

# Managing and analyzing multiple NGS samples with Bioconductor bamViews objects: application to RNA-seq

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# 1 Introduction

We consider a lightweight approach to Bioconductor-based management and interrogation of multiple samples to which NGS methods have been applied.

The basic data store is the binary SAM (BAM) format (Li et al., 2009). This format is widely used in the 1000 genomes project, and transformations between SAM/BAM and output formats of various popular alignment programs are well-established. Bioconductor's *Rsamtools* package allows direct use of important SAM data interrogation facilities from R.

# 2 Basic design

A collection of NGS samples is represented through the associated set of BAM files and BAI index files. These can be stored in the `inst/bam` folder of an R package to facilitate documented programmatic access through R file navigation facilities, or the BAM/BAI files can be accessed through arbitrary path or URL references.

The `bamViews` class is defined to allow reliable fine-grained access to the NGS data along with relevant metadata. A `bamViews` instance contains access path information for a set of related BAM/BAI files, along with sample metadata and an optional specification of genomic ranges of interest.

A key design aspect of the `bamViews` class is preservation of semantics of the `X[G, S]` idiom familiar from *ExpressionSet* objects for management of multiple microarrays. With `ExpressionSet` instances, `G` is a predicate specifying selection of microarray probes of interest. With *bamViews* instances, `G` is a predicate specifying selection of genomic features of interest. At present, for *bamViews*, selection using `G` involves ranges of genomic coordinates.

# 3 Illustration

Data from four samples from a yeast RNA-seq experiment (two wild type, two 'RLP' mutants) are organized in the *leeBamViews* package. The data are collected to allow regeneration of aspects of Figure 8 of Lee et al. (2008). We obtained all reads between bases 800000 and 900000 of yeast chromosome XIII.

We have not yet addressed durable serialization of manager objects, so the *bamViews* instance is created on the fly.

```
> library(leeBamViews) # bam files stored in package
> bpaths = dir(system.file("bam", package="leeBamViews"), full=TRUE, patt="bam$")
> #
> # extract genotype and lane information from filenames
> #
> gt = gsub(".*/", "", bpaths)
```

```

> gt = gsub("_.*", "", gt)
> lane = gsub(".*(.)$", "\\1", gt)
> geno = gsub(".$", "", gt)
> #
> # format the sample-level information appropriately
> #
> pd = DataFrame(geno=geno, lane=lane, row.names=paste(geno,lane,sep="."))
> prd = new("DataFrame") # protocol data could go here
> #
> # create the views object, adding some arbitrary experiment-level information
> #
> bs1 = BamViews(bamPaths=bpaths, bamSamples=pd,
+               bamExperiment=list(annotation="org.Sc.sgd.db"))
> bs1

BamViews dim: 0 ranges x 8 samples
names: isowt.5 isowt.6 ... xrn.1 xrn.2
detail: use bamPaths(), bamSamples(), bamRanges(), ...

> #
> # get some sample-level data
> #
> bamSamples(bs1)$geno

[1] "isowt" "isowt" "rlp"   "rlp"   "ssr"   "ssr"   "xrn"   "xrn"

```

We would like to operate on specific regions of chr XIII for all samples. Note that the aligner in use (bowtie) employed “Scchr13” to refer to this chromosome. We add a *GRanges* instance to the view to identify the region of interest.

```

> START = c(861250, 863000)
> END = c(862750, 864000)
> exc = GRanges(seqnames = "Scchr13", IRanges(start = START, end = END),
+               strand = "+")
> bamRanges(bs1) = exc
> bs1

```

```

BamViews dim: 2 ranges x 8 samples
names: isowt.5 isowt.6 ... xrn.1 xrn.2
detail: use bamPaths(), bamSamples(), bamRanges(), ...

