

Package ‘BiGGR’

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Type Package

Title Creates an interface to the BiGG database, provides a framework for simulation and produces flux graphs

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Depends R (>= 2.14.0), rsbml, hyperdraw, LIM

Imports hypergraph

Description This package provides an interface to simulate metabolic reconstruction from the BiGG database(<http://bigg.ucsd.edu/>) and other metabolic reconstruction databases. The package aids in performing flux balance analysis (FBA). Metabolic networks and estimated fluxes can be visualized using hypergraphs.

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biocViews NetworkAnalysis, Visualization, Metabolomics

LazyLoad yes

R topics documented:

BiGGR-package	2
buildSBMLFromBiGG	4
buildSBMLFromGenes	6
buildSBMLFromPathways	7
buildSBMLFromReactionIDs	9

createLIMFromBiGG	10
createLIMFromSBML	11
E.coli_iAF1260	13
E.coli_iJR904	14
E.coli_textbook	15
extractGeneAssociations	16
extractPathways	17
getPathwaysForSBML	18
getRates	19
Glycolysis	20
H.pylori_iIT341	20
H.sapiens_Recon_1	21
M.barkeri_iAF692	23
M.tuberculosis_iNJ661	24
P.putida_iJN746	25
Recon2	26
S.aureus_iSB619	27
S.cerevisiae_iND750	28
sbml2hyperdraw	29

Index 31

BiGGR-package	<i>Creates an interface to the BiGG database, provides a framework for simulation and produces flux graphs</i>
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Description

This package provides an interface to simulate metabolic reconstruction from the BiGG database(<http://bigg.ucsd.edu/>) and other metabolic reconstruction databases. The package aids in performing flux balance analysis (FBA). Metabolic networks and estimated fluxes can be visualized using hypergraphs.

Details

Package:	BiGGR
Type:	Package
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Depends:	R (>= 2.14.0), rsbml, hyperdraw, LIM
Imports:	hypergraph
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URL:	http://www.bioconductor.org/
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Index:

E.coli_iAF1260	Ecoli dataset with ORFs and thermodynamic information
E.coli_iJR904	Ecoli genome-scale model
Glycolysis	Metabolic reconstruction of Glycolysis pathway
H.pylori_ilt341	H.pylori in silico genome-scale characterization of sing
H.sapiens_Recon_1	Reconstruction of human metabolism from the BiGG databas
M.barkeri_iAF692	Metabolic reconstruction of M.barkeri from the BiGG data
Recon2	Human metabolic reconstruction Recon2
S.aureus_iSB619	Metabolic reconstruction of S.aureus from the BiGG databa
S.cerevisiae_iND750	Metabolic reconstruction of S.cerevisiae from the BiGG
buildSBMLFromBiGG	Build an SBML model from a given reactions file obtained
buildSBMLFromGenes	Build an SBML model for specific genes in a given databas
buildSBMLFromPathways	Build an SBML model for specific pathway(s) in a given d
buildSBMLFromReactionIDs	Build an SBML model for specific reactions in a given d
createLIMFromBiGG	Create a LIM model object from a BiGG database file
createLIMFromSBML	Create a LIM model object from an SBML file
extractGeneAssociations	Extract informations on genes from a given database
extractPathways	Extract all pathways from given database
getPathwaysForSBML	Extract all pathways from a database that are relevant f
getRates	Get Optimized Rates
sbml2hyperdraw	Returns a graph representation of an SBML model

Further information is available in the following vignettes:

BiGGR BiGGR (source, pdf)

Author(s)

