

# Metabolomics

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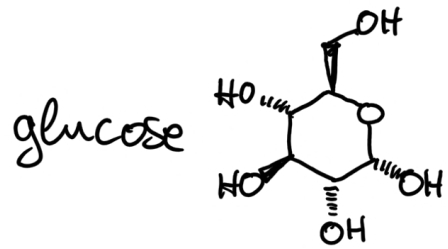
CSAMA 2019

# Content

- (Brief) introduction to metabolomics
- Preprocessing of LC-MS data
- Normalization
- Annotation/identification

# Metabolite? Metabolism?

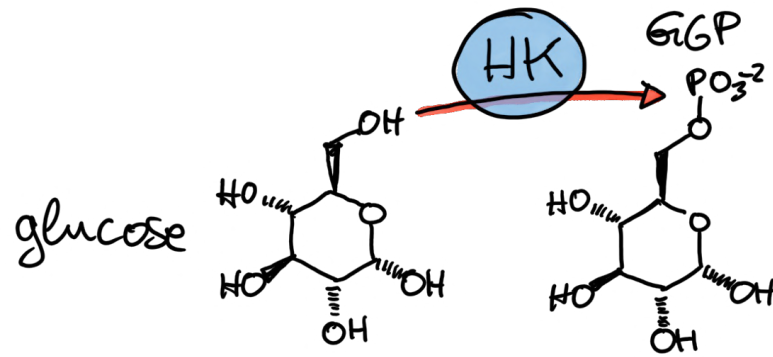
- Glycolysis



- Key metabolic pathway common to all cells.
- Creates energy by converting glucose to pyruvate.

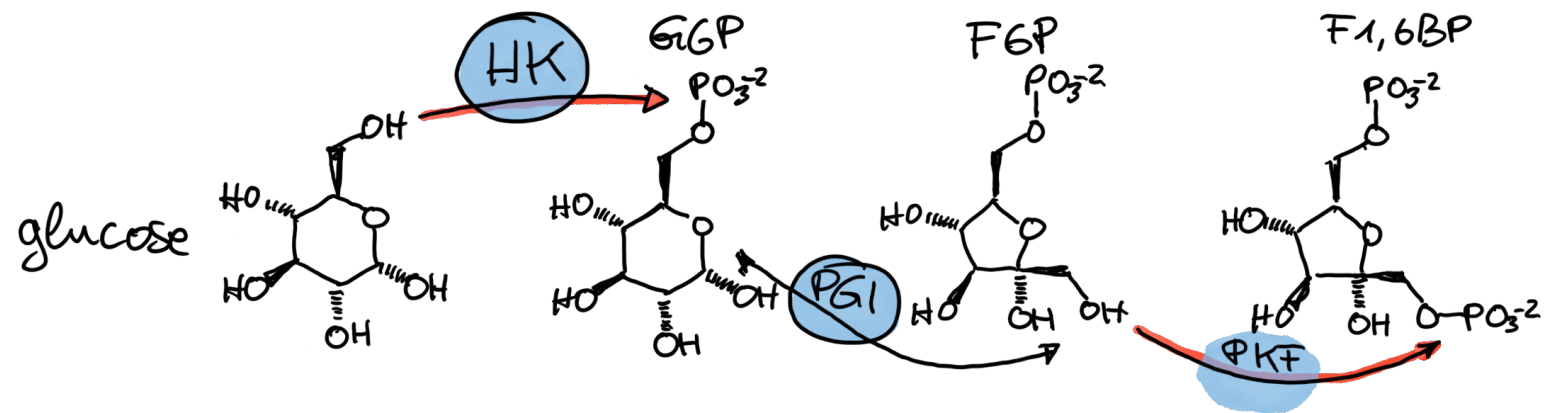
# Metabolite? Metabolism?