```

A common operation will be to extract coverage information. We use a transforming method, *readGappedAlignments*, from the *GenomicRanges* package to extract reads and metadata for each region and each sample.

```

> covex = RleList(lapply(bamPaths(bs1), function(x) coverage(readGappedAlignments(x))
> names(covex) = gsub(".bam$", "", basename(bamPaths(bs1)))
> head(covex, 3)

```

SimpleRleList of length 3

\$isowt5\_13e

'integer' Rle of length 900030 with 21818 runs

```

Lengths: 799974      2      2      1      6 ...      1      10      7      1
Values :      0      1      2      3      4 ...      5      4      3      2

```

\$isowt6\_13e

'integer' Rle of length 900035 with 21798 runs

```

Lengths: 799976      2      3     14     13 ...      1     17      1      4
Values :      0      1      2      3      4 ...      4      3      2      1

```

\$rlp5\_13e

'integer' Rle of length 900032 with 23036 runs

```

Lengths: 799974      2      6     25      3 ...      3      4      2     30
Values :      0      1      2      3      4 ...      2      3      2      1

```

Let's visualize what we have so far. We use *GenomeGraphs* and add some supporting software.

```

> library(GenomeGraphs)
> cov2baseTrack = function(rle, start, end, dp = DisplayPars(type = "l",
+   lwd = 0.5, color = "black"), countTx = function(x) log10(x +
+   1)) {
+   require(GenomeGraphs)
+   if (!is(rle, "Rle"))
+     stop("requires instance of Rle")
+   dat = runValue(rle)
+   loc = cumsum(runLength(rle))
+   ok = which(loc >= start & loc <= end)
+   makeBaseTrack(base = loc[ok], value = countTx(dat[ok]), dp = dp)
+ }
> trs = lapply(covex, function(x) cov2baseTrack(x, START[1], END[1],
+   countTx = function(x) pmin(x, 80)))
> ac = as.character
> names(trs) = paste(ac(bamSamples(bs1)$geno), ac(bamSamples(bs1)$lane),
+   sep = "")
> library(biomaRt)
> mart = useMart("ensembl", "scerevisiae_gene_ensembl")
> gr = makeGeneRegion(START, END, chromosome = "XIII", strand = "+",

```

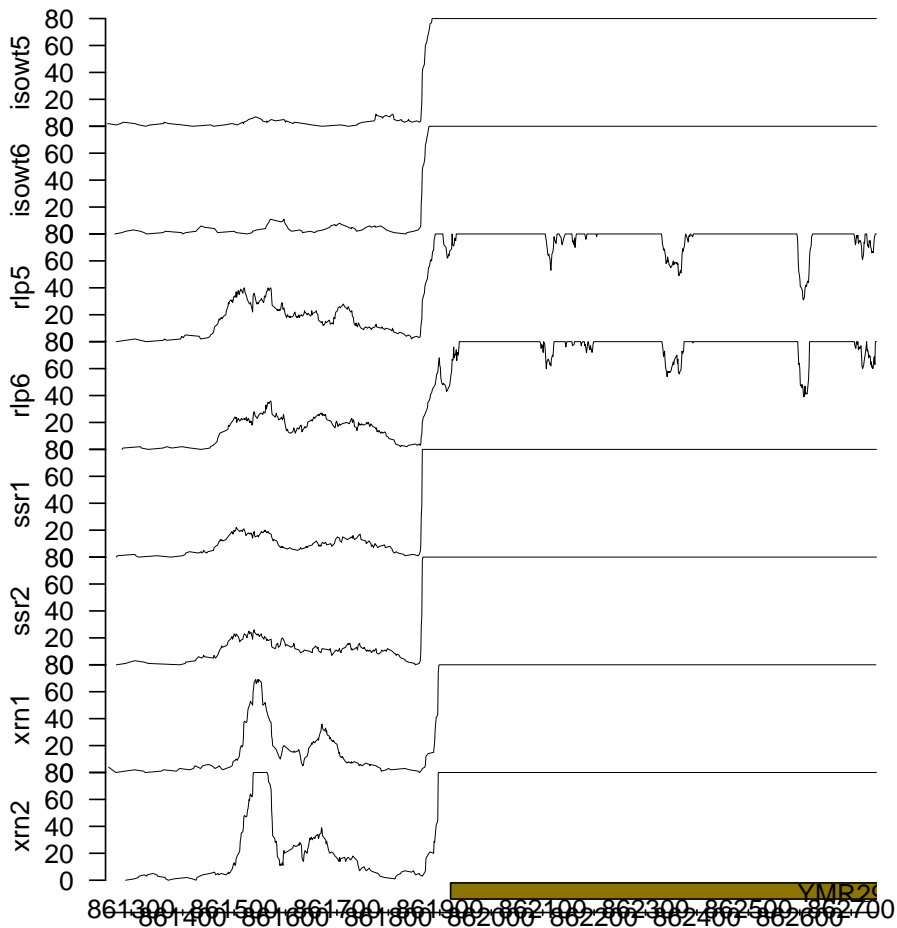
```

+   biomart = mart, dp = DisplayPars(plotId = TRUE, idRotation = 0,
+   idColor = "black"))
> trs[[length(trs) + 1]] = gr
> trs[[length(trs) + 1]] = makeGenomeAxis()

> print(gdPlot(trs, minBase = START[1], maxBase = END[1]))