Anand K. Gavai, Hannes Hettling

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See Also

rsbml

Examples

```
library("BiGGR")

library(help="BiGGR")

##load reaction identifiers from package examples
file.name <- system.file("extdata",
                        "Glycolysis_TCA_recon1_reactionIDs.txt",
                        package="BiGGR")
```

```

reaction.ids <- scan(file.name, what=" ")

##load database
data("H.sapiens_Recon_1")

##build SBML model
sbml.model <- buildSBMLFromReactionIDs(reaction.ids, H.sapiens_Recon_1)

##following term is to be maximized
maximize <- "R_ATPS4m - R_NDPK1m - R_HEX1 - R_PFK - R_PGK + R_PYK"

##specify the external metabolites of the system
externals <- c("M_glc_DASH_D_e", "M_lac_DASH_L_e",
              "M_ala_DASH_L_e", "M_gln_DASH_L_c", "M_h2o_e",
              "M_co2_e", "M_o2_e", "M_h_e", "M_pi_c",
              "M_o2s_m", "M_nh4_m", "M_adp_c",
              "M_atp_c", "M_nadp_c", "M_nadph_c", "M_h_c")

##specify the values of following fluxes:
##R_GLc1r=0.4, R_O2t=2.4, R_L_LAc2r=R_GLNtm=0

equation.vars <- c("R_GLc1r", "R_O2t", "R_L_LAc2r", "R_GLNtm")
equation.values <- c(0.4, 2.4, 0.0, 0.0)
eqns <- list(equation.vars, equation.values)

##create LIM file
limfile.name <- tempfile()
createLIMFromSBML(sbml.model, maximize, equations=eqns,
                 externals=externals, file.name=limfile.name)

rates <- getRates(limfile.name)

relevant.species <- c("M_glc_DASH_D_c", "M_g6p_c", "M_f6p_c",
                    "M_fdp_c", "M_dhap_c", "M_g3p_c",
                    "M_13dpg_c", "M_3pg_c", "M_2pg_c",
                    "M_pep_c", "M_pyr_c")

##generate graphical representation
hd <- sbml2hyperdraw(sbml.model, rates=rates,
                    relevant.species=relevant.species,
                    layoutType="dot", plt.margins=c(20, 0, 20, 0))

##plot hypergraph
plot(hd)

```

Description

Creates an SBML model containing all species, reactions and compartments that occur in a reactions file obtained from the BiGG database.

Usage

```
buildSBMLFromBiGG(reactions.filename, model.id=character(0), model.name=character(0))
```

Arguments

reactions.filename	name of the file containing the reactions extracted from BiGG
model.id	id for the SBML model created by the function. Defaults to reactions.filename
model.name	name for the SBML model created by the function. Defaults to reactions.filename

Value

a rsbml Model object containing all reactions, species and compartments that are associated with the reactions in the given input file.

Note

Note that it can be the case that species can be present in multiple compartments. In order to avoid any ambiguities in the model returned by the function, each species identifier is composed of the species identifier given in the reactions file and the compartment identifier, joined by "_". Example: adp in compartment c (cytosol) has the id adp_c.

Author(s)

Anand Gavai, Hannes Hettling

References

Schellenberger, J., Park, J. O., Conrad, T. C., and Palsson, B. , BiGG: a Biochemical Genetic and Genomic knowledgebase of large scale metabolic reconstructions, BMC Bioinformatics, 11:213, (2010). <http://bigg.ucsd.edu/>

See Also

[createLIMFromSBML](#)

Examples

```
##build model from file Reactions.txt from the package examples
path <- system.file("extdata", "Reactions.txt", package="BiGGR")
model <- buildSBMLFromBiGG(path, model.id="myid")
```

buildSBMLFromGenes *Build an SBML model for specific genes in a given database*

Description

Creates an SBML model containing all species, reactions and compartments that are associated with (a) specific gene(s) in the database document (e.g. Recon2) passed as an argument.

Usage

```
buildSBMLFromGenes(query, database, logical.fun="any")
```

Arguments

query	a character or a vector or list of character containing the query genes with identifiers as specified in the database.
database	an object of class SBMLDocument
logical.fun	function which specifies the logical relation of the query genes within the reactions (e.g. all or any, see details).

Details

The function all as argument logical.fun would mean that all genes in the query have to be associated with a certain reaction from the database in order to be included in the returned model. The default any means that a reaction is included if any of the query genes are associated with it. Custom functions are possible if they take a vector of type logical as an argument and return a logical. The argument of logical.fun is a vector of type logical having the same length as the query and for each gene the value is TRUE if it is associated with a specific reaction. See 'examples' section for an example of a custom function as logical.fun.

Value

a rsbml Model object containing all reactions, species and compartments that are present in the database and are associated with the query gene(s) or NULL if none of the genes in the database match the query.

Note

If the reactions in the database document provided in the argument database do not contain any "<notes>" with tags with gene information indicated by the string "GENE*ASSOCIATION" (the star stands for any character), no gene association information can be extracted and thus the returned SBML model is empty..

Author(s)

Anand Gavai, Hannes Hettling

References

Thiele, I. et al. Nat Biotech, 2013

See Also

[buildSBMLFromPathways extractGeneAssociations](#)