- Glycolysis



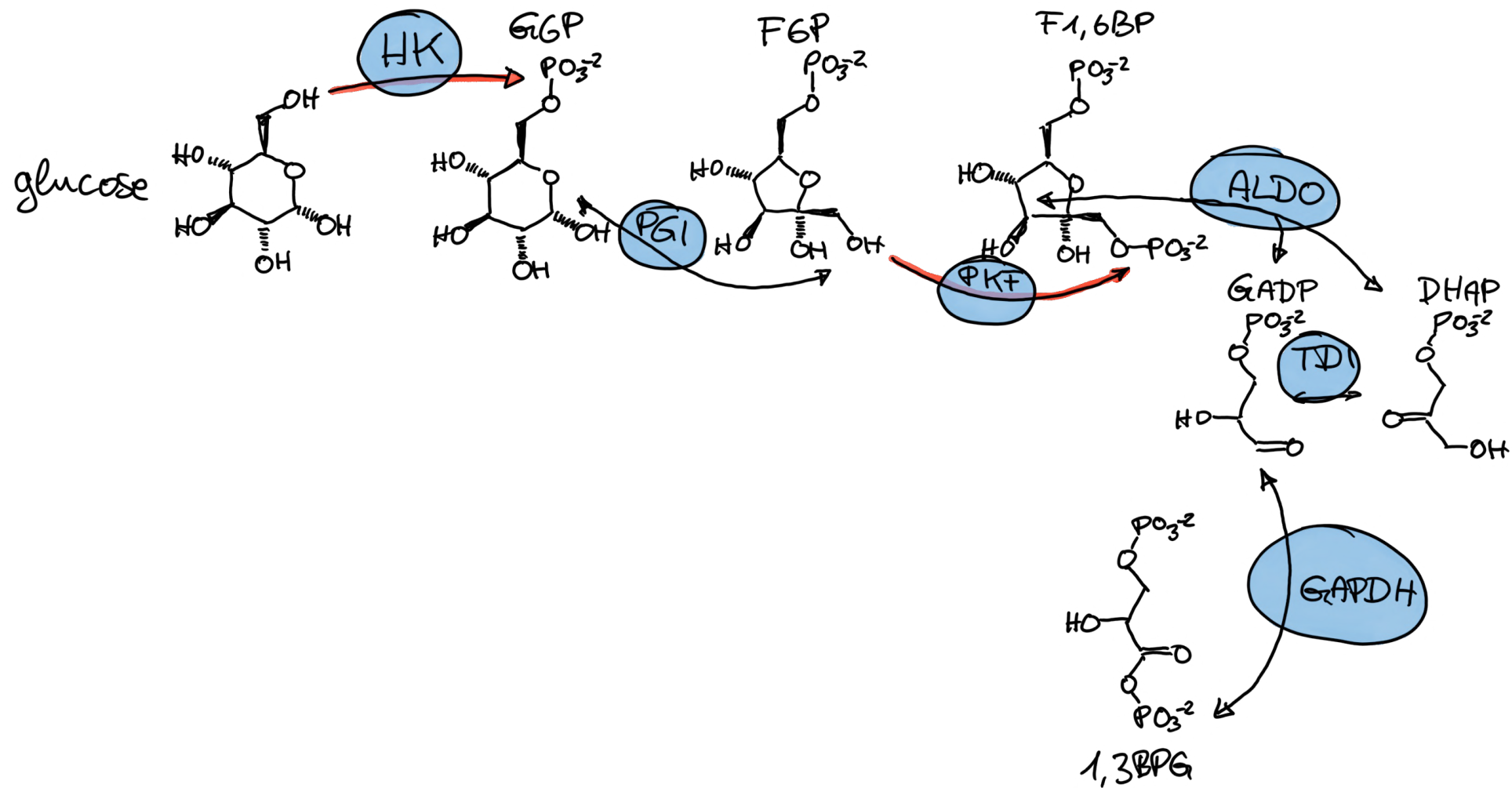
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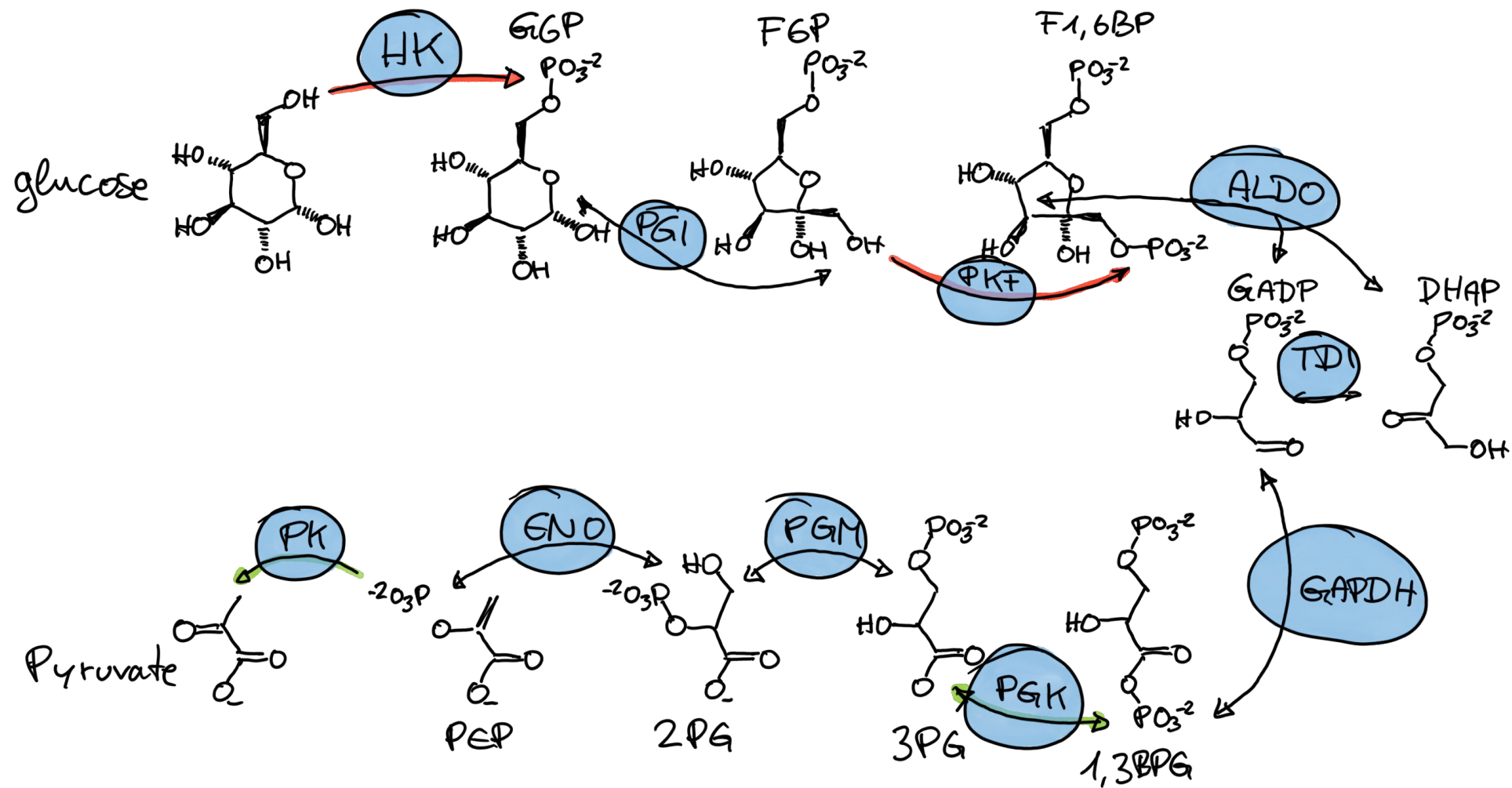
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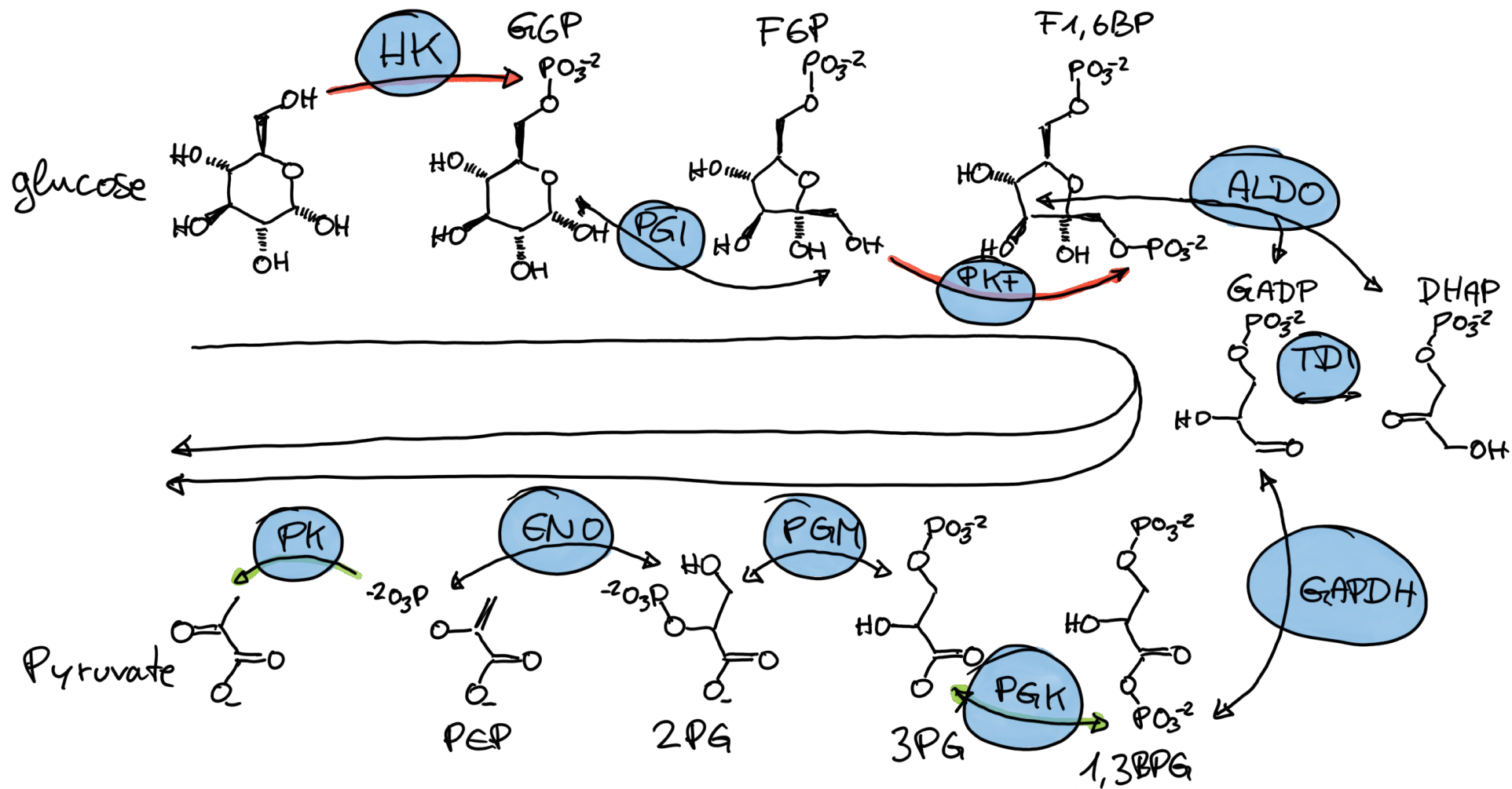
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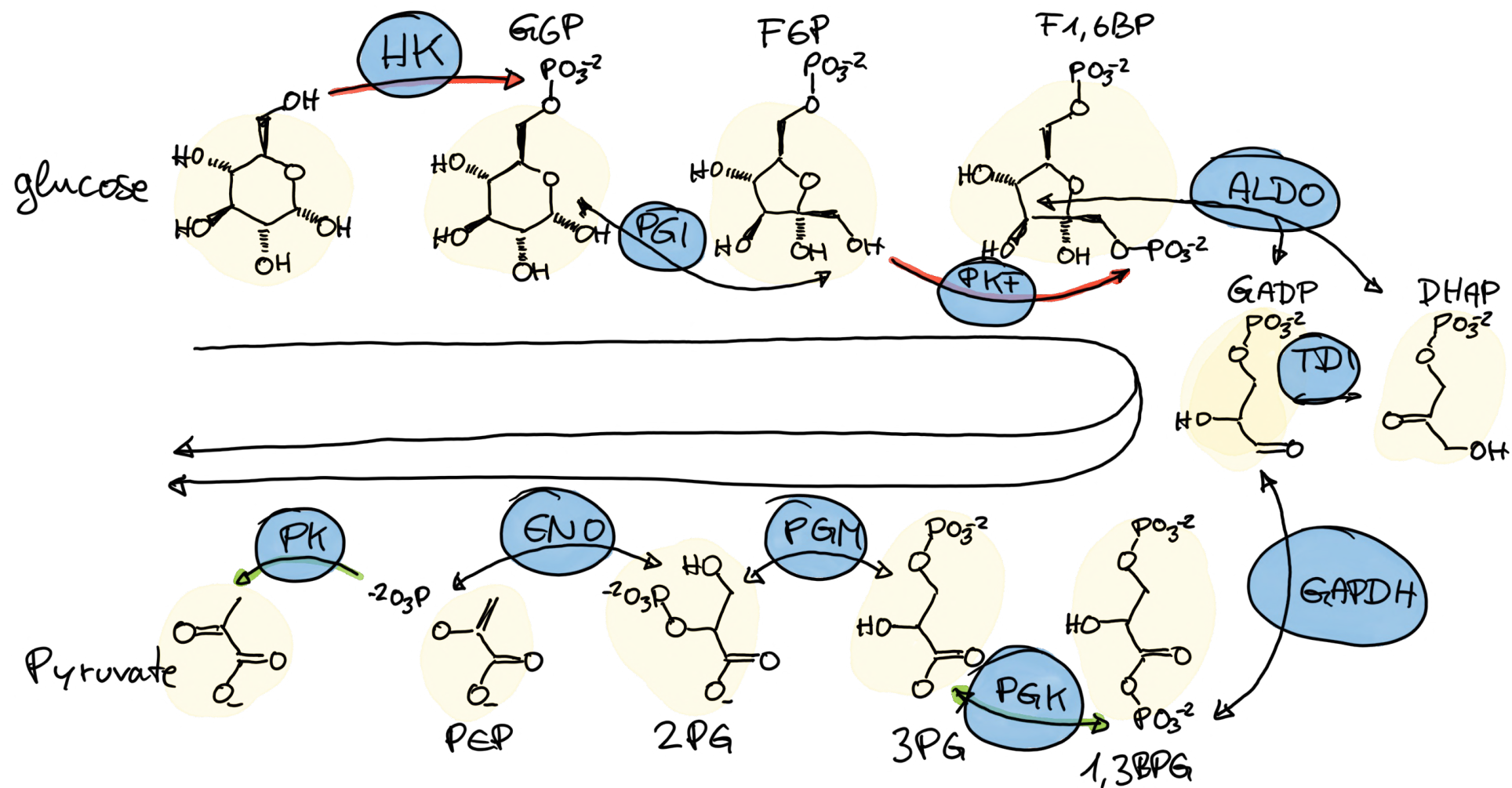
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# Metabolite? Metabolism?

- Glycolysis



- Metabolites: intermediates and products of cellular processes.

