vector : subgroups to retain
invert	string vector : subgroups which require inversion
rm_contaminants	TRUE or FALSE : rm contaminants ?
rm_reverse	TRUE or FALSE : rm reverse proteins ?
rm_missing_in_all_samples	TRUE or FALSE
impute	TRUE or FALSE: impute group-specific NA values?
plot	TRUE or FALSE: plot ?

label	fvar
pca	TRUE or FALSE: run pca ?
pls	TRUE or FALSE: run pls ?
fit	model engine: 'limma', 'lm', 'lme(r)', 'wilcoxon' or NULL
formula	model formula
block	model blockvar: string or NULL
coefs	model coefficients of interest: character vector or NULL
contrasts	coefficient contrasts of interest: character vector or NULL
palette	color palette : named character vector
verbose	TRUE or FALSE : message ?
...	maintain deprecated functions

Value

SummarizedExperiment

Examples

```
# fukuda20 - LFQ
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
pro <- read_maxquant_proteingroups(file = file)

# billing19 - Normalized Ratios
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
fastafile <- system.file('extdata/uniprot_hsa_20140515.fasta', package = 'autonomics')
subgroups <- sprintf('%s_STD', c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00'))
pro <- read_maxquant_proteingroups(file = file, subgroups = subgroups)
pro <- read_maxquant_proteingroups(file = file, fastafile = fastafile, subgroups = subgroups)
```

read_msigdt	<i>Read msigdb datatable</i>
-------------	------------------------------

Description

Read msigdb datatable

Usage

```
read_msigdt(
  file = defaultmsigfile(),
  collections = if (is.null(file)) NULL else switch(basename(file) %>% substr(nchar(.)
    - 4, nchar(.) - 3), Hs = c("C2:CP:REACTOME", "C5:GO:BP", "C5:GO:MF", "C5:GO:CC"), Mm
    = c("M2:CP:REACTOME", "M5:GO:BP", "M5:GO:MF", "M5:GO:CC"))
)
```

Arguments

file msigdb file: one of the files in dir(MSIGDB).
collections subset of names(MSIGCOLLECTIONS)

Examples

```
read_msigdt()
```

read_olink	<i>Read olink file</i>
------------	------------------------

Description

Read olink file

Usage

```
read_olink(file, sample_excel = NULL, sample_tsv = NULL, by.y = "SampleID")
```

Arguments

file olinkfile
sample_excel sample excel
sample_tsv sample tsv
by.y sample tsv mergeby column

Value

SummarizedExperiment

Examples

```
# Example data
npxdt <- data.table::data.table(OlinkAnalyze::npx_data1)[, c(1:11, 17)]
sampledt <- data.table::data.table(OlinkAnalyze::npx_data1)[, c(1, 12:15)]
sampledt %<>% extract(!grepl('CONTROL', SampleID))
sampledt %<>% unique()

# Write to file
file <- paste0(tempfile(), '.olink.csv')
samplefile <- paste0(tempfile(), '.samples.xlsx')
data.table::fwrite(npxdt, file)
writexl::write_xlsx(sampledt, samplefile)

# Read
object <- read_olink(file, sample_excel = samplefile)
biplot(pca(object), color = 'Time', group = 'Subject', shape = 'Treatment')
```

read_salmon	<i>Read salmon</i>
-------------	--------------------

Description

Read salmon

Usage

```
read_salmon(dir, sfile = NULL, by = NULL, ensdb = NULL)
```

Arguments

dir	salmon results rootdir
sfile	samplefile
by	samplefile column to merge by
ensdb	EnsDb object

Value

SummarizedExperiment

Examples

```
# dir <- '../bh/salmon_quants'
# sfile <- '../bh/samplesheet.csv'
# by <- 'salmonDir'
# ah <- AnnotationHub::AnnotationHub()
# ensdb <- ah[['AH98078']]
# read_salmon(dir, sfile = sfile, by = 'salmonDir', ensdb = ensdb)
```

read_uniprot	<i>Read fasta hdrs</i>
--------------	------------------------

Description

Read fasta hdrs

Usage

```
read_uniprot(fastafile, fastafields = FASTAFIELDS, verbose = TRUE)

parse_maxquant_hdrs(fastahdrs)

read_contaminantdt(force = FALSE, verbose = TRUE)
```

Arguments

fastafile	string (or charactervector)
fastafields	charactervector : which fastahdr fields to extract ?
verbose	bool
fastahdrs	character vector
force	whether to overwrite existing file

Value

data.table(uniprot, protein, gene, uniprot, reviewed, existence)

Note

existence values are always those of the canonical isoform (no isoform-level resolution for this field)

Examples

```
# uniprot hdrs
fastafile <- system.file('extdata/uniprot_hsa_20140515.fasta', package = 'autonomics')
read_uniprot(dt, fastafile)

# maxquant hdrs
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
dt <- .read_maxquant_proteingroups(file)
parse_maxquant_hdrs(dt$`Fasta headers`)

profile <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
fosfile <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics')
prodt <- .read_maxquant_proteingroups(profile)
fosdt <- .read_maxquant_phosphosites(fosfile, profile)
parse_maxquant_hdrs(prodt$`Fasta headers`)
parse_maxquant_hdrs(fosdt$`Fasta headers`)

# contaminant hdrs
read_contaminant(dt)
```

reexports	<i>Objects exported from other packages</i>
-----------	---

Description

These objects are imported from other packages. Follow the links below to see their documentation.

- data.table** [data.table](#)
- magrittr** [%<>%](#), [%>%](#), [extract](#)

reset_fit	<i>Reset fit</i>
-----------	------------------

Description

Reset fit

Usage

```
reset_fit(object, fit = fits(object), verbose = TRUE)
```

Arguments

object	SummarizedExperiment
fit	character vector
verbose	TRUE or FALSE

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %>% fdt()
object %>% linmod_limma() %>% fdt()
object %>% linmod_limma() %>% reset_fit() %>% fdt()
object %>% linmod_limma() %>% linmod_lm() %>% reset_fit('limma') %>% fdt()
object %>% linmod_limma() %>% linmod_lm() %>% reset_fit() %>% fdt()
```

rm_diann_contaminants	<i>Rm contaminants</i>
-----------------------	------------------------

Description

Rm contaminants from DIA-NN SumExp

Usage

```
rm_diann_contaminants(object, verbose = TRUE)
```

Arguments

object	SummarizedExperiment
verbose	TRUE or FALSE

Value

SummarizedExperiment

Examples

```
file <- download_data('dilution.report.tsv')
object <- read_diann_proteingroups(file)
object %<>% rm_diann_contaminants()
```

`rm_missing_in_all_samples`*Rm features missing in some samples*

Description

Rm features missing in some samples

Usage

```
rm_missing_in_all_samples(object, verbose = TRUE)

rm_missing_in_some_samples(object, verbose = TRUE)
```

Arguments

object	SummarizedExperiment
verbose	TRUE (default) or FALSE

Value

updated object

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
rm_missing_in_all_samples( object)
rm_missing_in_some_samples(object)
```