```

NULL



We can encapsulate this to something like:

```

> plotStrains = function(bs, query, start, end, snames, mart, chr,
+   strand = "+") {
+   filtbs = bs[query, ]
+   cov = lapply(filtbs, coverage)
+   covtrs = lapply(cov, function(x) cov2baseTrack(x[[1]], start,
+   end, countTx = function(x) pmin(x, 80)))
+   names(covtrs) = snames

```

```

+   gr = makeGeneRegion(start, end, chromosome = chr, strand = strand,
+     biomart = mart, dp = DisplayPars(plotId = TRUE, idRotation = 0,
+     idColor = "black"))
+   grm = makeGeneRegion(start, end, chromosome = chr, strand = "-",
+     biomart = mart, dp = DisplayPars(plotId = TRUE, idRotation = 0,
+     idColor = "black"))
+   covtrs[[length(covtrs) + 1]] = gr
+   covtrs[[length(covtrs) + 1]] = makeGenomeAxis()
+   covtrs[[length(covtrs) + 1]] = grm
+   gdPlot(covtrs, minBase = start, maxBase = end)
+ }

```

## 4 Comparative counts in a set of regions of interest

### 4.1 Counts in a regular partition

The supplementary information for the Lee paper includes data on unannotated transcribed regions reported in other studies. We consider the study of David et al., confining attention to chromosome XIII. If you wanted to study their intervals you could use code like:

```

> data(leeUnn)
> names(leeUnn)
> leeUnn[1:4, 1:8]
> table(leeUnn$study)
> l13 = leeUnn[leeUnn$chr == 13, ]
> l13d = na.omit(l13[l13$study == "David", ])
> d13r = GRanges(seqnames = "Scchr13", IRanges(l13d$start, l13d$end),
+   strand = ifelse(l13d$strand == 1, "+", ifelse(l13d$strand ==
+     "0", "*", "-")))
> elementMetadata(d13r)$name = paste("dav13x", 1:length(d13r),
+   sep = ".")
> bamRanges(bs1) = d13r
> d13tab = tabulateReads(bs1)

```

but our object `bs1` is too restricted in its coverage. Instead, we illustrate with a small set of subintervals of the basic interval in use:

```

> myrn = GRanges(seqnames = "Scchr13", IRanges(start = seq(861250,
+   862750, 100), width = 100), strand = "+")
> elementMetadata(myrn)$name = paste("til", 1:length(myrn), sep = ".")
> bamRanges(bs1) = myrn
> tabulateReads(bs1, "+")

