Package 'SNPchip'

September 24, 2012

Version 2.2.0

Title Visualizations for copy number alterations

Author Robert Scharpf <rscharpf@jhsph.edu> and Ingo Ruczinski

Maintainer Robert Scharpf <rscharpf@jhsph.edu>

Depends R (>= 2.14.0)

Imports graphics, lattice, grid, foreach, IRanges, utils, methods,oligoClasses (>= 1.17.25), Biobase

Suggests crlmm

Enhances doSNOW, VanillaICE, RColorBrewer

Description This package defines methods for visualizing high-throughput genomic data

License LGPL (>= 2)

LazyLoad yes

Collate AllGenerics.R coerce-methods.R xyplot-methods.R
grid-functions.R idiogram-functions.R panel-functions.R zzz.R

biocViews CopyNumberVariants, SNP, GeneticVariability, Visualization

URL http://www.biostat.jhsph.edu/~iruczins/software/snpchip.html

R topics documented:

arrangeSideBySide *Helper function to arrange two trellis objects side by side on a grid.*

Description

For visualizing copy number alterations, it is often helpful to plot estimates of copy number along with the corresponding estimate of the B allele frequencies. Creating a trellis object for the copy number estimates and a separate trellis object for the B allele frequencies, this function can be used to arrange the two trellis objects side by side on a grid.

Usage

```
arrangeSideBySide(object1, object2)
```

Arguments

object1	A trellis object (e.g., a trellis object of the copy number estimates).
object2	A trellis object (e.g., a trellis object of the B allele frequencies).

Author(s)

Rob Scharpf

See Also

xypanel, xyplot

centromere *Coordinates of centromere*

Description

Extracts coordinates of centromere for a particular chromosome

Usage

```
centromere(chromosome, build="hg18", verbose=FALSE)
```

Arguments

chromosome	character string: "1", ..., "22", "X", or "Y"
build	character string. Currently only build 'hg18' is provided
verbose	Logical. Displays build used to annotate the centromere coordinates when TRUE

Value

integer: start and stop coordinates of centromere in basepairs

Author(s)

R. Scharpf

Examples

```
centromere("1")
```

---

chromosomeAnnotation *chromosome annotation*

Description

Contains information on chromosomes 1-22, X and Y.

Usage

```
data(chromosomeAnnotation)
```

Format

A data frame with 24 observations on the following 3 variables.

`centromereStart` a numeric vector

`centromereEnd` a numeric vector

`chromosomeSize` a numeric vector

Examples

```
data(chromosomeAnnotation)
```

---

cytoband *Start and stop sites of cytoband*

Description

Contains start and stop sites of cytoband for Build 35

Usage

```
data(cytoband)
```

Format

A data frame with 862 observations on the following 5 variables.

`chrom` a factor with levels `chr1` `chr10` `chr11` `chr12` `chr13` `chr14` `chr15` `chr16` `chr17` `chr18` `chr19` `chr2` `chr20` `chr21` `chr22` `chr3` `chr4` `chr5` `chr6` `chr7` `chr8` `chr9` `chrX` `chrY`

`chromStart` a numeric vector

`chromEnd` a numeric vector

`name` a factor indicating which chromosomal arm

`gieStain` a factor with levels `acen` `gneg` `gpos100` `gpos25` `gpos50` `gpos75` `gvar` `stalk`

Source

http://pevsnerlab.kennedykrieger.org/snpscan_07_sourcecode.htm

Examples

```
data(cytoband)
```

`plotIdiogram` *Plots idiogram for one chromosome*

Description

Draw an idiogram for the specified chromosome.

Usage

```
plotIdiogram(chromosome, cytoband, cytoband.ycoords, xlim, ylim=c(0, 2),
new=TRUE, label.cytoband=TRUE, label.y=NULL, srt, cex.axis=1,
outer=FALSE, taper=0.15, verbose=FALSE, build="hg18", unit=c("bp", "Mb"), is.lattice=FALSE,...)
```

Arguments

`chromosome`	character string or integer: which chromosome to draw the cytoband
`cytoband`	data.frame containing cytoband information
`cytoband.ycoords`	numeric: y coordinates
`xlim`	x-axis limits
`ylim`	y-axis limits
`new`	logical: new plotting device
`label.cytoband`	logical: if TRUE, labels the cytobands
`label.y`	numeric: height (y-coordinate) for cytoband label
`srt`	string rotation for cytoband labels. See `par`
`cex.axis`	size of cytoband labels. See par
`outer`	logical: whether to draw the labels in the outer margins. See par
`taper`	tapering for the ends of the cytoband
`verbose`	Logical. If TRUE, displays human genome build used to annotated the cytoband coordinates.
`build`	Character string. Currently only "hg18" is allowed.
`unit`	Character string indicating the unit for physical position on the x-axis. Available options are basepairs (bp) or Mb.
`is.lattice`	logical indicating whether your drawing the cytoband on a lattice graphic.
`...`	additional arguments to plot

Author(s)

Robert Scharpf and Jason Ting

Examples

```
plotIdiogram("1")
```

xypanel *A panel function for plotting copy number versus physical position*

Description

A panel function for xyplot for plotting copy number versus physical position.