Examples

```
##Query genes in Recon 2 database
data("Recon2")
database <- Recon2
m1 <- buildSBMLFromGenes("8884.1", database)
m2 <- buildSBMLFromGenes(c("8884.1", "6509.1"), database)

##different databases
data(H.pylori_ilt341)
database <- H.pylori_ilt341
m3 <- buildSBMLFromGenes("HP0069", database)

data(M.barkeri_iAF692)
database <- M.barkeri_iAF692
m4 <- buildSBMLFromGenes(c("MBd0456", "MBd4814", "MBd4098"), database)

data(S.aureus_iSB619)
database <- S.aureus_iSB619
m5 <- buildSBMLFromGenes(c("SA0594", "SA1599", "SA0950", "SA0259"), database)

database <- Recon2
query <- c("218.1", "223.1")
m6 <- buildSBMLFromGenes(query, database)
m7 <- buildSBMLFromGenes(query, database, logical.fun="all")
##m6 has more reactions than m7
## because m7 has only reactions which match both genes in the query
length(m6@reactions) > length(m7@reactions)

##Custom logical function: Get model with all reactions
## which are not associated with the query gene
m8 <- buildSBMLFromGenes(query, database, logical.fun=function(x)!any(x))
```

buildSBMLFromPathways *Build an SBML model for specific pathway(s) in a given database*

Description

Creates an SBML model containing all species, reactions and compartments that are part of (a) given pathway(s) in the database document (e.g. Recon2) passed as an argument.

Usage

```
buildSBMLFromPathways(query, database, match.exact=TRUE)
```

Arguments

query	a character or a vector or list of character containing the names of the query pathways
database	an object of class SBMLDocument
match.exact	logical whether only the exact pathway name should be matched or whether a pathway should match if one keyword is in the pathway description in the database.

Value

a rsbml Model object containing all reactions, species and compartments that are present in the database for the query pathway(s) or NULL if none of the pathways in the database match the query.

Note

If the reactions in the database document provided in the argument database do not contain any "<notes>" with tags with pathway information indicated by the string "SUBSYSTEM", no pathway information can be extracted and thus the SBML model returned will be empty.

Author(s)

Anand Gavai, Hannes Hettling

References

Thiele, I. et al. Nat Biotech, 2013

See Also

[extractPathways](#)

Examples

```
data("Recon2")
database <- Recon2

##Get Model for specific pathway
m1 <- buildSBMLFromPathways("Arginine and Proline Metabolism", database)
```

```
##Get Model for specific pathway "Metabolism": does not exist!
m2 <- buildSBMLFromPathways("Metabolism", database)

##Get model of all pathways which contain keyword "metabolism"
m3 <- buildSBMLFromPathways("Metabolism", database, match.exact=FALSE)

##Multi-query:
query <- c("Transport, endoplasmic reticular", "Arginine and Proline Metabolism")
m4 = buildSBMLFromPathways(query, database)
m5 = buildSBMLFromPathways(query[1], database)
length(m4@species)
length(m5@species)

##different database
data(H.pylori_ilt341)
database <- H.pylori_ilt341
m7 <- buildSBMLFromPathways("Metabolism", database, match.exact=FALSE)
```

buildSBMLFromReactionIDs

Build an SBML model for specific reactions in a given database

Description

Creates an SBML model containing all species, reactions and compartments that are associated with a number of reaction identifiers in the database document (e.g. Recon2) passed as an argument.

Usage

```
buildSBMLFromReactionIDs(reaction.ids, database)
```

Arguments

reaction.ids	a character or a vector or list of character containing the names of the query reaction IDs
database	an object of class SBMLDocument

Value

a rsbml Model object containing all reactions, species and compartments that are present in the database for the query reaction(s) or NULL if none of the query reaction IDs is found in the database.

Author(s)

Anand Gavai, Hannes Hettling

References

Thiele, I. et al. Nat Biotech, 2013

See Also

[buildSBMLFromGenes](#)

Examples

```
##get list of reactions with Recon 2 identifiers from examples
path <- system.file("extdata", "Glycolysis_TCA_recon2_reactionIDs.txt", package="BiGGR")
reaction.ids <- scan(path, what=" ")

data("Recon2")
model <- buildSBMLFromReactionIDs(reaction.ids, Recon2)
```

createLIMFromBiGG *Create a LIM model object from a BiGG database file*

Description

Creates a LIM model object from a file containing reactions extracted from BiGG to be run for simulations of metabolic fluxes

Usage

```
createLIMFromBiGG(reactions.filename, ...)
```

Arguments

```
reactions.filename      file which contains the reactions extracted from the BiGG database
...                      arguments passed to createLIMfromSBML
```

Note

none

Author(s)

Anand K. Gavai <anand.gavai@bioinformatics.nl>, Hannes hettling <j.hettling@vu.nl>

References

Soetaert K, van Oevelen D (2009). LIM: Linear Inverse Model examples and solution methods. R package version 1.3

