

# Outreach

Stephanie Hicks



# Orchestrating single-cell analysis with Bioconductor

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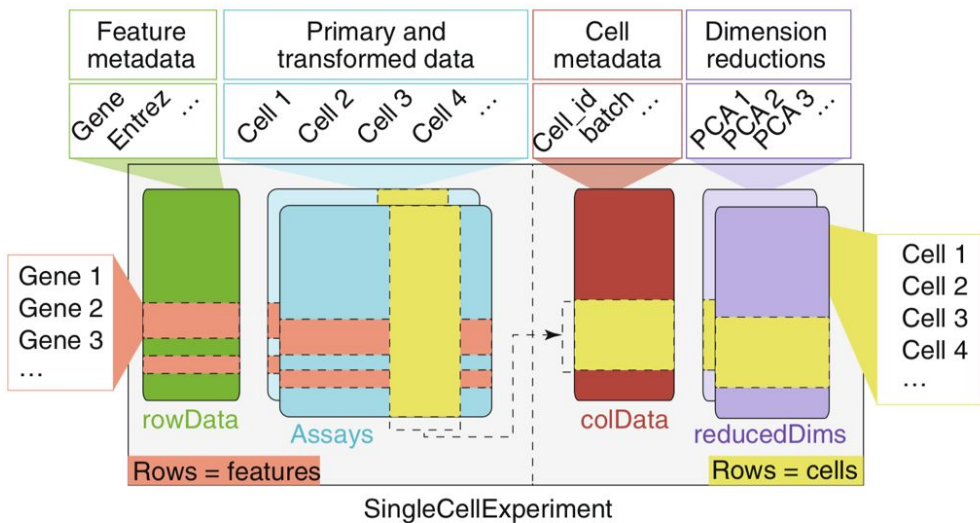
Recent technological advancements have enabled the profiling of a large number of genome-wide features in individual cells. However, single-cell data present unique challenges that require the development of specialized methods and software infrastructure to successfully derive biological insights. The Bioconductor project has rapidly grown to meet these demands, hosting community-developed open-source software distributed as R packages. Featuring state-of-the-art computational methods, standardized data infrastructure and interactive data visualization tools, we present an overview and online book (<https://osca.bioconductor.org>) of single-cell methods for prospective users.

Since 2001, the Bioconductor project<sup>1</sup> has attracted a rich community of developers and users from diverse scientific fields, driving the development of open-source software packages

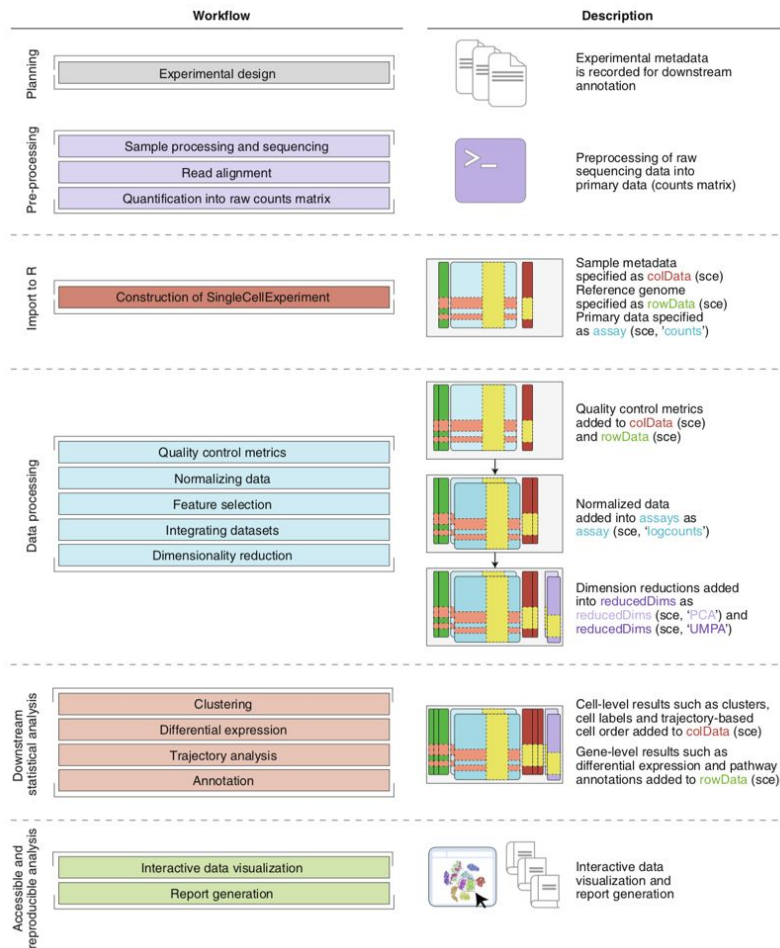
single-cell RNA-seq (scRNA-seq) data, much of the concepts mentioned are also generalizable to other types of single-cell assays. We cover data import, common data containers for storing single-cell