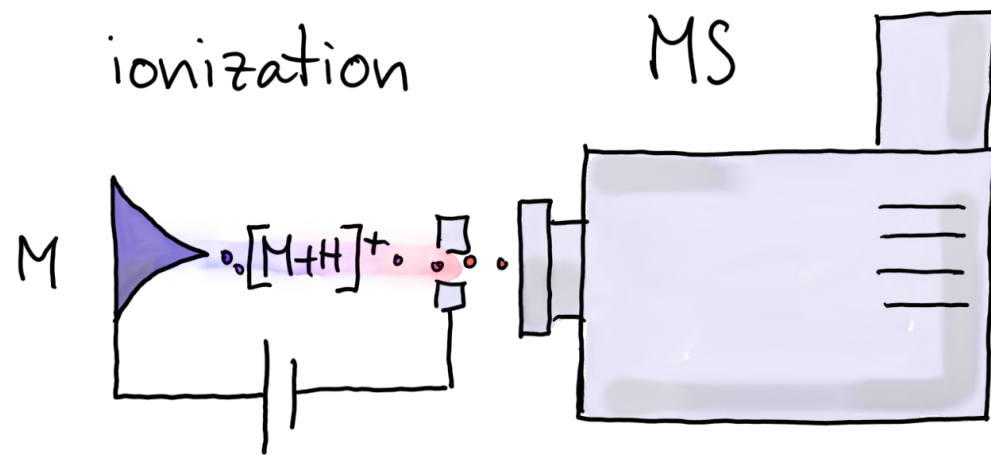
# Metabolomics?

- Large-scale study of small molecules (metabolites) in a system (cell, tissue, organism).
- Comparison of the different -omes:
- **Genome**: what can happen.
- **Transcriptome**: what appears to be happening.
- **Proteome**: what makes it happen.
- **Metabolome**: what actually happened.
- Metabolome influenced by genetic **and** environmental factors.

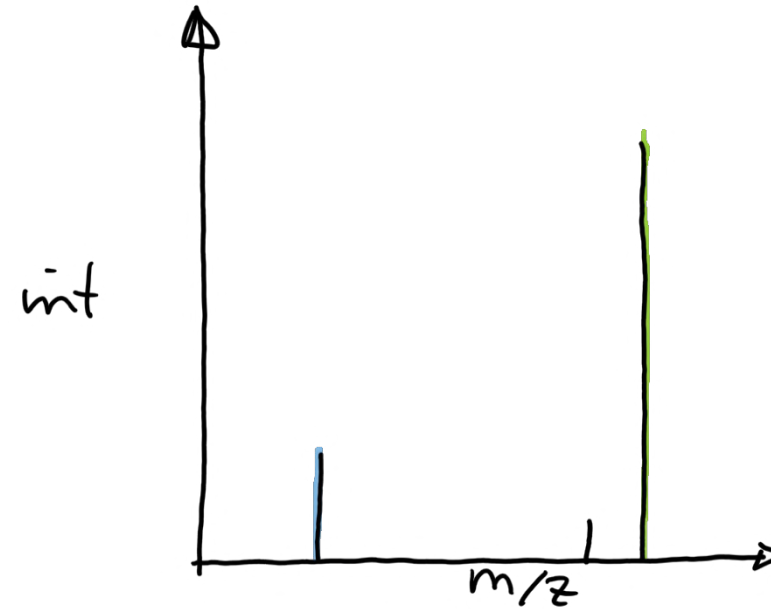
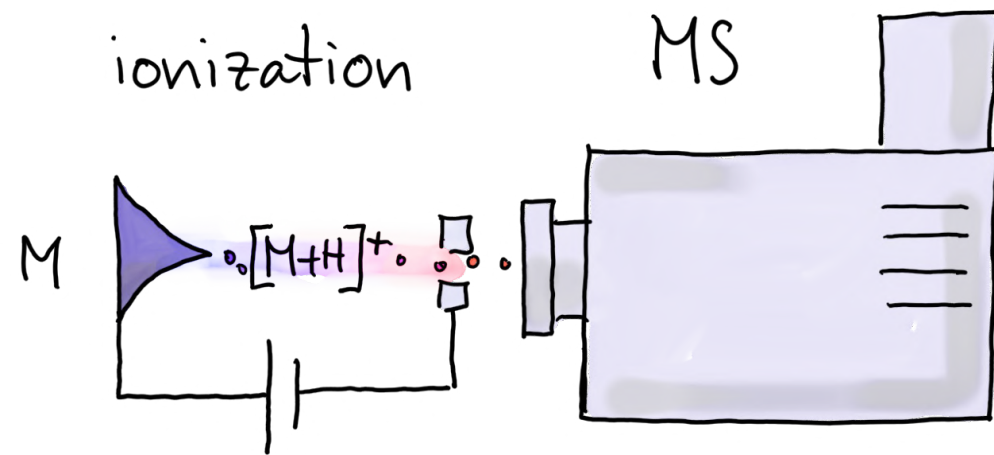
# How can we measure metabolites?

- Nuclear Magnetic Resonance (NMR) - not covered here.
- Mass spectrometry (MS)-based metabolomics.
- Metabolites small enough to be directly measured by MS.
- Most metabolites uncharged - need to create ions first.

# Mass Spectrometry (MS)



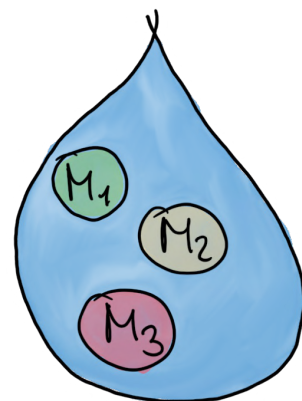
# Mass Spectrometry (MS)



- **Problem:** unable to distinguish between metabolites with the same/similar mass-to-charge ratio ( $m/z$ ).
- **Solution:** additional separation of metabolites prior to MS.

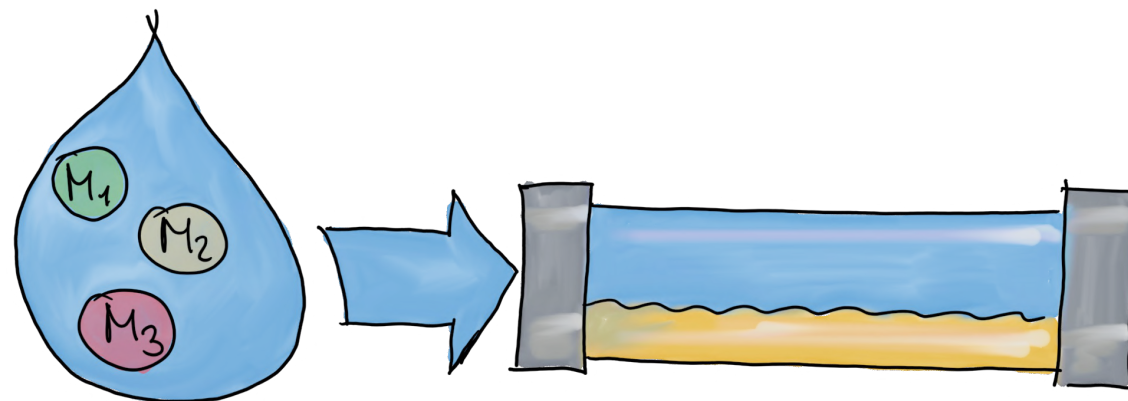
# Liquid chromatography

- Sample is dissolved in a fluid (mobile phase).



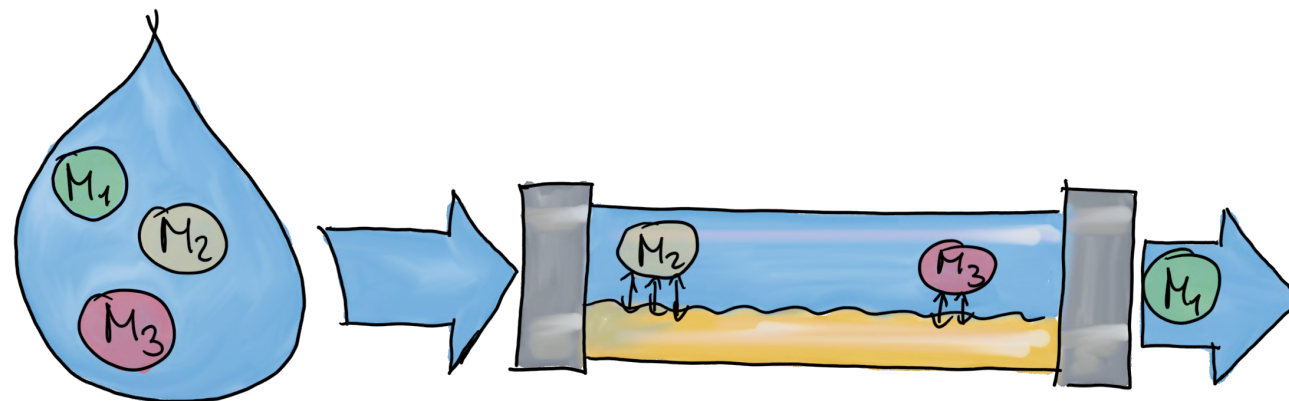
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- Mobile phase carries analytes through column (stationary phase).