See Also[createLIMFromSBML](#)**Examples**

```
##maximize flux for reaction R_PYK
maximize <- "R_PYK"

##setting equality constraint R_HEX = 1
equation_var <- "R_HEX1"
equation_value <- 1
eq <- list(equation_var, equation_value)

##range of possible fluxes for R_PYK
constraint <- list("R_PYK", 0, 1000)
externals <- c("glc_c", "pyr_c", "h_c", "nad_c",
              "nadh_c", "pi_c", "fad_m", "fadh2_m",
              "o2_c", "adp_c", "atp_c", "nadp_c",
              "co2_c", "o2_c", "gdp_c", "gtp_c")

##build LIM model from reactions file in package examples
path <- system.file("extdata", "Reactions.txt", package="BiGGR")
limfile.name <- tempfile()
createLIMFromBiGG(path, maximize, equations=eq, constraints=constraint,
                  externals=externals, file.name=limfile.name)
```

createLIMFromSBML *Create a LIM model object from an SBML file*

Description

creates a model file to be run for simulations of metabolic fluxes

Usage

```
createLIMFromSBML(model, maximize, equations=NULL,
                  constraints=NULL, externals=NULL, file.name="model.lim")
```

Arguments

model	an SBML object of reactions/metabolites participating in a metabolic pathway.
maximize	a character vector consisting the tag of the reaction(s) to be maximized or minimized
equations	a list specifying equality constraints on the system. The list must have two entries, the first one being a vector of class character containing the left hand side(s) of the equation(s), the second one being a vector of type character or numeric with the right hand side(s) of the equation(s). See also 'examples'.

constraints	a list specifying constrained on the solution space of the flux vector. The list must have three entries, the first one being a vector of class character with the reaction id(s) to be constrained, the second and third one a numeric vector with the lower and upper flux bounds, respectively, for the reactions to be constrained. is a character vector specifying the minimum and maximum values(boundary) under which the solution for the maximize reaction should fall
externals	a character vector of metabolites as provided by the user for specific pathways for which FBA (flux balance analysis needs to be performed)
file.name	a character string specifying the name of the LIM file created by the function.

Value

A model file with with extension ".lim" is created

Note

none

Author(s)

Anand K. Gavai <anand.gavai@bioinformatics.nl>, Hannes Hettling <j.hettling@vu.nl>

References

Soetaert K, van Oevelen D (2009). LIM: Linear Inverse Model examples and solution methods. R package version 1.3

Examples

```
##Create a LIM model file from a reactions file in the examples
path <- system.file("extdata", "Glycolysis_TCA_recon1_reactionIDs.txt", package="BiGGR")
reaction.ids <- scan(path, what=" ")
data("H.sapiens_Recon_1")
sbml.model <- buildSBMLFromReactionIDs(reaction.ids, H.sapiens_Recon_1)
maximize <- c("R_ATPS4m - R_NDPK1m - R_HEX1 - R_PFK - R_PGK + R_PYK")
externals <- c("M_glc_DASH_D_e", "M_lac_DASH_L_e",
              "M_ala_DASH_L_e", "M_gln_DASH_L_c", "M_h2o_e",
              "M_co2_e", "M_o2_e", "M_h_e", "M_pi_c",
              "M_o2s_m", "M_nh4_m", "M_adp_c",
              "M_atp_c", "M_nadp_c", "M_nadph_c", "M_h_c")
equation.vars <- c("R_GLc1r", "R_O2t", "R_L_LAcT2r", "R_GLNtm")
equation.values <- c(0.4, 2.4, 0.0, 0.0)
eqns <- list(equation.vars, equation.values)
constraints <- list(c("R_GLc1r", "R_CY00m3"), c(-1000, -1000), c(1000, 1000))
limfile.name <- tempfile()
createLIMFromSBML(sbml.model, maximize, equations=eqns,
                  constraints=constraints, externals=externals, file.name=limfile.name)
```

E.coli_iAF1260

Ecoli dataset with ORFs and thermodynamic information

Description

A genome-scale metabolic reconstruction for Escherichia coli K-12 MG1655 that accounts for 1260 ORFs and thermodynamic information. The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggs/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(E.coli_iAF1260)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggs/exportSelect.pl>

References

Feist, A.M., Henry, C.S., Reed, J.L., Krummenacker, M., Joyce, A.R., Karp, P.D., Broadbelt, L.J., Hatzimanikatis, V., Palsson, B.O., *A genome-scale metabolic reconstruction for Escherichia coli K-12 MG1655 that accounts for 1260 ORFs and thermodynamic information*, *Molecular Systems Biology*, 3:121 (2007)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggs/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\\\".+?\\\"", "", string)
E.coli_iAF1260 <- rsbml_read(text=string)