`rm_unmatched_samples` *rm unmatched/singleton samples*

Description

rm unmatched/singleton samples

Usage

```
rm_unmatched_samples(  
  object,  
  subgroupvar = "subgroup",  
  subgroupctr = slevels(object, subgroupvar)[1],  
  block,  
  verbose = TRUE  
)  
  
rm_singleton_samples(object, subgroupvar = "subgroup", verbose = TRUE)
```

Arguments

object	SummarizedExperiment
subgroupvar	subgroup variable (string)
subgroupctr	control subgroup (string)
block	block variable (string)
verbose	TRUE/FALSE

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.somascan.adat', package = 'autonomics')  
object <- read_somascan(file)  
object %<>% filter_samples(subgroup %in% c('t1', 't2'), verbose = TRUE)  
rm_singleton_samples(object, subgroupvar = 'Subject')  
rm_unmatched_samples(object, subgroupvar = 'subgroup', block = 'Subject')
```

sbind	<i>Sample/Feature/Assay bind</i>
-------	----------------------------------

Description

Sample/Feature/Assay bind

Usage

```
sbind(obj1, obj2)  
  
fbind(obj1, obj2)  
  
abind(obj1, obj2)
```

Arguments

obj1 SummarizedExperiment: nrow1 x ncol1
 obj2 SummarizedExperiment: nrow2 x ncol2

Value

SummarizedExperiment: nrow1+nrow2 x ncol1+ncol2

Examples

```
# Data
obj1 <- object1()
obj2 <- object2()
biplot( pca(obj1), color = 'age')
biplot( pca(obj2), color = 'age')

# Sample bind
obj <- sbind(obj1, obj2)
biplot( pca(obj), color = 'age', shape = 'set')
sdt(obj) # SET added
fdt(obj) # common fvars with differing content pasted together

# Feature bind
obj <- fbind(obj1, obj2)
biplot( pca(obj), color = 'age', nx = 2)
fdt(obj) # SET added
sdt(obj) # common svars with differing content pasted together

# Assay bind
obj <- abind(obj1, obj2)
plot( SummarizedExperiment::assays(abind(obj1, obj2))$SET1.exprs,
      SummarizedExperiment::assays(abind(obj1, obj2))$SET2.exprs)
fdt(obj) # common fvars with differing content pasted together
sdt(obj) # common svars with differing content pasted together
```

scaledlibsizes

Get tmm-scaled libsizes

Description

Get tmm-scaled libsizes

Usage

```
scaledlibsizes(counts)
```

Arguments

counts counts matrix

Value

scaled libsize vector

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file)
scaledlibsizes(counts(object))
```

scoremat	<i>Extract scores/loadings</i>
----------	--------------------------------

Description

Extract scores/loadings

Usage

```
scoremat(object, method = "pca", by = biplot_by(object, method), dim = 1:2)

scores(object, method = "pca", by = biplot_by(object, method), dim = 1)

loadingmat(object, method = "pca", by = biplot_by(object, method), dim = 1:2)

loadings(object, method = "pca", by = biplot_by(object, method), dim = 1)
```

Arguments

object	SummarizedExperiment
method	'pca', 'pls', etc.
by	svar (string)
dim	numeric vector

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% pca()
  scores(object)[1:2]
  loadings(object)[1:2]
  scoremat(object)[1:2, ]
  loadingmat(object)[1:2, ]
```

slevels	<i>Get slevels</i>
---------	--------------------

Description

Get svar levels

Usage

```
slevels(object, svar)

subgroup_levels(object)
```

Arguments

object	SummarizedExperiment, eSet, or eList
svar	sample var (character)

Value

svar values (character)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
slevels(object, 'subgroup')
subgroup_levels(object)
```

snames	<i>Get/Set snames</i>
--------	-----------------------

Description

Get/Set sample names

Usage

```
snames(object)

## S4 method for signature 'SummarizedExperiment'
snames(object)

snames(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,character'
snames(object) <- value
```

Arguments

object	SummarizedExperiment
value	string vector with sample names

Value

sample names vector (get) or updated eSet (set)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
head(snames(object))
head(snames(object) %<>% paste0('SAMPLE_', .))
```

split_samples	<i>Split samples</i>
---------------	----------------------

Description

Split samples by svar

Usage

```
split_samples(object, by = "subgroup")

cbind_imputed(objlist)

split_features(object, by)
```

Arguments

object	SummarizedExperiment
by	svar to split by (string)
objlist	SummarizedExperiment list

Value

SummarizedExperiment list

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
objlist <- split_features(object, by = 'PLATFORM')
objlist <- split_samples(object, 'Diabetes')
objlist %<>% Map(impute, .)
object <- cbind_imputed(objlist)
```

stepauc	<i>Compute step auc</i>
---------	-------------------------

Description

Compute step auc

Usage

```
stepauc(x, y, color = "group1", plot = FALSE)
```

Arguments

x	numeric vector
y	numeric vector
color	string
plot	TRUE or FALSE

Value

number

Examples

```
x <- c( 0, 4, 8, 27)
y <- c(100, 67, 33, 0)
stepauc(x, y, plot = TRUE)
```

stri_any_regex	<i>Does any string have a regex</i>
----------------	-------------------------------------

Description

Does any string have a regex

Usage

```
stri_any_regex(str, pattern)
```

Arguments

str	string vector
pattern	string

Value

TRUE or FALSE

Examples

```
str <- c('s1 Spectral Count', 's1 Unique Spectral Count')
patterns <- c('Spectral Count', '(?!Unique) Spectral Count', 'Intensity')
stringi::stri_detect_regex(str, pattern = patterns[1])
stringi::stri_detect_regex(str, pattern = patterns[2])
stringi::stri_detect_regex(str, pattern = patterns[3])
stri_any_regex(str, pattern = patterns)
```

stri_detect_fixed_in_collapsed

Detect fixed patterns in collapsed strings

Description

Detect fixed patterns in collapsed strings

Usage

```
stri_detect_fixed_in_collapsed(x, patterns, sep)
```

Arguments

x	vector with collapsed strings
patterns	vector with fixed patterns (strings)
sep	collapse separator (string) or NULL (if uncollapsed)

Value

boolean vector

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
x <- fdt(object)$uniprot
patterns <- c('A0A0R4IKT8', 'Q7T3G6')
table(stri_detect_fixed_in_collapsed(x = x, patterns = patterns, sep = ';'))
```

subgroup_array	<i>Get subgroup matrix</i>
----------------	----------------------------

Description

Arrange (subgroup)levels in matrix

Usage

```
subgroup_array(object, subgroupvar)
subgroup_matrix(object, subgroupvar)
```

Arguments

object	SummarizedExperiment
subgroupvar	subgroup svar

Value

matrix

Examples

```
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object$subgroup <- paste0(object$Diabetes, '.', object$subgroup)
subgroup_matrix(object, 'subgroup')
```

subtract_baseline	<i>Subtract baseline</i>
-------------------	--------------------------

Description

Subtract baseline level within block

Usage