# Liquid chromatography

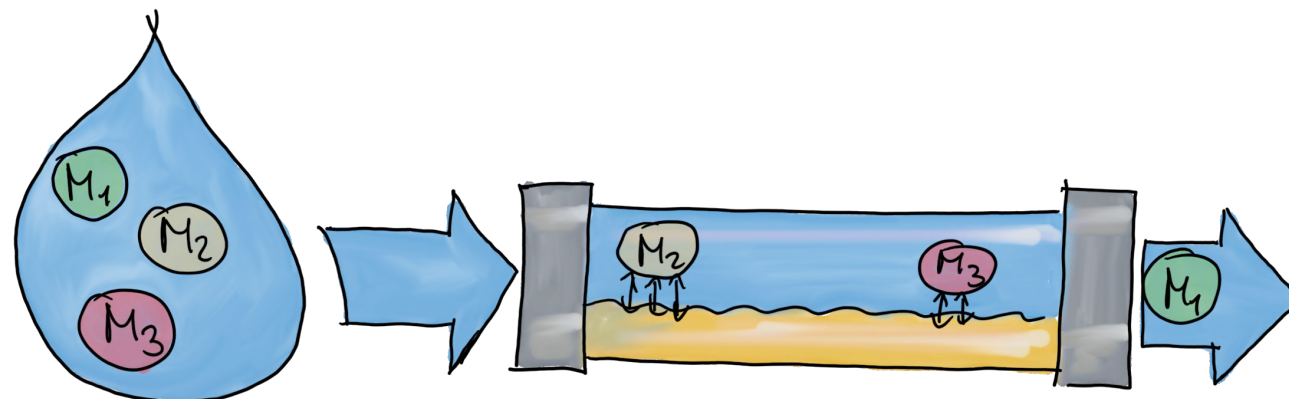
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- Separation based on affinity for the column's stationary phase.



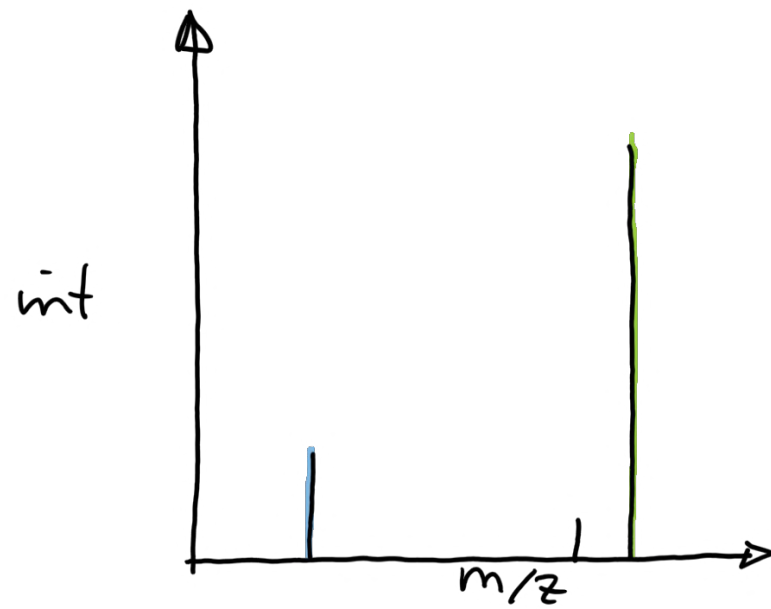
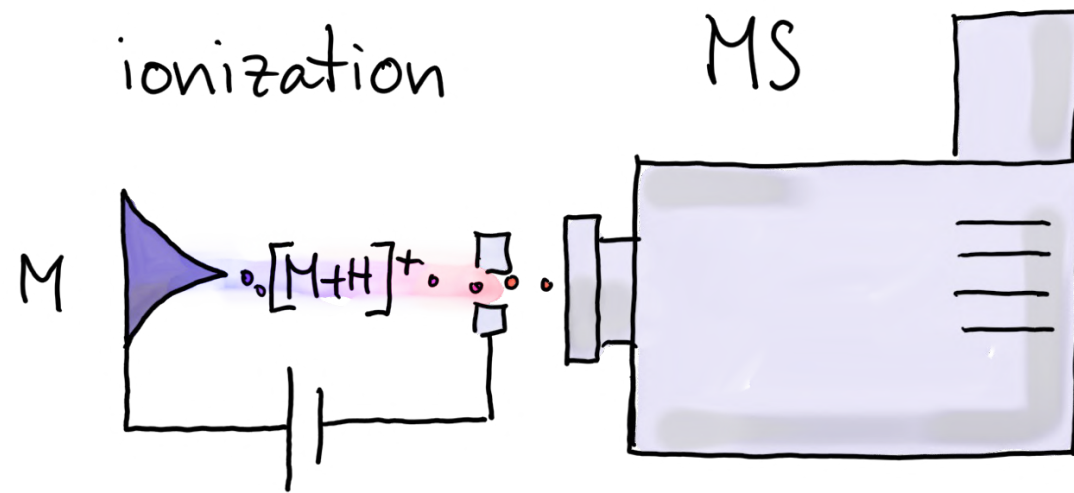


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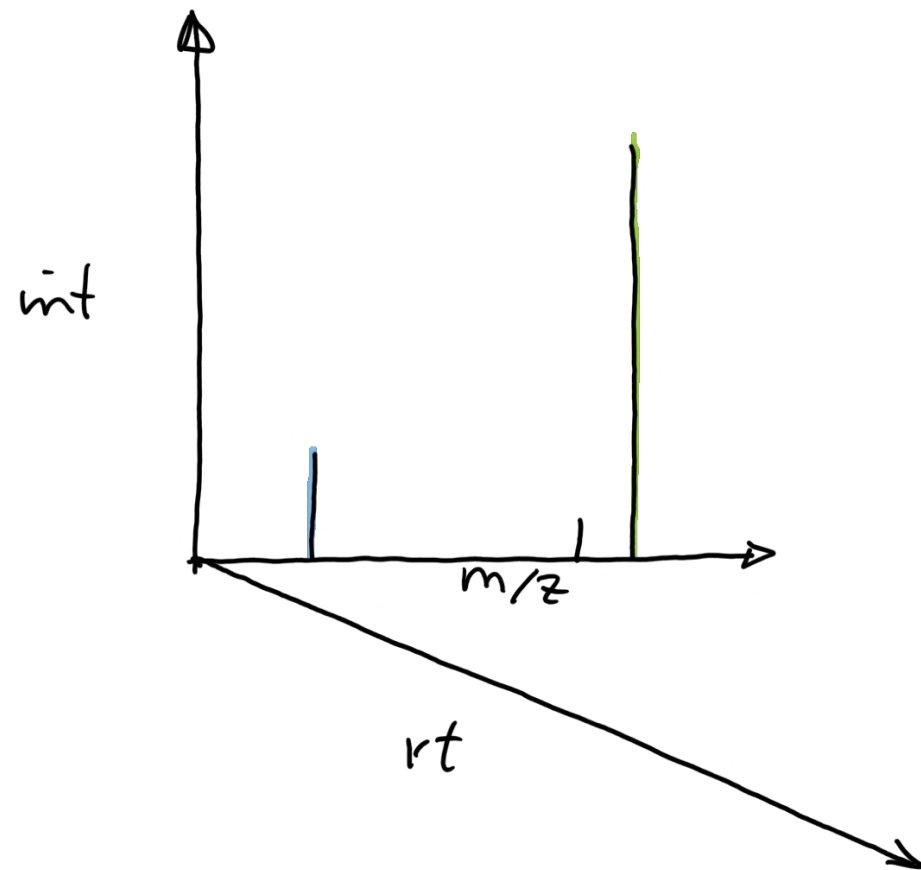
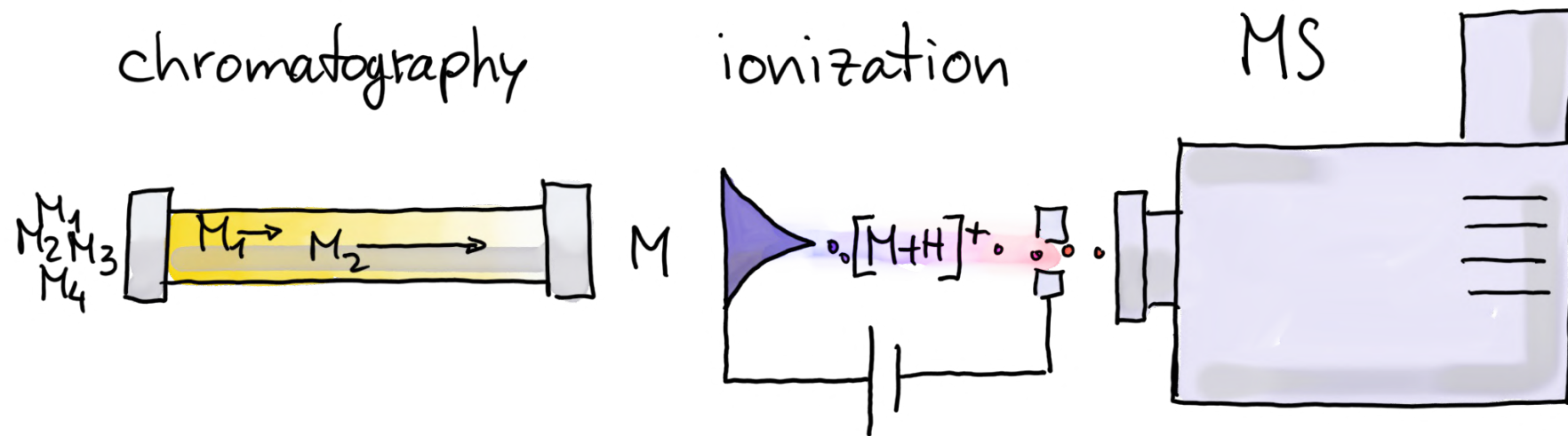
- Sample is dissolved in a fluid (mobile phase).
- Mobile phase carries analytes through column (stationary phase).
- Separation based on affinity for the column's stationary phase.
- [HILIC](#) (hydrophilic liquid interaction chromatography):
  - Hydrophilic, polar stationary phase.
  - Analytes solved in mobile phase.
  - Analytes separated by polarity: compounds with low polarity elute first, with high polarity later.