Usage

```
xypanel(x, y, gt, is.snp, range, col.hom = "grey20", fill.hom =
"lightblue", col.het = "grey20", fill.het = "salmon", col.np = "grey20",
fill.np = "grey60", show.state=TRUE, state.cex=1,  col.state="blue", ..., subscripts)
```

Arguments

x	Physical position in megabases.
y	Copy number estimates.
gt	Genotype calls.
is.snp	Logical. Whether the marker is polymorphic.
range	A RangedData or IRanges object. Note that we expect the units returned by start and end to be basepairs.
col.hom	A specification for the color of plotting symbols for homozygous genotypes.
fill.hom	A specification for the fill color of plotting symbols for homozygous genotypes.
col.het	A specification for the color of plotting symbols for heterozygous genotypes.
fill.het	A specification for the fill color of plotting symbols for heterozygous genotypes.
col.np	A specification for the color of plotting symbols for nonpolymorphic markers.
fill.np	A specification for the fill color of plotting symbols for nonpolymorphic genotypes.
show.state	Logical. Whether to display the predicted state in each panel.
state.cex	Numeric. cex for state label. Ignored if show.state is FALSE.
col.state	Character. color for state label. Ignored if show.state is FALSE.
...	Additional arguments passed to lattice functions xyplot, lpoints, and lrect.
subscripts	See the panel functions in lattice for more information.

Details

The order of plotting is (1) nonpolymorphic markers, (2), homozygous SNPs, and (3) heterozygous SNPs. Stretches of homozygosity should appear as blue using the default color scheme.

Note

To make the drawing of the range object border invisible, one can use border="white".

Author(s)

R. Scharpf

See Also

xyplot

Examples

```
## Not run:
if(require("crlmm") && require("VanillaICE") && require("IRanges")){
library(oligoClasses)
data(cnSetExample, package="crlmm")
cnSetExample <- chromosomePositionOrder(cnSetExample)
oligoSet <- as(cnSetExample, "oligoSnpSet")
fit2 <- hmm(oligoSet, p.hom=1)
xyplot(cn ~ x | range, data=oligoSet, range=fit2[1:10, ],
       frame=2e6,
       panel=xypanel, cex=0.3, pch=21, border="blue",
       scales=list(x="free"),
       col.hom="lightblue", col.het="salmon", col.np="grey60",
       fill.np="grey60",
       xlab="Mb")
## if xyplot method is masked by lattice, do
##xyplot <- VanillaICE:::xyplot
}

## End(Not run)
```

---

xyplot *Plot copy number and physical position for a set of genomic intervals.*

Description

Plot copy number and physical position given by a CNSet object for a set of genomic intervals stored in a RangedDataCVN object.

Usage

```
xyplot(x, data, ...)
xyplot2(x, data, range, frame=50e3L, ...)
```

Arguments

x	A formula. Currently, the formula must be one of cn~x, cn ~ x | id or cn ~ x | range when data is a CNSet. If data is a BeadStudioSet, the formula has the form lrr ~ x| range or baf ~ x | range.
data	A CNSet, BeadStudioSet, or SnpSet object.
...	A RangedDataCNV object must be passed by the name 'range'. Arguments for xyplot are passed to xyplot2. Additional arguments are passed to xypanel and panel.xyplot.
range	A RangedDataCNV object.
frame	The genomic distance (basepairs) to the left and right of the start and stop coordinates in the range object.

Details

For a given RangedDataCNV object, this function will plot the copy number estimates versus physical position. The function is particularly useful for multi-panel displays in which the copy number estimates for a single range of the RangedDataCNV object appears in one panel. The size of the multi-panel display depends on the number of ranges (rows) in the RangedDataCNV object. Typically, one would want to pass no more than 10 ranges to the xyplot function.

For genomic intervals of interest in the RangedDataCNV, it is often helpful to 'frame' the interval by plotting the data surrounding the interval. To facilitate this process, one may pass an argument called frame (an integer) that indicates the number of basepairs to the left and right of the start / stop points in the interval. By default, the first interval in the RangedDataCNV object will be plotted in the lower left panel and the last interval RangedDataCNV object will be plotted in the upper right panel. Overplotting the copy number data in each panel is a rectangle that indicates the start and stop coordinates in the RangedDataCNV object.

Value

An object of class trellis.

Note

Note that users must pass a RangedDataCNV object called 'range'. As mentioned previously, it can be helpful to pass an integer called 'frame' that indicates how much contextual data we should plot surrounding each genomic interval.

If the lattice package is loaded after loading VanillaICE, the generic definition for xyplot in VanillaICE will be masked. To unmask the S4 generic in VanillaICE, do

xyplot <- VanillaICE:::xyplot

Author(s)

R. Scharpf

See Also

xyplot, xypanel

To modify the plot appearance from the default, additional arguments can be passed to panel.xyplot, lpoints, and lrect.

Examples

```
## simulated data
library(oligoClasses)
library(IRanges)
library(VanillaICE)
data(oligoSetExample, package="oligoClasses")
data(hmmResults, package="VanillaICE")
## to visualize each range in it's own panel surrounded by a
## frame of 2,000,000 bases:
## (here the frames are overlapping, but the method could be
## applied more generally to a collection of ranges from
## different chromsomes and samples)
xyplot(cn~x | range, data=oligoSet,
       range=hmmResults,
       frame=2e6, panel=xypanel,
```

```
cex=2,
pch=".",
col.het="salmon",
fill.het="salmon",
col.hom="royalblue",
fill.hom="royalblue",
state.cex=0.5,
border="orange", scales=list(x="free"),
par.strip.text=list(cex=0.5),
xlab="Mb", ylab=expression(log[2]("copy number")))
```

Index