```

	til.1	til.2	til.3	til.4	til.5	til.6	til.7	til.8	til.9	til.10
start	861250	861350	861450	861550	861650	861750	861850	861950	862050	862150
end	861349	861449	861549	861649	861749	861849	861949	862049	862149	862249
isowt.5	1	1	3	6	2	7	299	605	406	380
isowt.6	2	6	9	12	7	4	306	664	456	382
rlp.5	1	5	65	53	36	11	158	247	186	145
rlp.6	3	2	47	48	36	16	123	238	163	158
ssr.1	2	5	35	27	21	8	423	699	539	496
ssr.2	2	6	43	37	26	13	443	838	613	509
xrn.1	7	8	74	78	24	5	180	443	357	288
xrn.2	4	8	96	109	30	8	225	610	464	356

	til.11	til.12	til.13	til.14	til.15	til.16
start	862250	862350	862450	862550	862650	862750
end	862349	862449	862549	862649	862749	862849
isowt.5	481	553	895	608	637	693
isowt.6	446	516	870	667	689	698
rlp.5	174	180	316	242	238	275
rlp.6	190	215	336	262	269	279
ssr.1	573	596	966	714	661	768
ssr.2	575	604	987	757	735	804
xrn.1	349	482	678	533	395	331
xrn.2	428	572	836	632	453	406

## 4.2 Counts in annotated intervals: genes

We can use Bioconductor annotation resources to acquire boundaries of yeast genes on our subregion of chromosome 13.

In the following chunk we generate annotated ranges of genes on the Watson strand.

```

> library(org.Sc.sgd.db)
> library(IRanges)
> c13g = get("13", revmap(org.Sc.sgdCHR)) # all genes on chr13
> c13loc = unlist(mget(c13g, org.Sc.sgdCHRLoc)) # their 'start' addresses
> c13locend = unlist(mget(c13g, org.Sc.sgdCHRLOCEND))
> c13locp = c13loc[c13loc>0] # confine attention to + strand
> c13locendp = c13locend[c13locend>0]
> ok = !is.na(c13locp) & !is.na(c13locendp)
> c13pr = GRanges(seqnames="Scchr13", IRanges(c13locp[ok], c13locendp[ok]),
+ strand="+") # store and clean up names
> elementMetadata(c13pr)$name = gsub(".13$", "", names(c13locp[ok]))
> c13pr

```

GRanges with 297 ranges and 1 elementMetadata value

seqnames	ranges	strand	name
----------	--------	--------	------

	<Rle>	<IRanges>	<Rle>		<character>
[1]	Scchr13	[268031, 268149]	+		CEN13
[2]	Scchr13	[267174, 267800]	+		YML001W
[3]	Scchr13	[260221, 261609]	+		YML005W
[4]	Scchr13	[253848, 255800]	+		YML007W
[5]	Scchr13	[247677, 250868]	+		YML010W
[6]	Scchr13	[246116, 246751]	+		YML012W
[7]	Scchr13	[244149, 245903]	+		YML013W
[8]	Scchr13	[243225, 244064]	+		YML014W
[9]	Scchr13	[236588, 238731]	+		YML017W
...	...	...	...	...	...
[289]	Scchr13	[768547, 768878]	+		YMRWdelta19
[290]	Scchr13	[808999, 809232]	+		YMRWdelta20
[291]	Scchr13	[837449, 837763]	+		YMRWdelta21
[292]	Scchr13	[503770, 504140]	+		YMRWtau2
[293]	Scchr13	[463553, 463624]	+		tD(GUC)M
[294]	Scchr13	[290800, 290871]	+		tE(UUC)M
[295]	Scchr13	[352279, 352369]	+		tF(GAA)M
[296]	Scchr13	[363063, 363134]	+		tH(GUG)M
[297]	Scchr13	[504894, 505007]	+		tL(CAA)M

```
seqlengths
```

```
Scchr13
NA
```

```
> c13pro = c13pr[ order(ranges(c13pr)), ]
```

That's the complete set of genes on the Watson strand of chromosome XIII. In the *leeBamViews* package, we do not have access to all these, but only those lying in a 100kb interval.

```
> lim = GRanges(seqnames = "Scchr13", IRanges(8e+05, 9e+05), strand = "+")
> c13pro1 = c13pro[which(c13pro %in% lim), ]
```

Now that we have a set of annotation-based genomic regions, we can tabulate read counts lying in those regions and obtain an annotated matrix.