# Liquid Chromatography Mass Spectrometry (LC-MS)



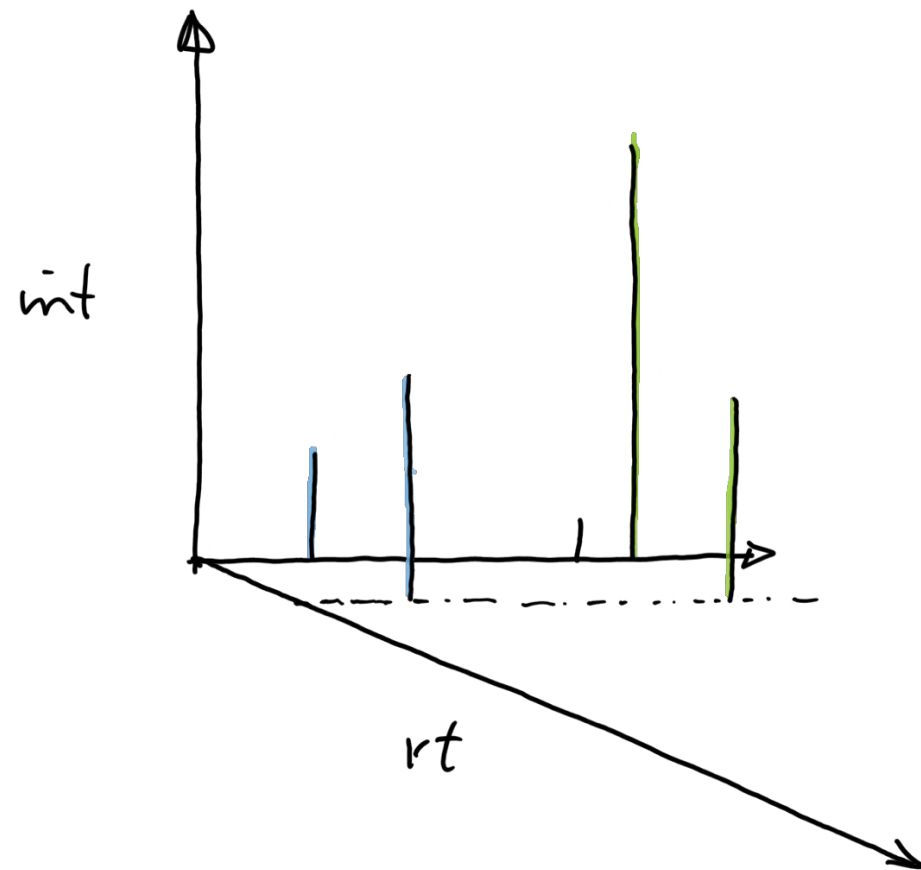
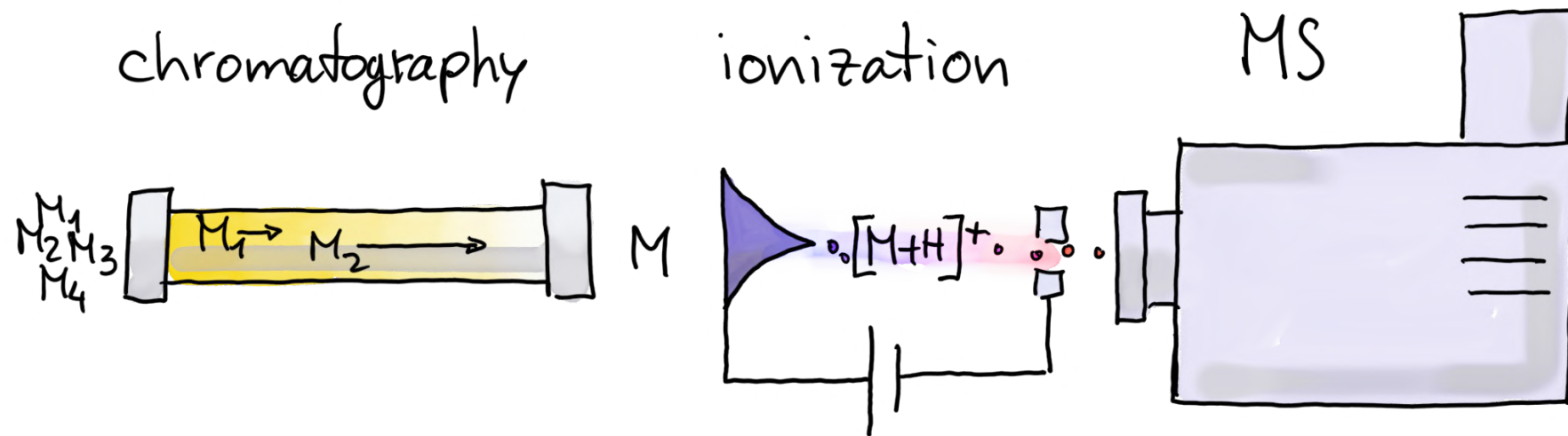
# Liquid Chromatography Mass Spectrometry (LC-MS)



We gain an additional dimension:

- retention time.