<https://osca.bioconductor.org>

# Standardized ways to store single-cell data



# Bioconductor workflows





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# Orchestrating Single-Cell Analysis with Bioconductor

2019-11-15

## Welcome

This is the website for “**Orchestrating Single-Cell Analysis with Bioconductor**”, a book that teaches users some common workflows for the analysis of single-cell RNA-seq data (scRNA-seq). This book will teach you how to make use of cutting-edge Bioconductor tools to process, analyze, visualize, and explore scRNA-seq data. Additionally, it serves as an online companion for the manuscript “**Orchestrating Single-Cell Analysis with Bioconductor**”.

While we focus here on scRNA-seq data, a newer technology that profiles transcriptomes at the single-cell level, many of the tools, conventions, and



<https://osca.bioconductor.org>

## 10 Clustering

### 10.1 Motivation

### 10.2 What is the “true clustering”?

### 10.3 Graph-based clustering

#### 10.3.1 Background

#### 10.3.2 Implementation

#### 10.3.3 Other parameters

#### 10.3.4 Assessing cluster separation

### 10.4 $k$ -means clustering

### 10.5 Hierarchical clustering

### 10.6 Subclustering

### Session Info

## 11 Marker gene detection

### 11.1 Motivation

### 11.2 Using pairwise $t$ -tests

### 11.3 Alternative testing regimes



## 10.2 What is the “true clustering”?

At this point, it is worth stressing the distinction between clusters and cell types. The former is an empirical construct while the latter is a biological truth (albeit a vaguely defined one). For this reason, questions like “what is the true number of clusters?” are usually meaningless. We can define as many clusters as we like, with whatever algorithm we like - each clustering will represent a different partitioning of the high-dimensional expression space, and is as “real” as any other clustering.

A more relevant question is “how well do the clusters approximate the cell types?” Unfortunately, this is difficult to answer given the context-dependent interpretation of biological truth. Some analysts will be satisfied with resolution of the major cell types; other analysts may want resolution of subtypes; and others still may require resolution of different states (e.g., metabolic activity, stress) within those subtypes. Moreover, two clusterings can be highly inconsistent yet both valid, simply partitioning the cells based on different aspects of biology. Indeed, asking for an unqualified “best” clustering is akin to asking for the best magnification on a microscope without any context.

It is helpful to realize that clustering, like a microscope, is simply a tool to explore the data. We can zoom in and out by changing the resolution of the clustering parameters, and we can experiment with different clustering algorithms to obtain alternative perspectives of the data. This iterative approach is entirely permissible for data exploration, which constitutes the majority of all scRNA-seq data analyses.

Lots of philosophical discussions about single-cell analyses



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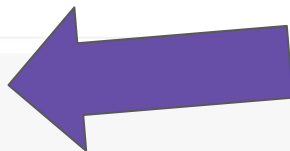
### 11.3 Alternative testing regimes