## End(Not run)
```

```
##load data and get all reaction IDs
data(E.coli_iAF1260)
model <- E.coli_iAF1260@model
##get all reaction identifiers
sapply(model@reactions, id)
```

E.coli_iJR904

Ecoli genome-scale model

Description

An expanded genome-scale model of Escherichia coli K-12 (iJR904 GSM/GPR). The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/bigg/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(E.coli_iJR904)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/bigg/exportSelect.pl>

References

Reed, J.L., Vo, T.D., Schilling, C.H., and Palsson, B.O., *An expanded genome-scale model of Escherichia coli K-12 (iJR904 GSM/GPR)*, *Genome Biology*, 4(9): R54.1-R54.12 (2003).

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggy/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\\.+?\\", "", string)
E.coli_iJR904 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(E.coli_iJR904)
model <- E.coli_iJR904@model
##get all reaction identifiers
sapply(model@reactions, id)
```

E.coli_textbook

Ecoli dataset from the BiGG database

Description

A metabolic reconstruction for Escherichia from text books. The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggy/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(E.coli_textbook)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggy/exportSelect.pl>

References

Feist, A.M., Henry, C.S., Reed, J.L., Krummenacker, M., Joyce, A.R., Karp, P.D., Broadbelt, L.J., Hatzimanikatis, V., Palsson, B.O., *A genome-scale metabolic reconstruction for Escherichia coli K-12 MG1655 that accounts for 1260 ORFs and thermodynamic information*, *Molecular Systems Biology*, 3:121 (2007)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggs/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\".+?\"", "", string)
E.coli_textbook <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(E.coli_textbook)
model <- E.coli_textbook@model
##get all reaction identifiers
sapply(model@reactions, id)
```

extractGeneAssociations

Extract informations on genes from a given database

Description

Extracts all information on genes associated to reactions from an rsbml document containing a metabolic reconstruction database (e.g. Recon2). The associated information is parsed from the "<notes>" tag of each reaction's SBML representation.

Usage

```
extractGeneAssociations(database)
```

Arguments

database an object of class [SBMLDocument](#)

Value

a list with length being the number of reactions in the database passed as argument each entry containing a character containing the associated gene identifiers and the reaction IDs as names. For reactions without gene annotation, the list will contain NA.

Note

If the reactions in the database document provided in the argument database do not contain any "<notes>" with tags with gene information indicated by the string "GENE*ASSOCIATION" (the star stands for any character), no gene association information can be extracted and thus the returned SBML mdel is empty..

Author(s)

Anand Gavai, Hannes Hettling

References

Thiele, I. et al. Nat Biotech, 2013

See Also

[buildSBMLFromGenes](#)

Examples

```
data("Recon2")
database <- Recon2
gene.info <- extractGeneAssociations(database)
```

extractPathways

Extract all pathways from given database

Description

Extracts all pathway information from an rsbml document containing a metabolic reconstruction database (e.g. Recon2). The pathway information is parsed from the "<notes>" tag of each reaction.

Usage

```
extractPathways(database)
```

Arguments

database an object of class [SBMLDocument](#)

Value

a list with length being the number of reactions in the database passed as argument each entry containing a character with the pathway information and the reaction IDs as names. For reactions without pathway annotation, the list will contain NA.

Note

If the reactions in the database document provided in the argument database do not contain any "<notes>" with tags with pathway information indicated by the string "SUBSYSTEM", no pathway information can be extracted.

Author(s)

Anand Gavai, Hannes Hettling

References

Thiele, I. et al. Nat Biotech, 2013

See Also

[buildSBMLFromPathways](#) [getPathwaysForSBML](#)

Examples

```
data(Recon2)
pathways.recon2 <- extractPathways(Recon2)
```

getPathwaysForSBML	<i>Extract all pathways from a database that are relevant for a given SBML model</i>
--------------------	--

Description

Extracts all pathway information from an rsbml document containing a metabolic reconstruction database (e.g. Recon2) and returns the subset of these pathways that is associated with the reactions in the given model. The pathway information is parsed from the "<notes>" tag of each reaction.

Usage

```
getPathwaysForSBML(model, database)
```

Arguments

database	an rsbml object of class SBMLDocument
model	an rsbml object of class Model

Value

A vector of type character that contains all the pathways relevant for the given model according to the specified database. Note that duplicate pathways do not appear twice in the return value.

Note

If the reactions in the database document provided in the argument database do not contain any "<notes>" with tags with pathway information indicated by the string "SUBSYSTEM", no pathway information can be extracted.

Author(s)

Anand Gavai, Hannes Hettling

References

Thiele, I. et al. Nat Biotech, 2013

See Also

[buildSBMLFromPathways](#) [buildSBMLFromGenes](#) [extractPathways](#)

Examples

```
##Build a model from query genes
data("Recon2")
database <- Recon2
query <- c("218.1", "223.1") ##query gene identifiers
m <- buildSBMLFromGenes(query, database)