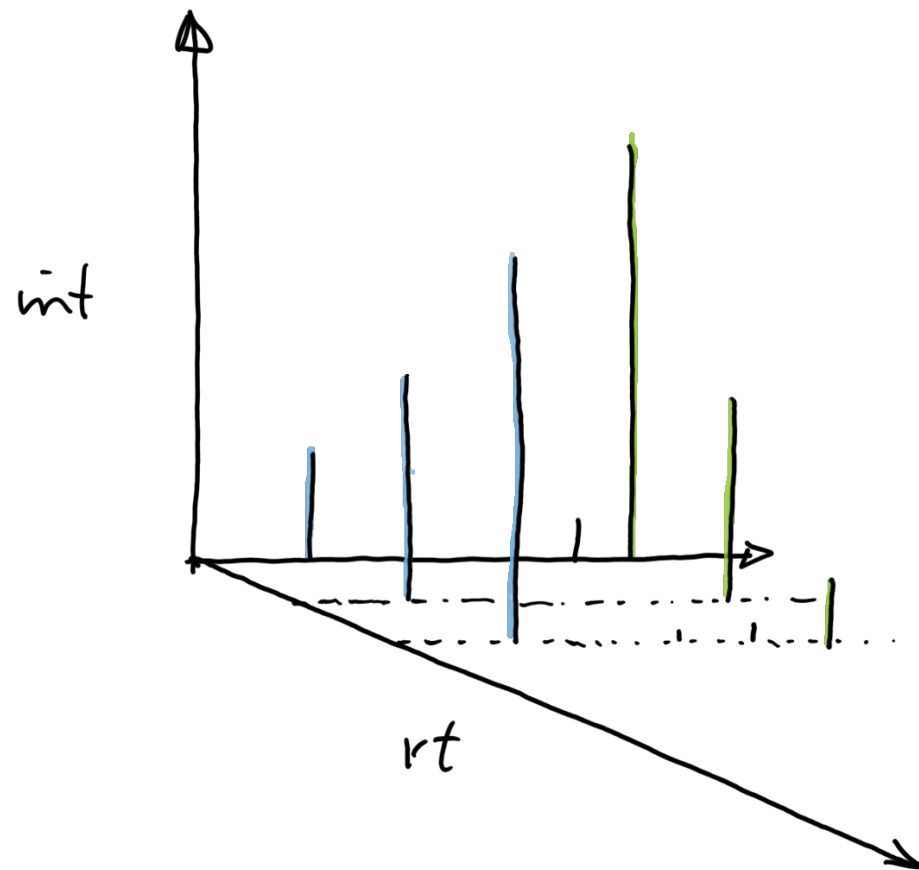
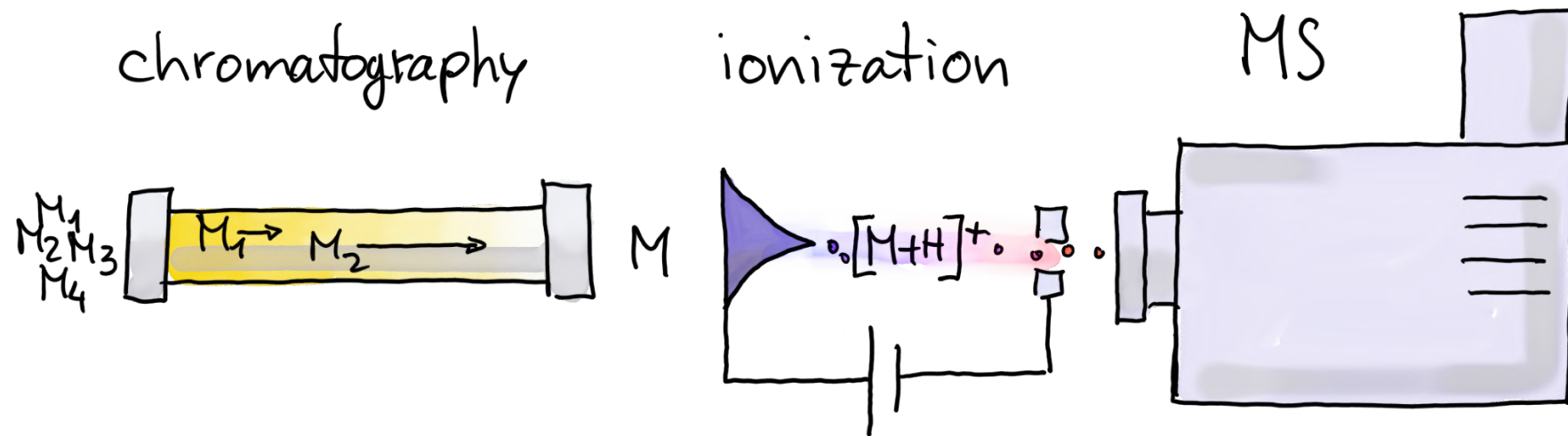
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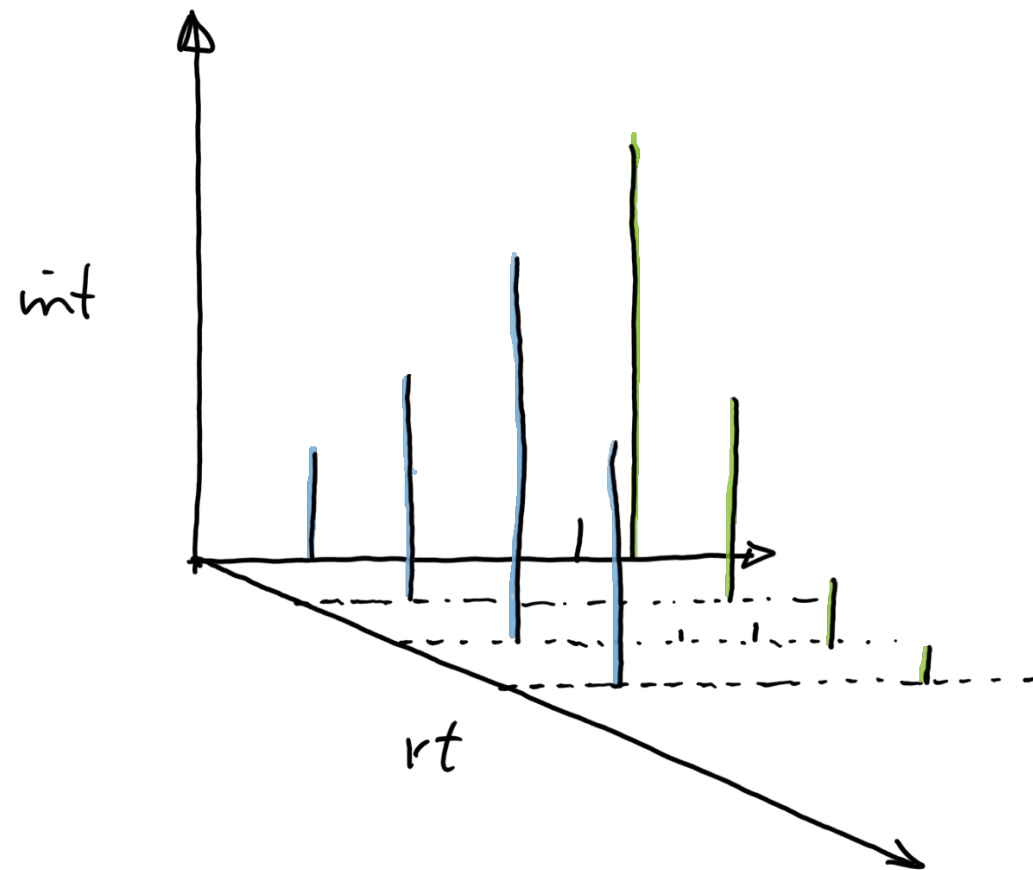
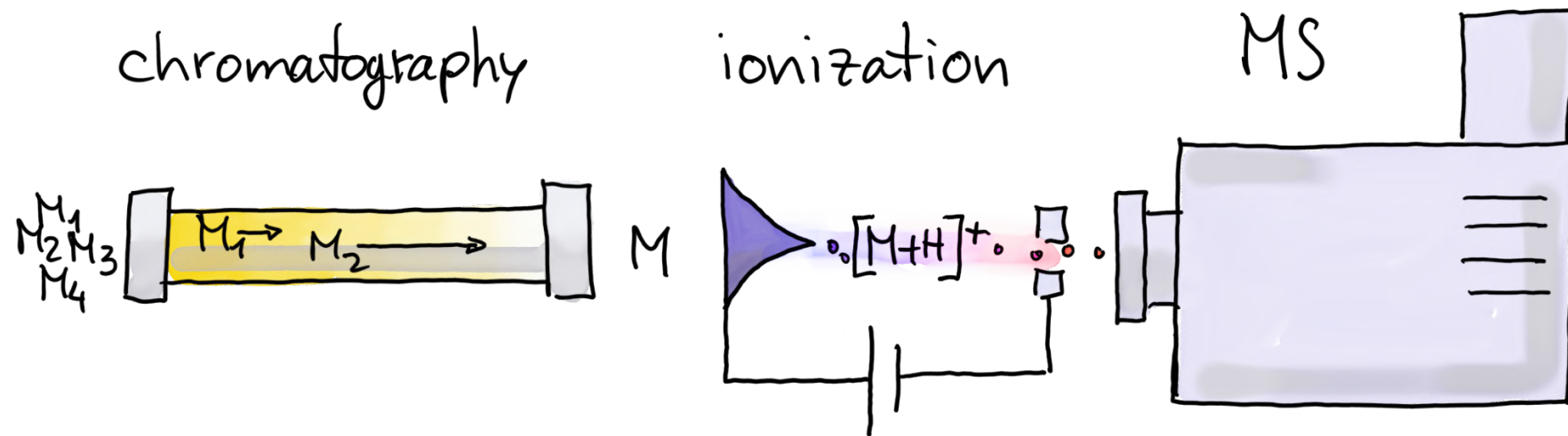
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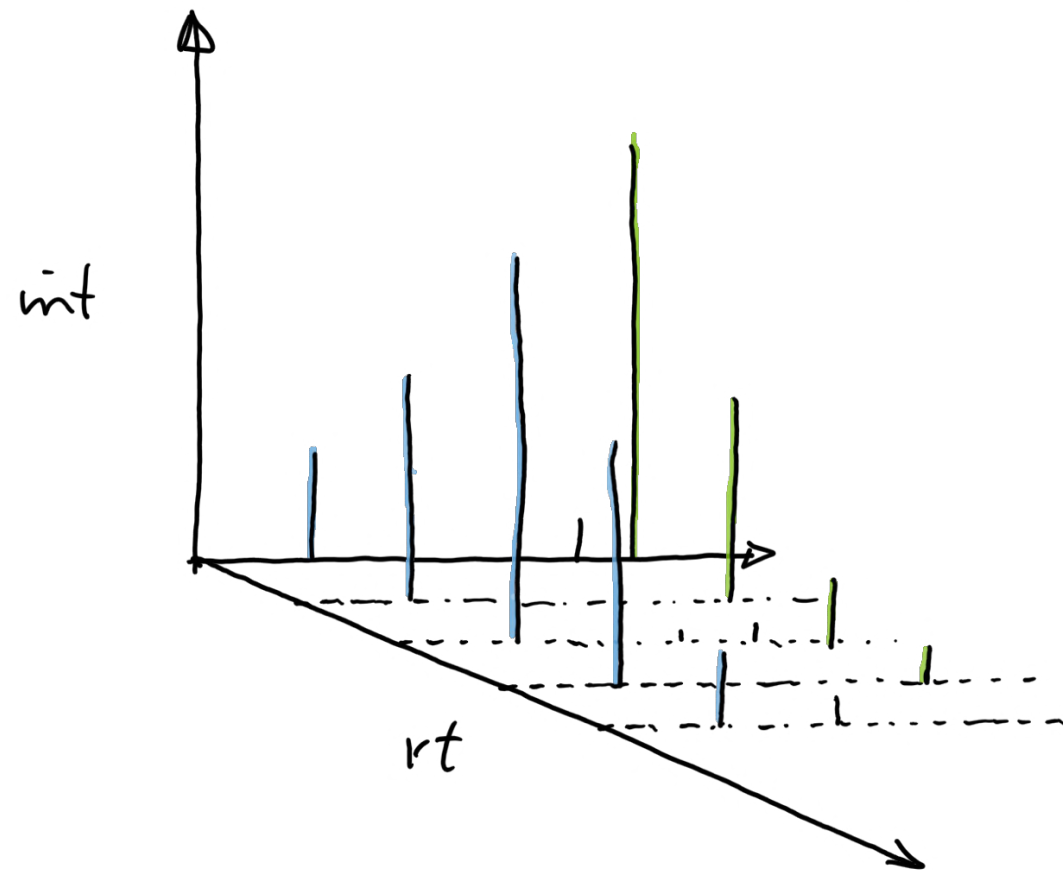