### 11.4 Handling blocking factors

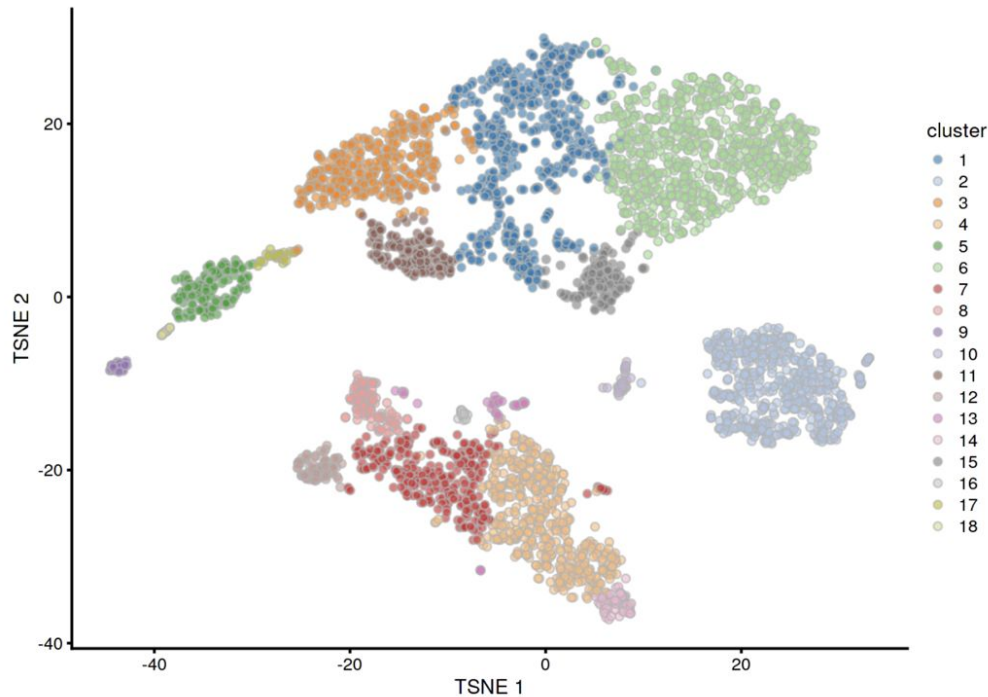
### 11.5 Invalidity of $p$ -values



```
library(scater)
sce.pbmc$cluster <- factor(clust)
plotReducedDim(sce.pbmc, "TSNE", colour_by="cluster")
```



Lots of R/Bioc  
code to analyze  
single-cell data





simpleSingleCell to trigger propagation of

Thursday, July 2nd ▾

Friday, July 3rd ▾

**Aaron Lun** 🇦🇺 12:30 AM

Excellent.

**Hervé Pagès** 7:15 PM

Same problem with pandoc 2.7.3. Updating to the latest pandoc (2.10) doesn't help either (just tried this on my laptop, still running Ubuntu 16.04 here). The HTML source code I get locally is the same as the online HTML:

```
<p>To inspect the object, we can simply type <code>sce</code> into the console to see some pertinent information, which will display an overview of the various slots available to us (which may or may not have any data).</p>
```

```
<div class="sourceCode" id="cb10"><pre class="sourceCode r"><code class="sourceCode r"><span id="cb10-1"><a href="data-infrastructure.html#cb10-1" aria-hidden="true"></a>sce</span></code></pre></div>
```

```
<pre><code>## class: SingleCellExperiment
```

```
## dim: 10 3
```

```
## metadata(0):
```

```
## assays(1): counts
```

```
## rownames(10): gene_1 gene_2 gene_3 gene_4 gene_5 gene_6 gene_7 gene_8 gene_9 gene_10
```



# Integrative analysis of sc data



#BIRSBioIntegration



**Banff International Research Station**  
for Mathematical Innovation and Discovery



**Aedín Culhane**  
Dana-Farber  
Cancer Institute/  
Harvard Chan



**Elana Fertig**  
Johns Hopkins  
University



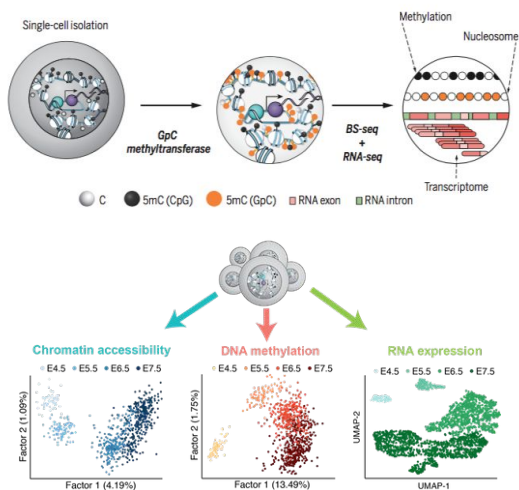
**Kim-Anh Lê Cao**  
The University of  
Melbourne

