

RSNPper: utilities for SNP data

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1 Introduction

This document describes *RSNPper* version 1.0, added to Bioconductor in October of 2003. This first version focuses on SNP metadata, with functions that retrieve SNP-related data from the Boston Children's Hospital Informatics Program *SNPper* web service ?.

Earlier non-released versions of this package included considerable code for working with *prettybase* format and for conducting other tasks in SNP discovery projects. That material has been moved to `inst/OLD` and may be re-introduced later. Users seeking legacy support should contact the author.

2 How it works

Loading required package: XML

The core of this package is the XML-RPC service at CHIP accessible through the following URL stub:

```
> print(.SNPperBaseURL)
```

```
[1] "http://snpper.chip.org/bio/rpcserv/dummy?cmd="
```

The `useSNPper` function allows you to work directly with the XML-RPC server by packing up appropriate command and argument strings.

```
> dput(useSNPper)
```

```
function (cmd, parmstring)
{
  targ <- url(paste(.SNPperBaseURL, cmd, parmstring, sep = ""))
  open(targ)
  on.exit(close(targ))
  readLines(targ)
}
```

```
> print(useSNPper("geneinfo", "&name=CRP")[1:7])
```

```
[1] " <SNPPER-RPC SOURCE=\"*RPCSERV-NAME*\" VERSION=\"\$$Revision: 1.38 $\" GENOME=\"hg17
[2] " <GENEINFO>"
[3] " <GENE ID=\"1440\">"
[4] " <GENEID>1440</GENEID>"
[5] " <NAME>CRP</NAME>"
[6] " <CHROM>chr1</CHROM>"
[7] " <STRAND>--</STRAND>"
```

The main functions of *RSNPper* attend to simplifying specification of parameters and parsing and packaging the XML results.

Note on auditability. All functions return textual information coupled with auditing information as a 'toolInfo' attribute, detailing the SNPper supplied information on the human genome sequence build, the dbSNP version, and the SNPper version from which the results are obtained. At present, there is one exception: when `itemsInRange` is invoked with `item='countsnps'`, no toolInfo data is obtained. This will be corrected once the `countsnps` command at SNPper returns valid XML element tags.

3 Overview of the functions

The current set of functions intended for investigative use is:

- `geneInfo` – general information about location and nomenclature
- `geneLayout` – information about exon locations

- `geneSNPs` – all SNPs associated with a given gene
- `SNPinfo` – detailed information on a SNP
- `itemsInRange` – supports chromosome scanning for genes, SNPs, or counts of SNPs

An omission: for SNP information, I have not collected information on submitter.

4 Demonstrations

4.1 Obtaining information on genes

The `geneInfo` function will collect some basic information on a gene. The gene may be specified by HUGO name, mRNA accession number, or SNPper id.

```
> print(geneInfo("CRP"))
```

```

                                snpper.ID           NAME
                                "1440"              "CRP"
                                CHROM              STRAND
                                "chr1"             "-"
                                PRODUCT             LOCUSLINK
"C-reactive protein, pentraxin-related"          "1401"
                                OMIM              UNIGENE
                                "123260"          "Hs.76452"
                                SWISSPROT         NSNPS
                                "P02741"         "101"
                                REFSEQACC         MRNAACC
                                " "             "NM_000567"
                                TRANSCRIPT.START   CODINGSEQ.START
                                "156495525"       "156496388"
                                TRANSCRIPT.END     CODINGSEQ.END
                                "156497437"       "156497348"

attr(,"toolInfo")
      SOURCE           VERSION           GENOME           DBSNP
"*RPCSERV-NAME*" "$Revision: 1.38 $" "hg17"          "123"

```

The `geneLayout` function provides information on exon locations.

```
> print(geneLayout("546"))
```

```

      ID           NAME           CHROM TRANSCRIPT.START
      " "          "RLF"          "chr1"  "40296154"
CODINGSEQ.START  TRANSCRIPT.END  CODINGSEQ.END  exon1.start

```

```

"40296165"      "40375684"      "40375212"      "40296154"
  exon1.end      exon2.start      exon2.end      exon3.start
"40296401"      "40323820"      "40323974"      "40325537"
  exon3.end      exon4.start      exon4.end      exon5.start
"40325618"      "40330397"      "40330529"      "40337177"
  exon5.end      exon6.start      exon6.end      exon7.start
"40337379"      "40357339"      "40357475"      "40366282"
  exon7.end      exon8.start      exon8.end
"40366423"      "40370557"      "40375684"
attr(,"toolInfo")
      SOURCE          VERSION          GENOME          DBSNP
"*RPCSERV-NAME*" "$Revision: 1.38 $" "hg17"          "123"

```

Information on all the genes catalogued in a certain chromosomal region can be obtained using `itemsInRange`.

```
> print(itemsInRange("genes", "chr1", "156400000", "156500000"))
```

```

[[1]]
      NAME          CHROM
      "CRP"         "chr1"
      PRODUCT      NSNPS
"C-reactive protein, pentraxin-related" "101"

```

```

$CHR
[1] "chr1"

```

```

$START
[1] "156400000"

```

```

$END
[1] "156500000"

```

```

$COUNT
[1] "1"

```

```

attr(,"toolInfo")
      SOURCE          VERSION          GENOME          DBSNP
"*RPCSERV-NAME*" "$Revision: 1.38 $" "hg17"          "123"