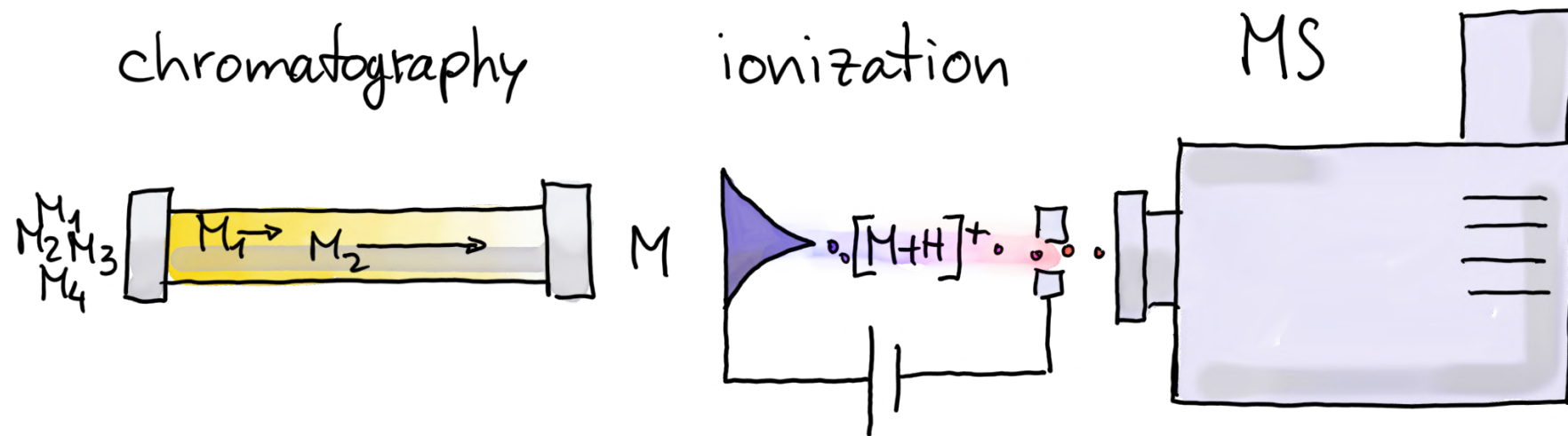
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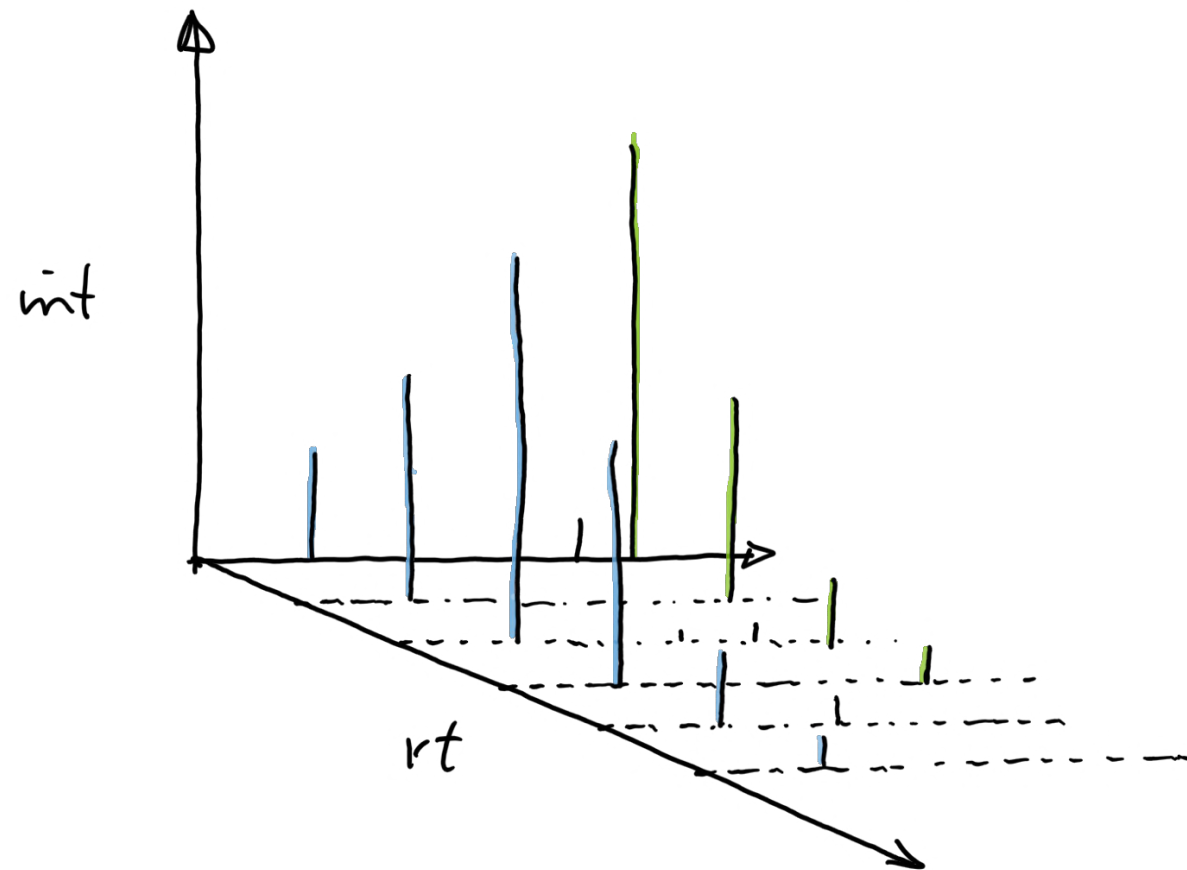
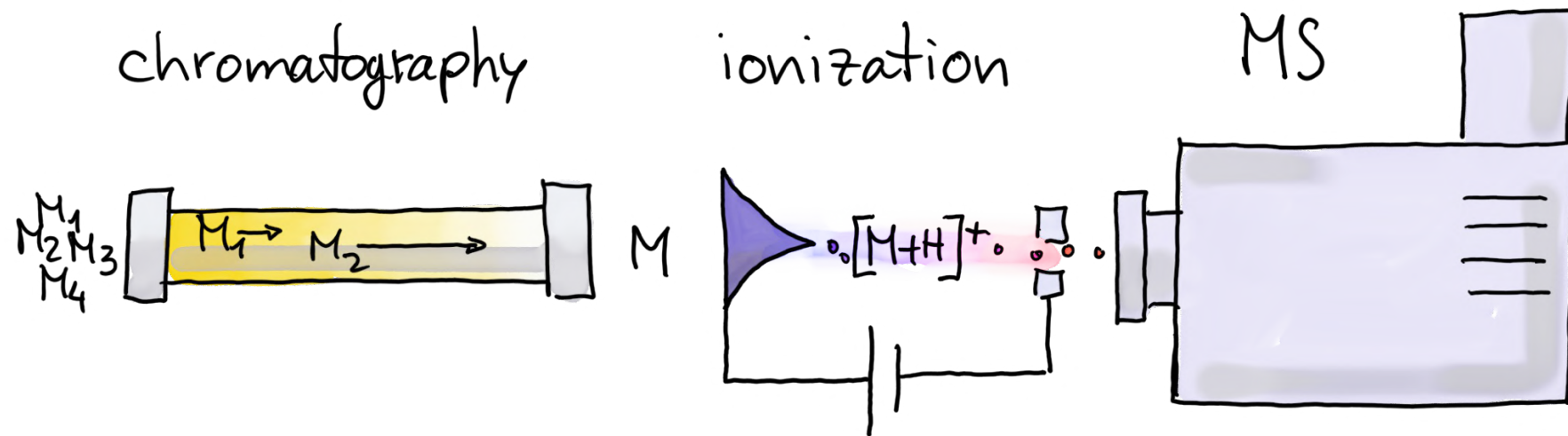
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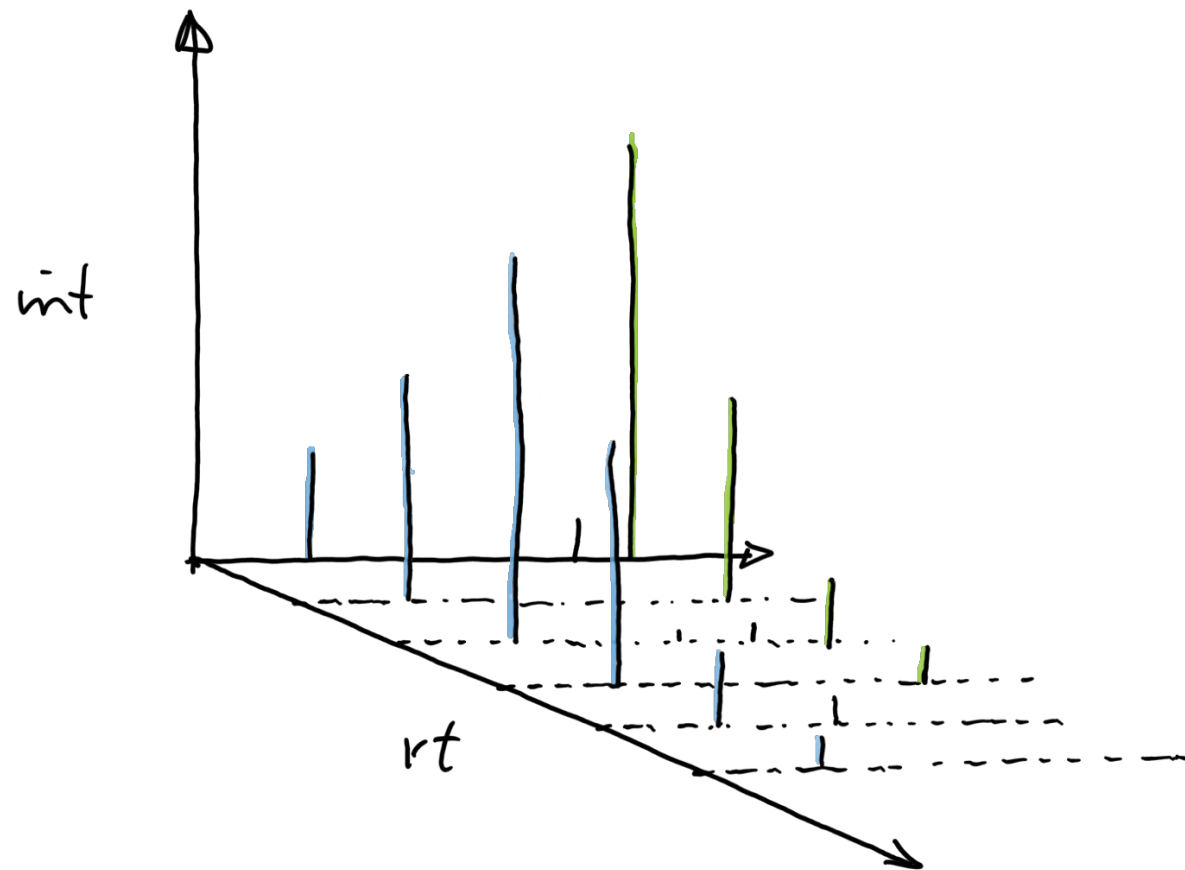
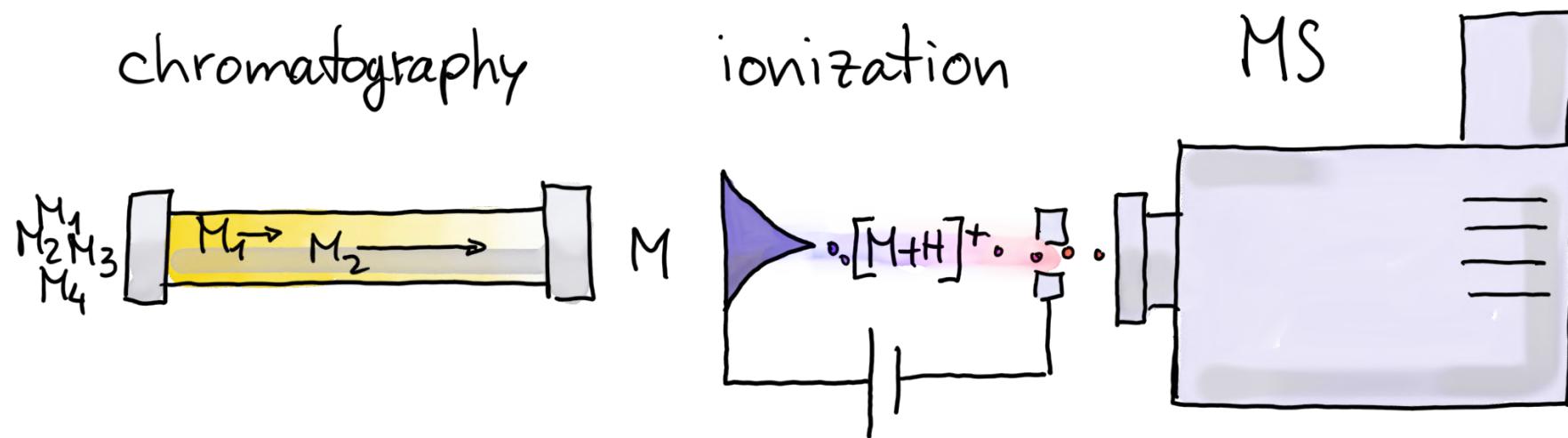