<https://www.birs.ca/events/2020/5-day-workshops/20w5197>

# 3 Hackathon Challenges

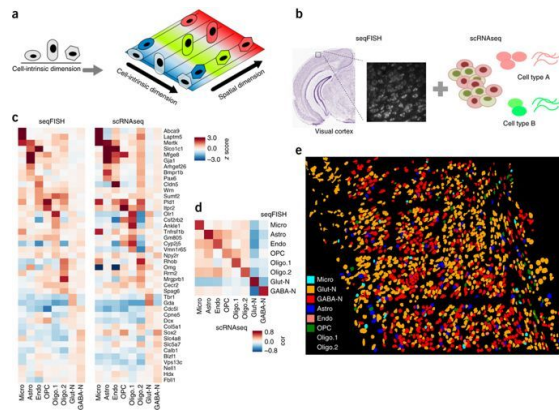
## Gastrulation (scNMT)

826 cells matching across all data sets (transcriptome, DNA accessibility and DNA methylation) after quality control and filtering.



## Adult mouse visual cortex seqFISH, scRNAseq

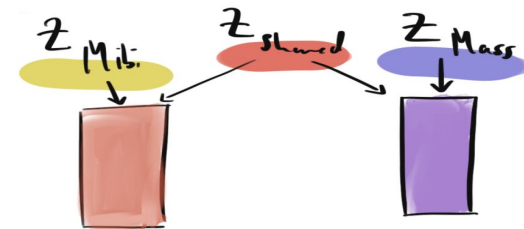
- seqFISH - 1,597 single cells x 125 genes mapped (Zhu *et al* 2018)
- scRNA-seq. ~1,600 cells (Tasic *et al* 2016)



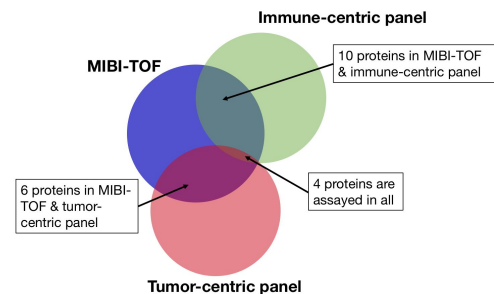
## Breast Cancer sc Proteomics

Non-overlapping patients

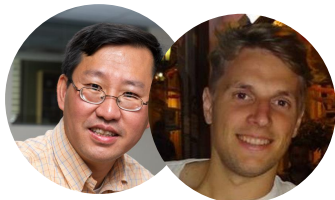
MIBI 40 TN, Mass Tag 7 TN



... with 20 overlapping proteins



# 6 Keynotes, 16 Contributed talks, 9 Brainstorming sessions



seqfish\_theme

**Guo-Cheng Yuan & Ruben Dries**

Dana-Farber Cancer Institute,  
Harvard TH Chan School of Public  
Health & Boston University



sc\_targ\_proteomics\_theme

**Aedin Culhane & Olga Vitek**

Dana-Farber Cancer Institute,  
Harvard TH Chan School of Public  
Health & Northeastern University



scNMT-seq\_theme

**Ricard Arguelaget & Oliver Stegle**

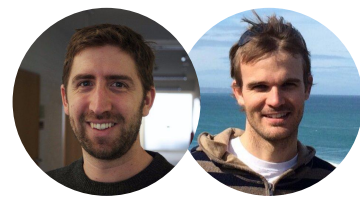
German Cancer Research Center &  
EMBL



summary\_analyses\_theme

**Kim-Anh Lê Cao & Casey Green**

University of Melbourne & Uni  
Pennsylvania



benchmark\_theme

**Mike Love & Matt Ritchie**

University of North Carolina-Chapel Hill  
& Walter and Eliza Hall Institute



**Susan Holmes**  
Stanford University

interpretation\_theme



**Vincent Carey**  
Harvard Medical School and  
Brigham & Women's Hospital

software\_theme



**Elana Fertig**  
Johns Hopkins University

future\_theme

# #Bioc2020

<http://bioc2020.bioconductor.org/>



## BioC 2020: Where Software and Biology Connect

When: July 29 - 31, 2020

What: Community/Developer Day, Main Conference

Where: [venue](#), Boston, USA

Slack: [Bioconductor Team](#) (#bioc2020 channel)

Twitter: [#bioc2020](#)

Now Virtual