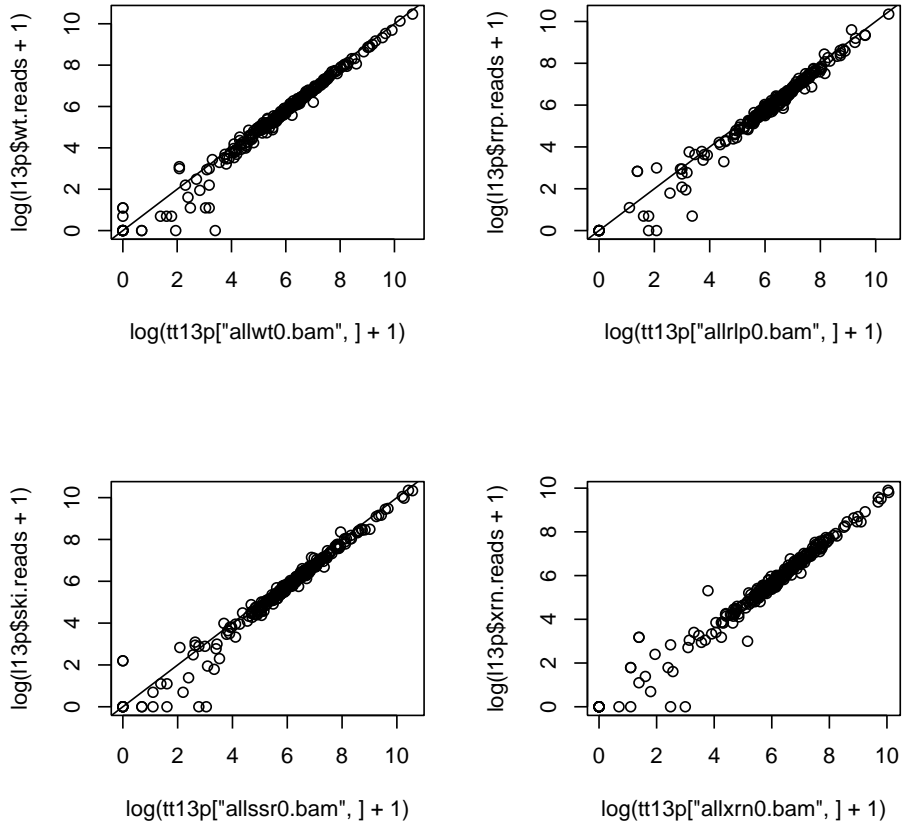
```
> bamRanges(bs1) = c13pro1
> annotab = tabulateReads(bs1, strandmarker = "+")
```

## 5 Larger scale sanity check

The following plot compares read counts published with the Lee et al. (2008) paper to those computed by the methods sketched here, for all regions noted on the plus strand of



chromosome XIII. Exact correspondence is not expected because of different approaches to read filtering.



## 6 Statistical analyses of differential expression

### 6.1 Using edgeR

Statistical analysis of read counts via negative binomial distributions with moderated dispersion is developed in Robinson and Smyth (2008). The *edgeR* differential expression statistics are computed using regional read counts, and total library size plays a role. We compute total read counts directly (the operation can be somewhat slow for very large BAM files):

```
> totcnts = totalReadCounts(bs1)
```

In the following demonstration, we will regard multiple lanes from the same genotype as replicates. This is probably inappropriate for this method; the original authors tested for lane effects and ultimately combined counts across lanes within strain.

```

> library(edgeR)
> #
> # construct an edgeR container for read counts, including
> # genotype and region (gene) metadata
> #
> dgel1 = DGEList( counts=t(annotab)[,-c(1,2)],
+   group=factor(bamSamples(bs1)$geno),
+   lib.size=totcnts, genes=colnames(annotab))
> #
> # compute a dispersion factor for the negative binomial model
> #
> cd = estimateCommonDisp(dgel1)
> #
> # test for differential expression between two groups
> # for each region
> #
> et12 = exactTest(cd)