```
subtract_baseline(
  object,
  subgroupvar,
  subgroupctr = slevels(object, subgroupvar)[1],
  block = NULL,
  assaynames = setdiff(assayNames(object), c("weights", "pepcounts")),
  verbose = TRUE
)
```

```

subtract_pairs(
  object,
  subgroupvar = "subgroup",
  subgroupctr = slevels(object, subgroupvar)[1],
  block,
  assaynames = assayNames(object)[1],
  verbose = TRUE
)

subtract_differences(object, block, subgroupvar, verbose = TRUE)

```

Arguments

<code>object</code>	SummarizedExperiment
<code>subgroupvar</code>	subgroup svar
<code>subgroupctr</code>	control subgroup
<code>block</code>	block svar (within which subtraction is performed)
<code>assaynames</code>	which assays to subtract for
<code>verbose</code>	TRUE/FALSE

Details

`subtract_baseline` subtracts baseline levels within block, using the medoid baseline sample if multiple exist.

`subtract_pairs` also subtracts baseline level within block. It cannot handle multiple baseline samples, but has instead been optimized for many blocks

`subtract_differences` subtracts differences between subsequent levels, again within block

Value

SummarizedExperiment

Examples

```

# read
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object0 <- read_metabolon(file)
pca(object0, plot = TRUE, color = 'Time')

# subtract_baseline: takes medoid of baseline samples if multiple
object <- subtract_baseline(object0, block = 'Subject', subgroupvar = 'Time')
pca(object, plot = TRUE, color = 'Time')

# subtract_pairs: optimized for many blocks
object <- subtract_pairs(object0, block = 'Subject', subgroupvar = 'Time')

```

```

pca(object, plot = TRUE, color = 'Time')

# subtract_differences
object <- subtract_differences(object0, block = 'Subject', subgroupvar = 'Time')
values(object) %<>% na_to_zero()
pca(object, plot = TRUE, color = 'Time')

```

sumexplist_to_longdt *SummarizedExperiment list to long data.table*

Description

SummarizedExperiment list to long data.table

Usage

```

sumexplist_to_longdt(
  sumexplist,
  svars = intersect("subgroup", autonomics::svars(sumexplist[[1]])),
  fvars = intersect("gene", autonomics::fvars(sumexplist[[1]])),
  setvarname = "set"
)

```

Arguments

sumexplist	list of SummarizedExperiments
svars	character vector
fvars	character vector
setvarname	string

Value

data.table

Examples

```

subgroups <- paste0(c('E00', 'E01', 'E02', 'E05', 'E15', 'E30', 'M00'), '_STD')
rnafile <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
profile <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
fosfile <- system.file('extdata/billing19.phosphosites.txt', package = 'autonomics')
rna <- read_rnaseq_counts(rnafile)
pro <- read_maxquant_proteingroups(file = profile, subgroups = subgroups)
fos <- read_maxquant_phosphosites(fosfile = fosfile, profile = profile, subgroups = subgroups)
pro$subgroup %<>% stringi::stri_replace_first_fixed('_STD', '')
fos$subgroup %<>% stringi::stri_replace_first_fixed('_STD', '')

sumexplist <- list(rna = rna, pro = pro, fos = fos)
dt <- sumexplist_to_longdt(sumexplist, setvarname = 'platform')
dt %<>% extract(gene %in% c('TNMD', 'TSPAN6'))

```

sumexp_to_tsv	<i>Write sumexp to tsv</i>
---------------	----------------------------

Description

Write sumexp to tsv

Usage

```
sumexp_to_tsv(object, assay = assayNames(object)[1], file)
```

Arguments

object	SummarizedExperiment
assay	string
file	filename

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file, fit = 'limma')
tsv <- file.path(tempdir(), 'fukuda20.proteingroups.tsv')
sumexp_to_tsv(object, file = tsv)
```

sumexp_to_widedt	<i>SummarizedExperiment to data.table</i>
------------------	---

Description

SummarizedExperiment to data.table

Usage

```
sumexp_to_widedt(
  object,
  fvars = autonomics::fvars(object),
  assay = assayNames(object)[1]
)

sumexp_to_longdt(
  object,
  fvars = intersect("feature_name", autonomics::fvars(object)),
  svars = intersect("subgroup", autonomics::svars(object)),
  assay = assayNames(object) %>% intersect(c(.[1], "is_imputed")),
  value.name = "value"
)

sumexp_to_groupdt(object, subgroup = subgroup)
```

Arguments

object	sumexp
fvars	additional fvars to include in table
assay	matrix in assays(object) to be used
svars	additional svars to include in table
value.name	string: passed to melt.data.table
subgroup	subgroup (sym)

Details

- sumexp_to_widedt: feature x sample
- sumexp_to_groupdt: feature.subgroup x replicate
- sumexp_to_longdt: feature.sample

Value

data.table

Examples

```
# Atkin Hypoglycemia
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
sumexp_to_widedt(object)
sumexp_to_longdt(object)
sumexp_to_groupdt(object)

# Fukuda
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
values(object)
fdt(object)
object %<>% impute()
table(fdt(object)$imputed)
sumexp_to_longdt(object)
sumexp_to_widedt(object)
sumexp_to_longdt(object)
```

summarize_fit	<i>Summarize fit</i>
---------------	----------------------

Description

Summarize fit

Usage

```

summarize_fit(object, ...)

## S3 method for class 'data.table'
summarize_fit(
  object,
  fit = fits(object),
  coefs = autonomics::coefs(object, fit = fit),
  ...
)

## S3 method for class 'SummarizedExperiment'
summarize_fit(
  object,
  fit = fits(object),
  coefs = autonomics::coefs(object, fit = fit),
  ...
)

```

Arguments

object	SummarizedExperiment or data.table
...	S3 dispatch
fit	'limma', 'lme', 'lm', 'lme', 'wilcoxon' or NULL
coefs	string vector

Value

data.table(contrast, nup, ndown)

Examples

```

file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma()
object %<>% linmod_lm()
summarize_fit(object, coefs = c('t1-t0', 't2-t0', 't3-t0'))

```

survobj

Survival analysis example

Description

Survival analysis example

Usage

```
survobj(verbose = TRUE)
```

Arguments

verbose TRUE or FALSE

Value

SummarizedExperiment

Examples

```
survobj()
```

svalues	<i>Get/Set svalues</i>
---------	------------------------

Description

Get/Set svar values

Usage

```
svalues(object, svar)

subgroup_values(object)

sampleid_values(object)

svalues(object, svar) <- value

## S4 replacement method for signature 'SummarizedExperiment,character'
svalues(object, svar) <- value
```

Arguments

object SummarizedExperiment
svar sample var (character)
value value vector

Value

character vector (get) or SummarizedExperiment (set)

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
svalues(object, 'subgroup')
subgroup_values(object)
```

svars	<i>Get/Set svars</i>
-------	----------------------

Description

Get/Set sample variables

Usage

```
svars(object)