##extract all pathways for that model
getPathwaysForSBML(m, database)
```

getRates

Get Optimized Rates

Description

getRates takes the model file as the argument and based on the description of the model file generates flux values for "minimum" or "maximum" reaction rates

Usage

```
getRates(modelFile)
```

Arguments

modelFile A model file as generated from the createBiggModel function

Value

The value returned is one dimensional numeric vector of flux rates for each reaction

Author(s)

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Examples

```
data("Glycolysis")
rates <-getRates(Glycolysis)
rates
```

Glycolysis	<i>Metabolic reconstruction of Glycolysis pathway</i>
------------	---

Description

Model of Glycolysis pathway

Usage

Glycolysis

Format

A LIM model file created as an example

H.pylori_iIT341	<i>H.pylori in silico genome-scale characterization of single and double deletion mutants</i>
-----------------	---

Description

An Expanded Metabolic Reconstruction of Helicobacter pylori (iIT341 GSM/GPR): An in silico genome-scale characterization of single and double deletion mutants. The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggy/exportSelect.pl>) which was subsequently converted into an object of class SBML using the rsbml_read function from the rsbml package.

Usage

```
data(H.pylori_iIT341)
```

Format

An sbml object of class rsbml

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggy/exportSelect.pl>

References

Thiele, I., Vo, T.D., Price, N.D. and Palsson, B., *An Expanded Metabolic Reconstruction of Helicobacter pylori* (iT341 GSM/GPR): An in silico genome-scale characterization of single and double deletion mutants, *Journal of Bacteriology*, 187(16): 5818-5830 (2005)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggy/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\".+?\"", "", string)
H.pylori_iIT341 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(H.pylori_iIT341)
model <- H.pylori_iIT341@model
##get all reaction identifiers
sapply(model@reactions, id)
```

Description

The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggy/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(H.sapiens_Recon_1)
```

Format

An sbml object of class rsbml

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggy/exportSelect.pl>

References

Duarte, N.D., Becker, S. A., Jamshidi, N., Thiele, I., Mo, M. L., Vo, T. D., Srivas, R., Palsson, B. O., Global reconstruction of the human metabolic network based on genomic and bibliomic data, Proc. Nat Acad. Sci. 104(6):1777-82 (2007)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggy/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\".+?\"", "", string)
H.sapiens_Recon_1 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(H.sapiens_Recon_1)
model <- H.sapiens_Recon_1@model
##get all reaction identifiers
sapply(model@reactions, id)
```

M.barkeri_iAF692

Metabolic reconstruction of M.barkeri from the BiGG database

Description

The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggy/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(M.barkeri_iAF692)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggy/exportSelect.pl>

References

Feist, A.M., Scholten, J.C.M., Palsson, B.O., Brockman, F.J., and Ideker, T., "Modeling methanogenesis with a genome-scale metabolic reconstruction of *Methanosarcina barkeri*", *Molecular Systems Biology*, 2(1):msb4100046-E1-E14 (2006)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggy/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\".+?\"", "", string)
M.barkeri_iAF692 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(M.barkeri_iAF692)
```

```
model <- M.barkeri_iAF692@model
##get all reaction identifiers
sapply(model@reactions, id)
```

M.tuberculosis_iNJ661 *Metabolic reconstruction of M.tuberculosis from the BiGG database*

Description

A metabolic reconstruction for tuberculosis. The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/bigg/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(M.tuberculosis_iNJ661)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

```
http://bigg.ucsd.edu/bigg/exportSelect.pl
```

References

Feist, A.M., Henry, C.S., Reed, J.L., Krummenacker, M., Joyce, A.R., Karp, P.D., Broadbelt, L.J., Hatzimanikatis, V., Palsson, B.O., *A genome-scale metabolic reconstruction for Escherichia coli K-12 MG1655 that accounts for 1260 ORFs and thermodynamic information*, *Molecular Systems Biology*, 3:121 (2007)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/bigg/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
```

```
string <- gsub("units=\\.+?\\\"", "", string)
M.tuberculosis_iNJ661 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(M.tuberculosis_iNJ661)
model <- M.tuberculosis_iNJ661@model
##get all reaction identifiers
sapply(model@reactions, id)
```

P.putida_iJN746

Metabolic reconstruction of P.putida from the BiGG database

Description

A metabolic reconstruction for *P. putida*. The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggs/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(P.putida_iJN746)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggs/exportSelect.pl>