```

Comparison of groups: rlp - isowt

```

> #
> # display statistics for the comparison
> #
> tt12 = topTags(et12)
> tt12

```

Comparison of groups: rlp - isowt

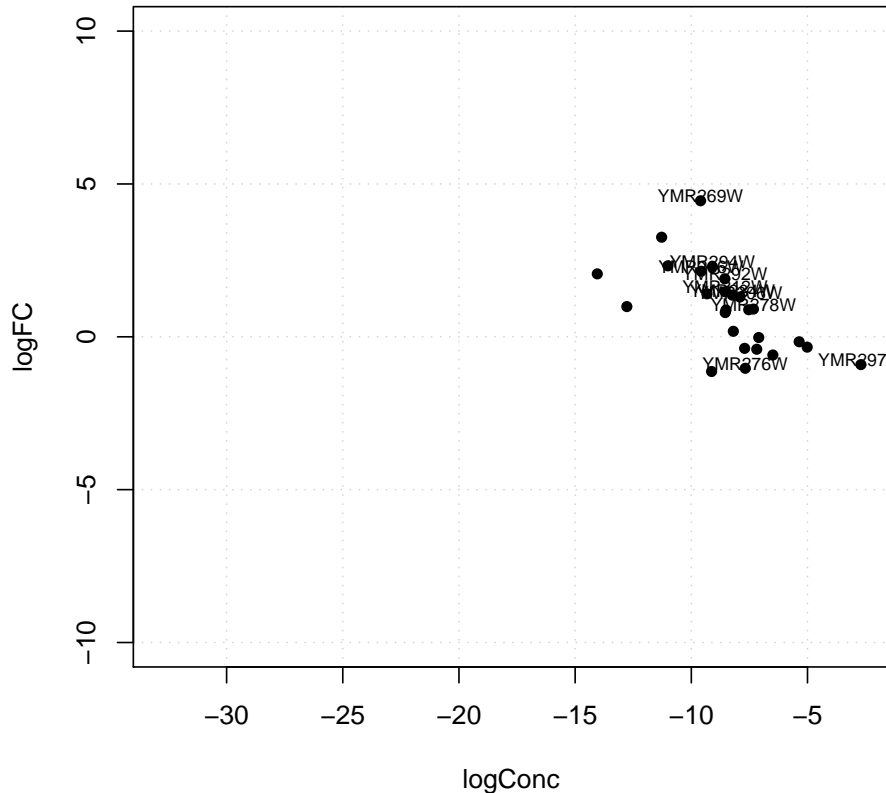
	genes	logConc	logFC	PValue	FDR
YMR297W	YMR297W	-2.701788	-0.9089939	6.640849e-260	1.793029e-258
YMR269W	YMR269W	-9.603251	4.4504807	9.748151e-88	1.316000e-86
YMR294W	YMR294W	-9.093662	2.2997348	1.199710e-31	1.079739e-30
YMR292W	YMR292W	-8.557110	1.8989068	4.386646e-31	2.960986e-30
YMR306W	YMR306W	-7.909224	1.3029908	5.286705e-23	2.854821e-22
YMR284W	YMR284W	-8.234354	1.3643876	1.380457e-20	6.212055e-20
YMR286W	YMR286W	-9.577072	2.1461646	5.032338e-20	1.941045e-19
YMR312W	YMR312W	-8.544790	1.4744256	2.254859e-19	7.610150e-19
YMR278W	YMR278W	-7.327946	0.9077038	6.124400e-17	1.837320e-16
YMR276W	YMR276W	-7.678076	-1.0290934	1.161190e-16	3.135214e-16