## S4 method for signature 'SummarizedExperiment'
svars(object)

## S4 method for signature 'MultiAssayExperiment'
svars(object)

svars(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,character'
svars(object) <- value

## S4 replacement method for signature 'MultiAssayExperiment,character'
svars(object) <- value
```

Arguments

object	SummarizedExperiment
value	string factor with variable names

Value

sample variable names (get) or updated SummarizedExperiment

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
svars(object)[1]
(svars(object)[1] %<>% paste0('1'))
```

systematic_nas	<i>Is systematic/random/full NA</i>
----------------	-------------------------------------

Description

Is systematic/random/full NA

Usage

```
systematic_nas(object, by = "subgroup", frac = 0.5)
```

```
random_nas(object, by = "subgroup")
```

```
no_nas(object)
```

Arguments

object	SummarizedExperiment
by	svar (string)
frac	fraction

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
table(systematic_nas(object)) # missing in some subgroups, present in others
table(random_nas(object))    # missing in some samples, independent of subgroup
table(no_nas(object))        # missing in no samples
```

tag_features	<i>Tag features</i>
--------------	---------------------

Description

Tag features

Usage

```
tag_features(
  object,
  keyvar,
  sep,
  features,
  tagvar = get_name_in_parent(features),
  verbose = TRUE
)
```

Arguments

object	SummarizedExperiment
keyvar	string : intersection fvar
sep	string : keyvar collapse separator
features	character vector : intersection set
tagvar	string :
verbose	TRUE or FALSE

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/atkin.somascan.adat', package = 'autonomics')
object <- read_somascan(file)
features <- AnnotationDbi::keys(org.Hs.eg.db::org.Hs.eg.db, keytype = 'SYMBOL')
object %<>% tag_features(keyvar = 'EntrezGeneSymbol', sep = ' ', features)
table(fdt(object)$features)
```

tag_hdlproteins	<i>Tag hdlproteins</i>
-----------------	------------------------

Description

Tag hdlproteins

Usage

```
tag_hdlproteins(object, verbose = TRUE)
```

Arguments

object	SummarizedExperiment
verbose	TRUE or FALSE

Value

SummarizedExperiment

Examples

```
file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
object %<>% tag_hdlproteins()
fdt(object)
```

TAXON_TO_ORGNAME	<i>Annotation Maps</i>
------------------	------------------------

Description

Annotation Maps

Usage

TAXON_TO_ORGNAME

ABBREV_TO_ORGNAME

REVIEWED_TO_NUMBER

EXISTENCE_TO_NUMBER

Format

An object of class character of length 7.
An object of class character of length 4.
An object of class character of length 2.
An object of class numeric of length 4.

Examples

```
TAXON_TO_ORGNAME['9606']  
ABBREV_TO_ORGNAME['HSA']  
REVIEWED_TO_NUMBER['reviewed']  
EXISTENCE_TO_NUMBER['Evidence at protein level']
```

TESTS	<i>Statistical models supported in autonomics</i>
-------	---

Description

Statistical models supported in autonomics

Usage

TESTS

Format

An object of class character of length 5.

Examples

TESTS

tpm	<i>Get/Set tpm</i>
-----	--------------------

Description

Get / Set tpm matrix

Usage

```
tpm(object)

## S4 method for signature 'SummarizedExperiment'
tpm(object)

tpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
tpm(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
tpm(object) <- value
```

Arguments

object	SummarizedExperiment
value	tpm matrix (features x samples)

Value

tpm matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')
object <- read_rnaseq_counts(file, plot=FALSE)
tpm(object) <- values(object)
tpm(object)[1:3, 1:3]
```

TRANSFORMENGINES	<i>Data Transformation Methods</i>
------------------	------------------------------------

Description

Data Transformation Methods

Usage

TRANSFORMENGINES

TRANSFORMSTRICT

Format

An object of class character of length 7.

An object of class character of length 5.

Details

- TRANSFORMENGINES: c('center', 'center_mean', 'center_median', 'invnorm', 'quantnorm', 'vsn', 'zscore')
- TRANSFORMSTRICT: c('center', 'invnorm', 'quantnorm', 'vsn', 'zscore')

twofactor_sumexp	<i>twofactor sumexp</i>
------------------	-------------------------

Description

twofactor sumexp

Usage

twofactor_sumexp()

Value

SummarizedExperiment

uncollapse	<i>Uncollapse/Recollapse</i>
------------	------------------------------

Description

Uncollapse data.table cols

Usage

```
uncollapse(dt, ..., sep = ";")  
  
recollapse(dt, by, sep = ";")
```

Arguments

dt	data.table
...	cols
sep	string
by	string

Examples

```
# Example data  
(dt <- data.table::data.table(  
  uniprot = 'Q9BQL6;Q96AC1;Q96AC1-3',  
  protein = 'FERM1_HUMAN;FERM2_HUMAN',  
  gene    = 'FERMT1;FERMT2',  
  family  = 'FERM'))  
  
# Uncollapse  
uncollapse(dt, protein, gene, sep = ';')  
recollapse( uncollapse(dt, protein, gene, sep = ';'), by = 'uniprot')  
  
# Unchanged when no sep  
uncollapse(dt, family, sep = ';')  
uncollapse(dt, family, sep = 'NOSEP')
```

values	<i>Get/Set expr values</i>
--------	----------------------------

Description

Get/Set value matrix

Usage

```

values(object)

## S4 method for signature 'SummarizedExperiment'
values(object)

values(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,matrix'
values(object) <- value

## S4 replacement method for signature 'SummarizedExperiment,numeric'
values(object) <- value

```

Arguments

object	SummarizedExperiment
value	ratio matrix (features x samples)

Value

value matrix (get) or updated object (set)

Examples

```

file <- system.file('extdata/billing19.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
values(object)[1:3, 1:3]
values(object) <- 0
values(object)[1:3, 1:3]

```

varlevels_dont_clash *Are varlevels unique*

Description

Are varlevels unique

Usage

```

varlevels_dont_clash(object, ...)

## S3 method for class 'data.table'
varlevels_dont_clash(object, vars = names(object), ...)

## S3 method for class 'SummarizedExperiment'
varlevels_dont_clash(object, vars = svars(object), ...)

```

Arguments

object	SummarizedExperiment or data.table
...	required for s3 dispatch
vars	character vector

Value

TRUE or FALSE

Examples

```
require(data.table)
object1 <- data.table(expand.grid(genome = c('WT', 'MUT'), treat = c('control', 'drug')))
object2 <- data.table(expand.grid(mutant = c('YES', 'NO'), treated = c('YES', 'NO')))
varlevels_dont_clash(object1)
varlevels_dont_clash(object2)
```

venn_detects	<i>Venn detects</i>
--------------	---------------------

Description

Venn diagram full/consistent/random detects

Usage

```
venn_detects(object, by = "subgroup")
```

Arguments

object	SummarizedExperiment
by	svar (string)

Value

NULL

Examples

```
file <- system.file('extdata/fukuda20.proteingroups.txt', package = 'autonomics')
object <- read_maxquant_proteingroups(file)
venn_detects(object, 'subgroup')
```

weights	<i>Get/Set weights</i>
---------	------------------------

Description

Get/Set weight matrix

Usage

```
weights(object, ...)  
  