References

Feist, A.M., Henry, C.S., Reed, J.L., Krummenacker, M., Joyce, A.R., Karp, P.D., Broadbelt, L.J., Hatzimanikatis, V., Palsson, B.O., *A genome-scale metabolic reconstruction for Escherichia coli K-12 MG1655 that accounts for 1260 ORFs and thermodynamic information*, *Molecular Systems Biology*, 3:121 (2007)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggs/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\\.+?\\", "", string)
P.putida_iJN746 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(P.putida_iJN746)
model <- P.putida_iJN746@model
##get all reaction identifiers
sapply(model@reactions, id)
```

Recon2

Human metabolic reconstruction Recon2

Description

The dataset was generated by downloading the SBML file of the reconstruction (<http://www.ebi.ac.uk/biomodels-main/MODEL1109130000>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(Recon2)
```

Format

An sbml object of class `rsbml`

Source

<http://www.ebi.ac.uk/biomodels-main/MODEL1109130000>

References

Thiele I, Swainston N, et al., "A community-driven global reconstruction of human metabolism", *Nature Biotechnology* 31, 419-425 (2013), doi:10.1038/nbt.2488

Examples

```
## Not run:
##The dataset was generated as follows:
##MODEL1109130000.xml was downloaded from http://www.ebi.ac.uk/biomodels-main/MODEL1109130000
Recon2 <- rsbml_read("MODEL1109130000.xml")

## End(Not run)

##load data and get all reaction IDs
data(Recon2)
model <- Recon2@model
##get all reaction identifiers
sapply(model@reactions, id)
```

S.aureus_iSB619

Metabolic reconstruction of S.aureus from the BiGG database

Description

The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/bigg/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(S.aureus_iSB619)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/bigg/exportSelect.pl>

References

Becker, S.A. and Palsson, B.O., Genome-scale reconstruction of the metabolic network in *Staphylococcus aureus* N315: an initial draft to the two-dimensional annotation, *BMC Microbiology*, 5(1):8 (2005)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggs/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\\.+?\\", "", string)
S.aureus_iSB619 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(S.aureus_iSB619)
model <- S.aureus_iSB619@model
##get all reaction identifiers
sapply(model@reactions, id)
```

S.cerevisiae_iND750 *Metabolic reconstruction of S.cerevisiae from the BiGG database*

Description

The dataset was generated by downloading the SBML file of the reconstruction (<http://bigg.ucsd.edu/biggs/exportSelect.pl>) which was subsequently converted into an object of class SBML using the `rsbml_read` function from the `rsbml` package.

Usage

```
data(S.cerevisiae_iND750)
```

Format

An sbml object of class `rsbml`

Details

Note that the files in the BiGG database fail the unit consistency check of the `rsbml_read` function. To avoid unit checking when creating SBML objects, the substance units in the reaction tags were parsed out from the database SBML files (see example below).

Source

<http://bigg.ucsd.edu/biggs/exportSelect.pl>

References

Duarte, N.C., Herrgard, M.J., and Palsson, B.O., " Reconstruction and Validation of Saccharomyces cerevisiae iND750, a Fully Compartmentalized Genome-scale Metabolic Model", Genome Research, 14: 1298-1309 (2004)

Examples

```
## Not run:
##The dataset was generated as follows:
##SBML_export.xml was downloaded from http://bigg.ucsd.edu/biggy/exportSelect.pl
##and a newline was added at the end of the file
file <- "SBML_export.xml"
string <- paste(readLines(file), collapse="\n")
##Parse out units to avoid validation error
string <- gsub("units=\".+?\"", "", string)
S.cerevisiae_iND750 <- rsbml_read(text=string)

## End(Not run)

##load data and get all reaction IDs
data(S.cerevisiae_iND750)
model <- S.cerevisiae_iND750@model
##get all reaction identifiers
sapply(model@reactions, id)
```

sbml2hyperdraw

Returns a graph representation of an SBML model

Description

Convert an SBML model to a RagraphBPH using hypergraph. Metabolites are displayed as nodes and reactions are displayed as directed edges connecting the nodes. If a vector of rates is given, edge widths are weighted according to the rates. For negative rates, edges are drawn in red and the arrow between the metabolites is reversed to represent the correct direction of the flux.

Usage