An analog of the “MA-plot” familiar from microarray studies is available for this analysis. The ‘concentration’ is the log proportion of reads present in each gene, and the “log fold change” is the model-based estimate of relative abundance. In the following display we label the top 10 genes (those with smallest FDR).

```

> plotFC(et12, cex = 0.8, ylim = c(-10, 10))
> text(tt12$table$logConc, tt12$table$logFC + 0.15, as.character(tt12$table$genes),
+      cex = 0.65)

```



## 7 Summary

- The BAM format provides reasonably compact and comprehensive information about a alignments of short reads obtained in a sequencing experiment. `samtools` utilities permit efficient random access to read collections of interest.
- *Rsamtools* brings `samtools` functionality into R, principally through the `scanBam` method, which is richly parameterized so that many details of access to and filtering of reads from BAM files can be controlled in R.
- *Rsamtools* defines the `bamViews` container for management of collections of BAM files. Read data are managed external to R; data on aligned reads can be imported efficiently, and “streaming read” models for scanning large collections of

reads can be used. Many embarrassingly parallel operations can be accomplished concurrently using *multicore* or similar packages.

- The *leeBamViews* package provides small excerpts from BAM files generated after bowtie alignment of FASTQ records available through the NCBI short read archives. These excerpts can be analyzed using code shown in this vignette.
- After the count data have been generated, various approaches to inference on differential expression are available. We consider the moderated negative binomial models of *edgeR* above; more general variance modeling is available in the developmental *DESeq* package.

## References

Albert Lee, Kasper Daniel Hansen, James Bullard, Sandrine Dudoit, Gavin Sherlock, and Michael Snyder. Novel low abundance and transient rnas in yeast revealed by tiling microarrays and ultra high-throughput sequencing are not conserved across closely related yeast species. *PLoS Genet*, 4(12):e1000299, Dec 2008. doi: 10.1371/journal.pgen.1000299.t002.

Heng Li, Bob Handsaker, Alec Wysoker, Tim Fennell, Jue Ruan, Nils Homer, Gabor Marth, Goncalo Abecasis, Richard Durbin, and 1000 Genome Project Data Processing Subgroup. The sequence alignment/map format and samtools. *Bioinformatics*, 25(16): 2078–9, Aug 2009. doi: 10.1093/bioinformatics/btp352.

Mark D Robinson and Gordon K Smyth. Small-sample estimation of negative binomial dispersion, with applications to sage data. *Biostatistics (Oxford, England)*, 9(2):321–32, Apr 2008. doi: 10.1093/biostatistics/kxm030. URL <http://biostatistics.oxfordjournals.org/cgi/content/full/9/2/321>.

## 8 Session data

```
> sessionInfo()
```

```
R version 2.11.0 (2010-04-22)
```

```
x86_64-unknown-linux-gnu
```

```
locale:
```

```
[1] LC_CTYPE=en_US           LC_NUMERIC=C             LC_TIME=en_US
[4] LC_COLLATE=en_US        LC_MONETARY=C           LC_MESSAGES=en_US
[7] LC_PAPER=en_US          LC_NAME=C               LC_ADDRESS=C
[10] LC_TELEPHONE=C          LC_MEASUREMENT=en_US    LC_IDENTIFICATION=C
```

attached base packages:

```
[1] grid      tools      stats      graphics  grDevices  utils      datasets
[8] methods   base
```

other attached packages:

```
[1] edgeR_1.6.4          org.Sc.sgd.db_2.4.1  RSQLite_0.9-0
[4] DBI_0.2-5            AnnotationDbi_1.10.1 GenomeGraphs_1.8.0
[7] biomaRt_2.4.0        leeBamViews_0.99.7   BSgenome_1.16.1
[10] Rsamtools_1.0.1      Biostrings_2.16.0    GenomicRanges_1.0.1
[13] IRanges_1.6.1        Biobase_2.8.0
```

loaded via a namespace (and not attached):

```
[1] limma_3.4.0 Rcurl_1.4-2 XML_3.1-0
```