```

4.2 Obtaining information on SNPs

Suppose you want information on the SNP with dbSNP id rs25.

```
> print(SNPinfo("25"))
```

```
      DBSNPID      TSCID CHROMOSOME  POSITION  ALLELES  ROLE  RELPOS
      "rs25"      " "    "chr7"  "11357382"  "A/G"    " "   " "
      AMINO      AMINOPOS
      " "      " "
attr(,"toolInfo")
      SOURCE          VERSION          GENOME          DBSNP
      "*RPCSERV-NAME*" "$Revision: 1.38 $"          "hg17"          "123"
```

Suppose instead you want information on all the SNPs cataloged in a certain chromosomal region.

```
> ird <- itemsInRange("snps", "chr1", "156400000", "156500000")
> print(length(ird))
```

```
[1] 314
```

```
> print(ird[1:3])
```

```
[[1]]
      DBSNPID      TSCID CHROMOSOME  POSITION  ALLELES  ROLE
      "rs2263016"  " "    "chr1"  "156400511"  "A/G"    " "
      RELPOS      AMINO  AMINOPOS
      " "      " "    " "
[[2]]
      DBSNPID      TSCID CHROMOSOME  POSITION  ALLELES  ROLE
      "rs2263017"  " "    "chr1"  "156400521"  "A/C"    " "
      RELPOS      AMINO  AMINOPOS
      " "      " "    " "
[[3]]
      DBSNPID      TSCID CHROMOSOME  POSITION  ALLELES  ROLE
      "rs7531018"  " "    "chr1"  "156400582"  "C/T"    " "
```

Note that the start and end locations are supplied as strings. This is to avoid coercion to textual scientific notation.

Additional detail on the count of SNPs can be obtained more briefly:

```
> print(itemsInRange("countsnps", "chr1", "156400000", "156500000"))
```

```
total exonic nonsyn
  310      7      0
```

To see all the SNPs associated with a given gene, use the `geneSNPs` function. This requires knowledge of the SNPper gene id, which can be obtained using `geneInfo`.

```
> gs <- geneSNPs("546")
> print(length(gs))
```

```
[1] 164
```

```
> print(gs[1:3])
```

```
[[1]]
```

DBSNPID	TSCID
"rs6679879"	" "
CHROMOSOME	POSITION
"chr1"	"40286528"
ALLELES	ROLE
"C/G"	"Promoter"
RELPOS	AMINO
"-9637"	" "
AMINOPOS	HUGO
" "	"RLF"
LOCUSLINK	NAME
"6018"	"rearranged L-myc fusion sequence"
MRNA	
"NM_012421"	

```
[[2]]
```

DBSNPID	TSCID
"rs7550355"	" "
CHROMOSOME	POSITION
"chr1"	"40287075"
ALLELES	ROLE
"C/T"	"Promoter"
RELPOS	AMINO
"-9090"	" "
AMINOPOS	HUGO
" "	"RLF"
LOCUSLINK	NAME
"6018"	"rearranged L-myc fusion sequence"
MRNA	

```

"NM_012421"

[[3]]
      DBSNPID          TSCID
"rs12096261"         " "
      CHROMOSOME      POSITION
      "chr1"          "40288041"
      ALLELES         ROLE
      "G/T"           "Promoter"
      RELPOS          AMINO
      "-8124"         " "
      AMINOPOS        HUGO
      " "             "RLF"
      LOCUSLINK       NAME
      "6018" "rearranged L-myc fusion sequence"
      MRNA
      "NM_012421"

```

5 Application: SNP density on chr 1

Human chromosome 1 is approximately 300Mb, and 142,629 SNPs have been recorded as of dbSNP build 106, according to NCBI SNP/maplists/maplist-newmap.html on 13 Sep 03. Let's see if these facilities can recover this sort of information. Counting the number of SNPs on a long chromosomal region seems to take a long time for SNPper, so we will break up the task.

```

> print(itemsInRange("countsnp", "chr1", "1", "100000"))

total exonic nonsyn
  80      1      0

> system("sleep 2")
> print(itemsInRange("countsnp", "chr1", "100001", "200000"))

total exonic nonsyn
   2      0      0

> system("sleep 2")
> print(itemsInRange("countsnp", "chr1", "200001", "300000"))

total exonic nonsyn
   4      0      0

```

```
> system("sleep 2")
```

These runs complete in a reasonable amount of time. Here we will just look at the first 2Mb in intervals of .1Mb.

```
> starts <- as.character(as.integer(seq(1, 2000001, 1e+05)))
> ends <- as.character(as.integer(as.integer(starts) + 99999))
> out <- matrix(NA, nr = 20, nc = 3)
> for (i in 1:20) {
+   cat(i)
+   out[i, ] <- itemsInRange("countsnps", "chr1", starts[i],
+     ends[i])
+   system("sleep 2")
+ }
```

```
1234567891011121314151617181920
```

```
> print(out)
```

	[,1]	[,2]	[,3]
[1,]	80	1	0
[2,]	2	0	0
[3,]	4	0	0
[4,]	0	0	0
[5,]	4	0	0
[6,]	28	0	0
[7,]	126	0	0
[8,]	377	23	2
[9,]	405	7	1
[10,]	356	36	12
[11,]	370	33	5
[12,]	361	31	9
[13,]	334	51	15
[14,]	190	23	9
[15,]	251	30	7
[16,]	392	33	7
[17,]	154	12	5
[18,]	190	14	2
[19,]	209	10	1
[20,]	292	2	1