```
sbml2hyperdraw(sbml.model, rates ,relevant.species,
relevant.reactions,layoutType, lwd.max, lwd.min, plt.margins)
```

Arguments

sbml.model	an rsbml Model object
rates	a named vector with the rates of the reactions in the model. The names of the rates must agree with the reaction identifiers in the sbml.model

<code>relevant.species</code>	a vector of type character defining a subset of species in the <code>sbml.model</code> to be plotted. Defaults to all species identifiers in the <code>sbml.model</code> .
<code>relevant.reactions</code>	a vector of type character defining a subset of reactions in the <code>sbml.model</code> to be plotted. Defaults to all reactions identifiers in the <code>sbml.model</code> .
<code>layoutType</code>	is a character string representing the layout engine to be used for visualization. Current supported layouts are "dot", "twopi", "neato", "fdp", "sfdp" and "circo". Defaults to "dot". See <code>?GraphvizLayouts</code> for further documentation.
<code>lwd.max</code>	a numeric given the maximum edge width. Defaults to 3.
<code>lwd.min</code>	a numeric given the minimum edge width. Defaults to 0.5.
<code>plt.margins</code>	A numerical vector of the form <code>c(bottom, left, top, right)</code> giving additional white space around the graph (in case long node or edge labels fall outside the plotting region). Defaults to <code>c(150,150,150,150)</code> .

Value

Object of class `RagraphBPH` with the hypergraph representation of the SBML object.

Author(s)

Hannes Hettling <j.hettling@vu.nl>, Anand K. Gavai <anand.gavai@bioinformatics.nl>

See Also

`RagraphBPH hyperdraw`

Examples

```
##Generate an example model
path <- system.file("extdata", "Glycolysis_TCA_recon2_reactionIDs.txt", package="BiGGR")
reaction.ids <- scan(path, what=" ")

data("Recon2")
model <- buildSBMLFromReactionIDs(reaction.ids, Recon2)

##Plot ATP and ADP in cytosol and mitochondrion in model without rates
rel.sp <- c("M_adp_c", "M_atp_c", "M_adp_m", "M_atp_m")
hd <- sbml2hyperdraw(model, relevant.species=rel.sp)
plot(hd)

##Plot model with random rates
rates <- rnorm(length(model@reactions))
names(rates) <- sapply(model@reactions, id)
hd <- sbml2hyperdraw(model, rates=rates, relevant.species=rel.sp, lwd.max=4)
plot(hd)
```

Index

- *Topic **BiGG**
 - buildSBMLFromBiGG, 4
 - *Topic **Linear Inverse Models**
 - createLIMFromBiGG, 10
 - createLIMFromSBML, 11
 - *Topic **Linear Optimization**
 - getRates, 19
 - *Topic **Linear optimization model file**
 - createLIMFromBiGG, 10
 - createLIMFromSBML, 11
 - *Topic **SBML object**
 - createLIMFromSBML, 11
 - *Topic **biggr**
 - BiGGR-package, 2
 - *Topic **ecoli**
 - E.coli_iAF1260, 13
 - E.coli_iJR904, 14
 - E.coli_textbook, 15
 - *Topic **flux rates**
 - getRates, 19
 - *Topic **gene**
 - buildSBMLFromGenes, 6
 - extractGeneAssociations, 16
 - *Topic **glycolysis**
 - Glycolysis, 20
 - *Topic **hpylori**
 - H.pylori_ilT341, 20
 - *Topic **hsapiens**
 - H.sapiens_Recon_1, 21
 - *Topic **hyperdraw**
 - sbml2hyperdraw, 29
 - *Topic **hypergraph**
 - sbml2hyperdraw, 29
 - *Topic **mbarkeri**
 - M.barkeri_iAF692, 23
 - *Topic **pathways**
 - buildSBMLFromPathways, 7
 - *Topic **pathway**
 - extractPathways, 17
 - getPathwaysForSBML, 18
 - *Topic **putida**
 - P.putida_iJN746, 25
 - *Topic **reactions**
 - buildSBMLFromReactionIDs, 9
 - *Topic **recon2**
 - Recon2, 26
 - *Topic **saureus**
 - S.aureus_iSB619, 27
 - *Topic **scervisiae**
 - S.cerevisiae_iND750, 28
 - *Topic **tuberculosis**
 - M.tuberculosis_iNJ661, 24
- BiGGR (BiGGR-package), 2
- BiGGR-package, 2
- buildSBMLFromBiGG, 4
- buildSBMLFromGenes, 6, 10, 17, 19
- buildSBMLFromPathways, 7, 7, 18, 19
- buildSBMLFromReactionIDs, 9
- createLIMFromBiGG, 10
- createLIMFromSBML, 5, 11, 11
- E.coli_iAF1260, 13
- E.coli_iJR904, 14
- E.coli_textbook, 15
- extractGeneAssociations, 7, 16
- extractPathways, 8, 17, 19
- getPathwaysForSBML, 18, 18
- getRates, 19
- Glycolysis, 20
- H.pylori_ilT341, 20
- H.sapiens_Recon_1, 21
- M.barkeri_iAF692, 23
- M.tuberculosis_iNJ661, 24
- Model, 18

P.putida_iJN746, [25](#)

Recon2, [26](#)

S.aureus_iSB619, [27](#)

S.cerevisiae_iND750, [28](#)

sbml2hyperdraw, [29](#)

SBMLDocument, [6](#), [8](#), [9](#), [16–18](#)