## S4 method for signature 'SummarizedExperiment'  
weights(object)  
  
weights(object) <- value  
  
## S4 replacement method for signature 'SummarizedExperiment,matrix'  
weights(object) <- value  
  
## S4 replacement method for signature 'SummarizedExperiment,numeric'  
weights(object) <- value  
  
## S4 replacement method for signature 'SummarizedExperiment,NULL'  
weights(object) <- value
```

Arguments

object	SummarizedExperiment
...	additional params
value	ratio matrix (features x samples)

Value

weight matrix (get) or updated object (set)

Examples

```
file <- system.file('extdata/billing19.rnacounts.txt', package = 'autonomics')  
object <- read_rnaseq_counts(file)  
weights(object)[1:3, 1:2]  
weights(object) <- 1  
weights(object)[1:3, 1:2]
```

write_xl	<i>Write xl</i>
----------	-----------------

Description

Write xl

Usage

```
write_xl(  
  object,  
  file,  
  fitcoefs = autonomics::fitcoefs(object),  
  assays = assayNames(object)[0],  
  verbose = TRUE  
)  
  
write_ods(  
  object,  
  file,  
  fitcoefs = autonomics::fitcoefs(object),  
  assays = assayNames(object)[0],  
  verbose = TRUE  
)
```

Arguments

object	SummarizedExperiment
file	file
fitcoefs	character vector
assays	assayNames subset
verbose	TRUE or FALSE

Value

filepath

Examples

```
# linmod  
file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')  
object <- read_metabolon(file)  
object %<>% linmod_limma(~Diabetes/Time)  
xlfile <- file.path(tempdir(), 'linmod.atkin.metabolon.xlsx')  
odsfile <- file.path(tempdir(), 'linmod.atkin.metabolon.ods' )  
write_xl( object, xlfile) # linmod.xlsx: fdt + stats  
write_xl( object, xlfile, assays = SummarizedExperiment::assayNames(object)[1] ) # fdt + stats + a
```

```
write_xl( object, xlfile, assays = SummarizedExperiment::assayNames(object)[1:2])      # fdt + stats + a
write_ods(object, odsfile)                                                           # ods: fdt + stats
write_ods(object, odsfile, assays = SummarizedExperiment::assayNames(object)[1] )      # fdt + stats + a
write_ods(object, odsfile, assays = SummarizedExperiment::assayNames(object)[1:2])      # fdt + stats +

# awblinmod
object <- read_metabolon(file)
object %<>% awblinmod_limma(c('Diabetes', 'Time'), block = 'Subject')
xlfile <- file.path(tempdir(), 'awblinmod.atkin.metabolon.xlsx')
odsfile <- file.path(tempdir(), 'awblinmod.atkin.metabolon.ods')
write_xl( object, xlfile)                                                           # awblinmod xlsx: fdt + stats
write_xl( object, xlfile, assay = SummarizedExperiment::assayNames(object)[1] )      # fdt + stats +
write_xl( object, xlfile, assay = SummarizedExperiment::assayNames(object)[1:2])      # fdt + stats +
write_ods(object, odsfile)                                                           # ods: fdt + stats
write_ods(object, odsfile, assay = SummarizedExperiment::assayNames(object)[1] )      # fdt + stats +
write_ods(object, odsfile, assay = SummarizedExperiment::assayNames(object)[1:2])      # fdt + stats +
```

X	<i>Model based prediction</i>
---	-------------------------------

Description

Model based prediction

Usage

```
X(
  object,
  formula = default_formula(object),
  drop = varlevels_dont_clash(object, all.vars(formula)),
  coding = "code_control"
)

beta(object, fit = fits(object)[1])
```

Arguments

object	SummarizedExperiment or data.frame
formula	formula
drop	TRUE or FALSE
coding	string: codingfunname
fit	'limma', 'lm', 'lme', 'wilcoxon'

Value

beta matrix (nlevel x nfeature)

Examples

```

file <- system.file('extdata/atkin.metabolon.xlsx', package = 'autonomics')
object <- read_metabolon(file)
object %<>% linmod_limma(block = 'Subject', coefs = model_coefs(object)) # intercept required!
beta(object)                #   betas : nlevel x nfeature
X(object)                   #   design : nlevel x nlevel
X(object) %*% beta(object)  # response : nlevel x nfeature

```

zero_to_na	<i>Change nondetect representation</i>
------------	--

Description

Change nondetect representation

Usage

```

zero_to_na(x, verbose = FALSE)

nan_to_na(x, verbose = FALSE)

na_to_zero(x, verbose = FALSE)

inf_to_na(x, verbose = FALSE)

minusinf_to_na(x, verbose = FALSE)

na_to_string(x)

```

Arguments

x	matrix
verbose	logical(1)

Value

Updated matrix

Examples

```

matrix(c(0, 7), nrow=1)
matrix(c(0, 7), nrow=1) %>% zero_to_na(verbose=TRUE)

matrix(c(NA, 7), nrow=1)
matrix(c(NA, 7), nrow=1) %>% na_to_zero(verbose=TRUE)

matrix(c(NaN, 7), nrow=1)
matrix(c(NaN, 7), nrow=1) %>% nan_to_na(verbose=TRUE)

```

```
matrix(c(Inf, 7), nrow=1)
matrix(c(Inf, 7), nrow=1) %>% inf_to_na(verbose=TRUE)

matrix(c(-Inf, 7), nrow=1)
matrix(c(-Inf, 7), nrow=1) %>% minusinf_to_na(verbose=TRUE)
```

Index

* datasets

- AUTONOMICS_DATASETS, [42](#)
- COMPOUNDDISCOVERER_PATTERNS, [55](#)
- DATADIR, [64](#)
- DIMREDUN, [71](#)
- LINMODENGINES, [122](#)
- MAXQUANT_PATTERNS, [133](#)
- MSIGCOLLECTIONSHUMAN, [146](#)
- MSIGDIR, [146](#)
- OPENTARGETSDIR, [148](#)
- PRECURSOR_QUANTITY, [181](#)
- TAXON_TO_ORGNAME, [215](#)
- TESTS, [215](#)
- TRANSFORMENGINES, [217](#)

* internal

- reexports, [195](#)
- .coxph, [7](#)
- .densities, [8](#)
- .extract_effectsize_features
 (.extract_p_features), [8](#)
- .extract_fdr_features
 (.extract_p_features), [8](#)
- .extract_n_features
 (.extract_p_features), [8](#)
- .extract_p_features, [8](#)
- .fit_survival, [11](#)
- .logrank(.coxph), [7](#)
- .merge, [14](#)
- .read_compounddiscoverer, [15](#)
- .read_compounddiscoverer_masslist, [15](#)
- .read_diann_precursors, [16](#)
- .read_diann_proteingroups
 (.read_diann_precursors), [16](#)
- .read_maxquant_phosphosites
 (.read_maxquant_proteingroups),
 [19](#)
- .read_maxquant_proteingroups, [19](#)
- .read_metabolon, [20](#)
- .read_rectangles, [22](#)

- .read_rnaseq_bams, [24](#)
- .read_rnaseq_counts
 (.read_rnaseq_bams), [24](#)
- .read_somascan, [27](#)
- .survdiff(.coxph), [7](#)
- %<>% (reexports), [195](#)
- %>% (reexports), [195](#)
- %<>%, [195](#)
- %>%, [195](#)
- ABBREV_TO_ORGNAME (TAXON_TO_ORGNAME),
 [215](#)
- abind(sbind), [198](#)
- abstract_fit, [29](#)
- abstractvar(modelvar), [140](#)
- abstractvec(modelvar), [140](#)
- add_adjusted_pvalues, [30](#)
- add_assay_means, [31](#)
- add_facetvars, [32](#)
- add_opentargets_by_uniprot, [33](#)
- add_psp, [34](#)
- add_smiles, [35](#)
- all_non_numeric(is_non_numeric), [111](#)
- altenrich, [35](#)
- analysis, [36](#)
- analysis, SummarizedExperiment-method
 (analysis), [36](#)
- analysis<- (analysis), [36](#)
- analysis<-, SummarizedExperiment, list-method
 (analysis), [36](#)
- analyze, [37](#)
- annotate_compounddiscoverer, [39](#)
- annotate_maxquant, [39](#)
- annotate_uniprot_rest, [41](#)
- assert_character_matrix
 (is_character_matrix), [103](#)
- assert_compounddiscoverer_output
 (is_diann_report), [106](#)
- assert_correlation_matrix
 (is_correlation_matrix), [105](#)

- `assert_diann_report(is_diann_report)`, 106
- `assert_fastadt(is_fastadt)`, 107
- `assert_fragpipe_tsv(is_diann_report)`, 106
- `assert_is_fraction(is_fraction)`, 108
- `assert_is_valid_sumexp`, 41
- `assert_maxquant_phosphosites(is_diann_report)`, 106
- `assert_maxquant_proteingroups(is_diann_report)`, 106
- `assert_positive_number(is_positive_number)`, 112
- `assert_scalar_subset(is_scalar_subset)`, 113
- `assert_valid_formula(is_valid_formula)`, 114
- `assert_weakly_positive_number(is_positive_number)`, 112
- `AUTONOMICS_DATASETS`, 42
- `awblinmod`, 42
- `awblinmod_limma(awblinmod)`, 42
- `awblinmod_lm(awblinmod)`, 42
- `awblinmod_lme(awblinmod)`, 42
- `awblinmod_lmer(awblinmod)`, 42
- `beta(X)`, 223
- `bin(factorize)`, 78
- `bin_assay(factorize)`, 78
- `biplot`, 44
- `biplot_corrections`, 45
- `biplot_covariates`, 46
- `biplot_transforms(plot_densities_transforms)`, 159
- `biplot_transforms_assays(plot_densities_transforms)`, 159
- `block2limma`, 47
- `block2lm`, 48
- `block2lme`, 49
- `block2lmer`, 50
- `block_has_two_levels`, 51
- `cbind_imputed(split_samples)`, 202
- `center`, 51
- `center_mean(center)`, 51
- `center_median(center)`, 51
- `code`, 52
- `code_control(code)`, 52
- `code_deviation(code)`, 52
- `code_deviation_first(code)`, 52
- `code_diff(code)`, 52
- `code_diff_forward(code)`, 52
- `code_helmert(code)`, 52
- `code_helmert_forward(code)`, 52
- `coefs(fits)`, 88
- `coefs_estimable(pvalues_estimable)`, 183
- `collapse_in(count_in)`, 61
- `collapsed_entrezg_to_symbol`, 55
- `COMPOUNDDISCOVERER_PATTERNS`, 55
- `contr.diff(code)`, 52
- `contr.treatment.explicit(code)`, 52
- `contrast_coefs`, 57
- `contrast_subgroup_cols`, 58
- `contrast_subgroup_rows(contrast_subgroup_cols)`, 58
- `contrastdt`, 56
- `count_in`, 61
- `count_out(count_in)`, 61
- `counts`, 58
- `counts, SummarizedExperiment-method(counts)`, 58
- `counts2cpm`, 59
- `counts2tpm`, 60
- `counts<-(counts)`, 58
- `counts<-, SummarizedExperiment, matrix-method(counts)`, 58
- `counts<-, SummarizedExperiment, NULL-method(counts)`, 58
- `counts<-, SummarizedExperiment, numeric-method(counts)`, 58
- `cpm`, 62
- `cpm, SummarizedExperiment-method(cpm)`, 62
- `cpm2counts(counts2cpm)`, 59
- `cpm<-(cpm)`, 62
- `cpm<-, SummarizedExperiment, matrix-method(cpm)`, 62
- `cpm<-, SummarizedExperiment, numeric-method(cpm)`, 62
- `create_design`, 63
- `data.table`, 195
- `data.table(reexports)`, 195
- `DATADIR`, 64
- `default_formula`, 66
- `default_geom`, 67

- default_sfile, 68
- defaultsigfile, 66
- demultiplex, 68
- densities (.densities), 8
- dequantify, 69
- dequantify_compounddiscoverer, 70
- DIMREDENGINES (DIMREDUN), 71
- DIMREDSUPER (DIMREDUN), 71
- DIMREDUN, 71
- downfeatures (modelvar), 140
- download_data (DATADIR), 64
- download_gtf, 71
- download_mcclain21, 72
- dt2mat, 73
- effectdt (modelvar), 140
- effectmat (modelvar), 140
- effectsize (modelvar), 140
- effectvar (modelvar), 140
- effectvec (modelvar), 140
- enrichment, 73
- ens2org, 75
- entrezg_to_symbol, 76
- EXISTENCE_TO_NUMBER (TAXON_TO_ORGNAME), 215
- exp2transform (log2transform), 128
- extract, 195
- extract (reexports), 195
- extract_contrast_features
 (.extract_p_features), 8
- extract_rectangle, 76
- factor.vars (left.vars), 117
- factor.vars, formula, data.table-method
 (left.vars), 117
- factor.vars, formula, SummarizedExperiment-method
 (left.vars), 117
- factor2logical (logical2factor), 129
- factorize, 78
- factorize_assay (factorize), 78
- fbind (sbind), 198
- fcluster, 81
- fcor (mdsplot), 135
- fdata, 82
- fdata, SummarizedExperiment-method
 (fdata), 82
- fdata<- (fdata), 82
- fdata<-, SummarizedExperiment, data.frame-method
 (fdata), 82
- fdist (mdsplot), 135
- fdr2p, 84
- fdrmat (modelvar), 140
- fdrvar (modelvar), 140
- fdrvec (modelvar), 140
- fdt (fdata), 82
- fdt, SummarizedExperiment-method
 (fdata), 82
- fdt<- (fdata), 82
- fdt<-, SummarizedExperiment, data.table-method
 (fdata), 82
- filter_exprs_replicated_in_some_subgroup, 85
- filter_features, 86
- filter_medoid, 86
- filter_samples, 87
- fit_limma (LINMOD), 118
- fit_lm (LINMOD), 118
- fit_lme (LINMOD), 118
- fit_lmer (LINMOD), 118
- fit_survival (.fit_survival), 11
- fit_wilcoxon (LINMOD), 118
- fitcoefs (fits), 88
- fits, 88
- fix_xlgenes, 89
- flevels, 89
- fnames, 90
- fnames, SummarizedExperiment-method
 (fnames), 90
- fnames<- (fnames), 90
- fnames<-, SummarizedExperiment, character-method
 (fnames), 90
- formula2str, 91
- fscale (log2transform), 128
- ftype, 91
- fvalues, 92
- fvars, 93
- fvars, SummarizedExperiment-method
 (fvars), 93
- fvars<- (fvars), 93
- fvars<-, SummarizedExperiment, character-method
 (fvars), 93
- genome_to_orfdb, 93
- group_by_level, 94
- guess_compounddiscoverer_quantity, 95
- guess_fitsep, 96
- guess_maxquant_quantity, 96
- guess_sep, 97

- has_multiple_levels, 98
- hdlproteins, 100
- impute, 100
- inf_to_na (zero_to_na), 224
- installed, 102
- invert_subgroups, 103
- invnorm (log2transform), 128
- is_character_matrix, 103
- is_collapsed_subset, 104
- is_compounddiscoverer_output, 105
- is_correlation_matrix, 105
- is_diann_report, 106
- is_fastadt, 107
- is_file, 107
- is_fraction, 108
- is_fragpipe_tsv, 109
- is_imputed, 109
- is_imputed, SummarizedExperiment-method
(is_imputed), 109
- is_imputed<- (is_imputed), 109
- is_imputed<-, SummarizedExperiment, matrix-method
(is_imputed), 109
- is_imputed<-, SummarizedExperiment, NULL-method
(is_imputed), 109
- is_maxquant_phosphosites, 110
- is_maxquant_proteingroups, 111
- is_non_numeric, 111
- is_positive_number, 112
- is_scalar_subset, 113
- is_sig, 114
- is_valid_formula, 114
- is_weakly_positive_number
(is_positive_number), 112
- keep_estimable_features, 116
- label2index, 116
- lda (pca), 151
- left.vars, 117
- LINMOD, 118
- linmod_limma (LINMOD), 118
- linmod_lm (LINMOD), 118
- linmod_lme (LINMOD), 118
- linmod_lmer (LINMOD), 118
- linmod_wilcoxon (LINMOD), 118
- LINMODENGINES, 122
- list2mat, 122
- list_files, 123
- loadingmat (scoremat), 200
- loadings (scoremat), 200
- log2counts, 123
- log2counts, SummarizedExperiment-method
(log2counts), 123
- log2counts<- (log2counts), 123
- log2counts<-, SummarizedExperiment, matrix-method
(log2counts), 123
- log2counts<-, SummarizedExperiment, numeric-method
(log2counts), 123
- log2cpm, 124
- log2cpm, SummarizedExperiment-method
(log2cpm), 124
- log2cpm<- (log2cpm), 124
- log2cpm<-, SummarizedExperiment, matrix-method
(log2cpm), 124
- log2cpm<-, SummarizedExperiment, numeric-method
(log2cpm), 124
- log2diffs, 125
- log2diffs, SummarizedExperiment-method
(log2diffs), 125
- log2diffs<- (log2diffs), 125
- log2diffs<-, SummarizedExperiment, matrix-method
(log2diffs), 125
- log2diffs<-, SummarizedExperiment, numeric-method
(log2diffs), 125
- log2proteins, 126
- log2proteins, SummarizedExperiment-method
(log2proteins), 126
- log2proteins<- (log2proteins), 126
- log2proteins<-, SummarizedExperiment, matrix-method
(log2proteins), 126
- log2proteins<-, SummarizedExperiment, numeric-method
(log2proteins), 126
- log2sites, 126
- log2sites, SummarizedExperiment-method
(log2sites), 126
- log2sites<- (log2sites), 126
- log2sites<-, SummarizedExperiment, matrix-method
(log2sites), 126
- log2sites<-, SummarizedExperiment, numeric-method
(log2sites), 126
- log2tpm, 127
- log2tpm, SummarizedExperiment-method
(log2tpm), 127
- log2tpm<- (log2tpm), 127
- log2tpm<-, SummarizedExperiment, matrix-method
(log2tpm), 127

log2tpm<- , SummarizedExperiment, numeric-methodpls (pca), 151
 (log2tpm), 127
 log2transform, 128
 logical2factor, 129

 make_alpha_palette, 130
 make_colors, 131
 make_volcano_dt, 131
 map_fvalues, 132
 mat2dt (dt2mat), 73
 matrix2sumexp, 133
 MAXQUANT_PATTERNS, 133
 mclust_breaks, 134
 mclust_parameters (overall_parameters), 150
 mdsplot, 135
 merge_compounddiscoverer, 136
 merge_fdata (merge_sdata), 138
 merge_fdt (merge_sdata), 138
 merge_ffile (merge_sample_file), 137
 merge_sample_excel, 136
 merge_sample_file, 137
 merge_sdata, 138
 merge_sdt (merge_sdata), 138
 message_df, 139
 minusinf_to_na (zero_to_na), 224
 mixtools_breaks (mclust_breaks), 134
 mixtools_parameters
 (overall_parameters), 150
 model_coefs (contrast_coefs), 57
 modeldt (modelvar), 140
 modelfeatures (modelvar), 140
 modelmat (modelvar), 140
 modelvar, 140
 modelvec (modelvar), 140
 MSIGCOLLECTIONSHUMAN, 146
 MSIGCOLLECTIONSMOUSE
 (MSIGCOLLECTIONSHUMAN), 146
 MSIGDIR, 146

 na_to_string (zero_to_na), 224
 na_to_zero (zero_to_na), 224
 nan_to_na (zero_to_na), 224
 nfactors, 147
 no_nas (systematic_nas), 213

 object1, 148
 object2 (object1), 148
 OPENTARGETSDIR, 148

 order_on_effect (order_on_p), 149
 order_on_p, 149
 order_on_t (order_on_p), 149
 overall_parameters, 150

 parse_maxquant_hdrs (read_uniprot), 194
 pca, 151
 pdt (modelvar), 140
 pg_to_canonical, 153
 pg_to_isoforms (pg_to_canonical), 153
 plot_coef_densities, 154
 plot_contrast_venn, 155
 plot_contrastogram, 154
 plot_data, 156
 plot_densities, 157, 176
 plot_densities_transforms, 159
 plot_design, 162
 plot_detections (plot_sample_nas), 170
 plot_exprs, 163, 176
 plot_exprs_per_coef, 166
 plot_feature_boxplots (plot_exprs), 163
 plot_feature_densities
 (plot_densities), 157
 plot_feature_violins (plot_violins), 174
 plot_fit_summary, 167
 plot_heatmap, 168
 plot_matrix, 169
 plot_sample_boxplots, 159
 plot_sample_boxplots (plot_exprs), 163
 plot_sample_densities, 165, 167
 plot_sample_densities (plot_densities), 157
 plot_sample_nas, 170
 plot_sample_violins, 159, 165, 167
 plot_sample_violins (plot_violins), 174
 plot_subgroup_nas (plot_sample_nas), 170
 plot_subgroup_points, 171
 plot_subgroup_violins (plot_violins), 174
 plot_summarized_detections
 (plot_sample_nas), 170
 plot_summary, 172
 plot_survival (.fit_survival), 11
 plot_venn, 173
 plot_venn_heatmap, 174
 plot_violins, 174

- plot_violins_transforms
(plot_densities_transforms),
159
- plot_volcano, 177
- plot_x_density, 179
- plot_xy_density (plot_x_density), 179
- plot_xy_scatter (plot_x_density), 179
- plot_y_density (plot_x_density), 179
- plotmat, 155
- pls (pca), 151
- pmat (modelvar), 140
- PRECURSOR_QUANTITY, 181
- prep_survival (.fit_survival), 11
- preprocess_rnaseq_counts, 181
- pull_columns, 183
- pvalues_estimable, 183
- pvar (modelvar), 140
- pvec (modelvar), 140

- quantile_breaks (mclust_breaks), 134
- quantnorm (log2transform), 128

- random_nas (systematic_nas), 213
- read_affymetrix, 185
- read_compounddiscoverer, 186
- read_contaminantdt (read_uniprot), 194
- read_diann (.read_diann_precursors), 16
- read_diann_pgmatrix, 187
- read_diann_phosphodiffs
(read_diann_pgmatrix), 187
- read_diann_phosphosites
(read_diann_pgmatrix), 187
- read_diann_proteingroups
(.read_diann_precursors), 16
- read_fragpipe, 188
- read_maxquant_phosphosites, 188
- read_maxquant_proteingroups, 190
- read_metabolon (.read_metabolon), 20
- read_msigt, 36, 192
- read_olink, 193
- read_phosphosites
(read_maxquant_phosphosites),
188
- read_proteingroups
(read_maxquant_proteingroups),
190
- read_rectangles (.read_rectangles), 22
- read_rnaseq_bams (.read_rnaseq_bams), 24
- read_rnaseq_counts (.read_rnaseq_bams),
24
- read_salmon, 194
- read_somascan (.read_somascan), 27
- read_uniprot, 194
- recollapse (uncollapse), 218
- reexports, 195
- reset_fit, 196
- REVIEWED_TO_NUMBER (TAXON_TO_ORGNAME),
215
- right.vars (left.vars), 117
- rm_diann_contaminants, 196
- rm_missing_in_all_samples, 197
- rm_missing_in_some_samples
(rm_missing_in_all_samples),
197
- rm_singleton_samples
(rm_unmatched_samples), 197
- rm_unmatched_samples, 197

- sampleid_values (svalues), 211
- sbind, 198
- scaledlibsizes, 199
- scor (mdsplot), 135
- scoremat, 200
- scores (scoremat), 200
- sdata (fdata), 82
- sdata, SummarizedExperiment-method
(fdata), 82
- sdata<- (fdata), 82
- sdata<- , SummarizedExperiment, data.frame-method
(fdata), 82
- sdata<- , SummarizedExperiment, DataFrame-method
(fdata), 82
- sdist (mdsplot), 135
- sdt (fdata), 82
- sdt, SummarizedExperiment-method
(fdata), 82
- sdt<- (fdata), 82
- sdt<- , SummarizedExperiment, data.table-method
(fdata), 82
- slevels, 201
- sma (pca), 151
- snames, 201
- snames, SummarizedExperiment-method
(snames), 201
- snames<- (snames), 201
- snames<- , SummarizedExperiment, character-method
(snames), 201

- split_extract(nfactors), 147
- split_extract_fixed(nfactors), 147
- split_extract_regex(nfactors), 147
- split_features(split_samples), 202
- split_samples, 202
- spls(pca), 151
- sscale(log2transform), 128
- stepauc, 203
- stri_any_regex, 203
- stri_detect_fixed_in_collapsed, 204
- subgroup_array, 205
- subgroup_levels(slevels), 201
- subgroup_matrix(subgroup_array), 205
- subgroup_values(svalues), 211
- subtract_baseline, 205
- subtract_differences
 - (subtract_baseline), 205
- subtract_pairs(subtract_baseline), 205
- sumexp_to_groupdt(sumexp_to_widedt), 208
- sumexp_to_longdt(sumexp_to_widedt), 208
- sumexp_to_tsv, 208
- sumexp_to_widedt, 208
- sumexplist_to_longdt, 207
- summarize_fit, 209
- survobj, 210
- svalues, 211
- svalues<- (svalues), 211
- svalues<-, SummarizedExperiment, character-method
 - (svalues), 211
- svars, 212
- svars, MultiAssayExperiment-method
 - (svars), 212
- svars, SummarizedExperiment-method
 - (svars), 212
- svars<- (svars), 212
- svars<-, MultiAssayExperiment, character-method
 - (svars), 212
- svars<-, SummarizedExperiment, character-method
 - (svars), 212
- systematic_nas, 213
- tag_features, 213
- tag_hdlproteins, 214
- taxon2org(ens2org), 75
- TAXON_TO_ORGNAME, 215
- tdt(modelvar), 140
- TESTS, 215
- tmat(modelvar), 140
- tpm, 216
- tpm, SummarizedExperiment-method(tpm), 216
- tpm<- (tpm), 216
- tpm<-, SummarizedExperiment, matrix-method
 - (tpm), 216
- tpm<-, SummarizedExperiment, numeric-method
 - (tpm), 216
- TRANSFORMENGINES, 217
- TRANSFORMSTRICT (TRANSFORMENGINES), 217
- tvar(modelvar), 140
- tvec(modelvar), 140
- twofactor_sumexp, 217
- uncollapse, 218
- upfeatures(modelvar), 140
- values, 218
- values, SummarizedExperiment-method
 - (values), 218
- values<- (values), 218
- values<-, SummarizedExperiment, matrix-method
 - (values), 218
- values<-, SummarizedExperiment, numeric-method
 - (values), 218
- varlevels_dont_clash, 219
- venn_detects, 220
- vsr(log2transform), 128
- weights, 221
- weights, SummarizedExperiment-method
 - (weights), 221
- weights<- (weights), 221
- weights<-, SummarizedExperiment, matrix-method
 - (weights), 221
- weights<-, SummarizedExperiment, NULL-method
 - (weights), 221
- weights<-, SummarizedExperiment, numeric-method
 - (weights), 221
- write_ods(write_xl), 222
- write_xl, 222
- X, 223
- zero_to_na, 224
- zscore(log2transform), 128