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# Liquid Chromatography Mass Spectrometry (LC-MS)



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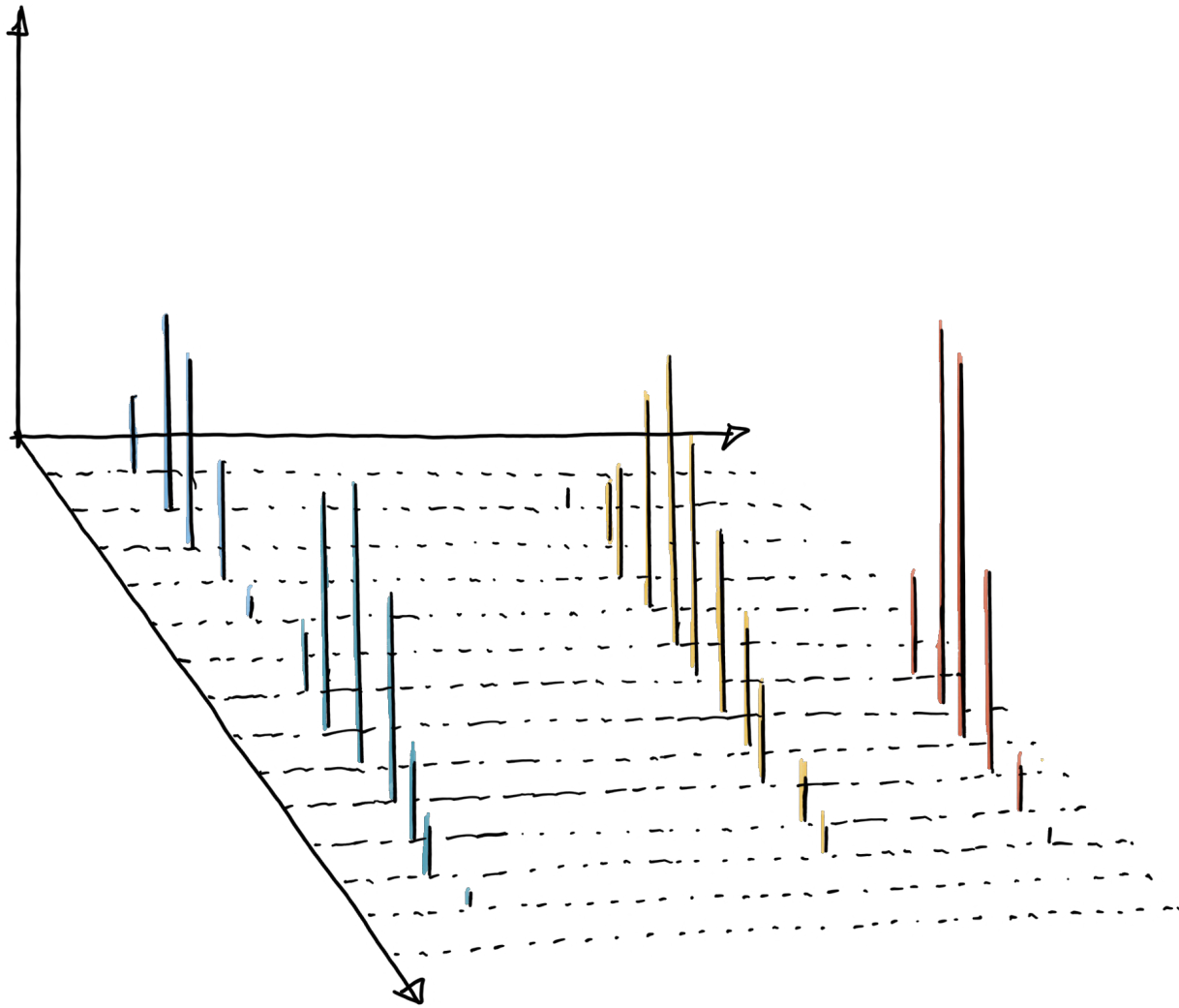
- retention time.
- LC-MS: analyze data along retention time.

# LC-MS data preprocessing

- Chromatographic peak detection
- Alignment
- Correspondence

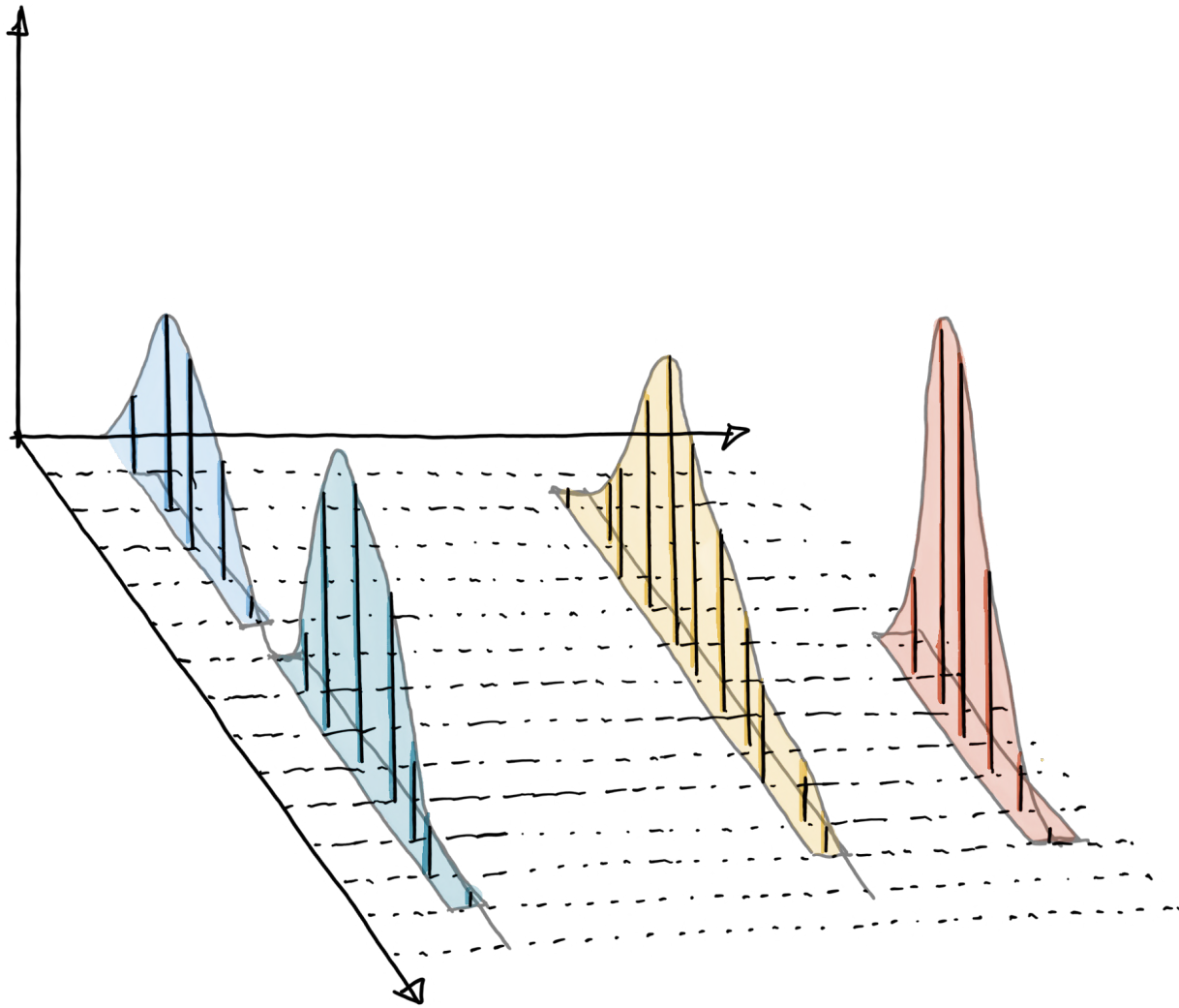
# Chromatographic peak detection

- **Aim:** identify chromatographic peaks in the data.



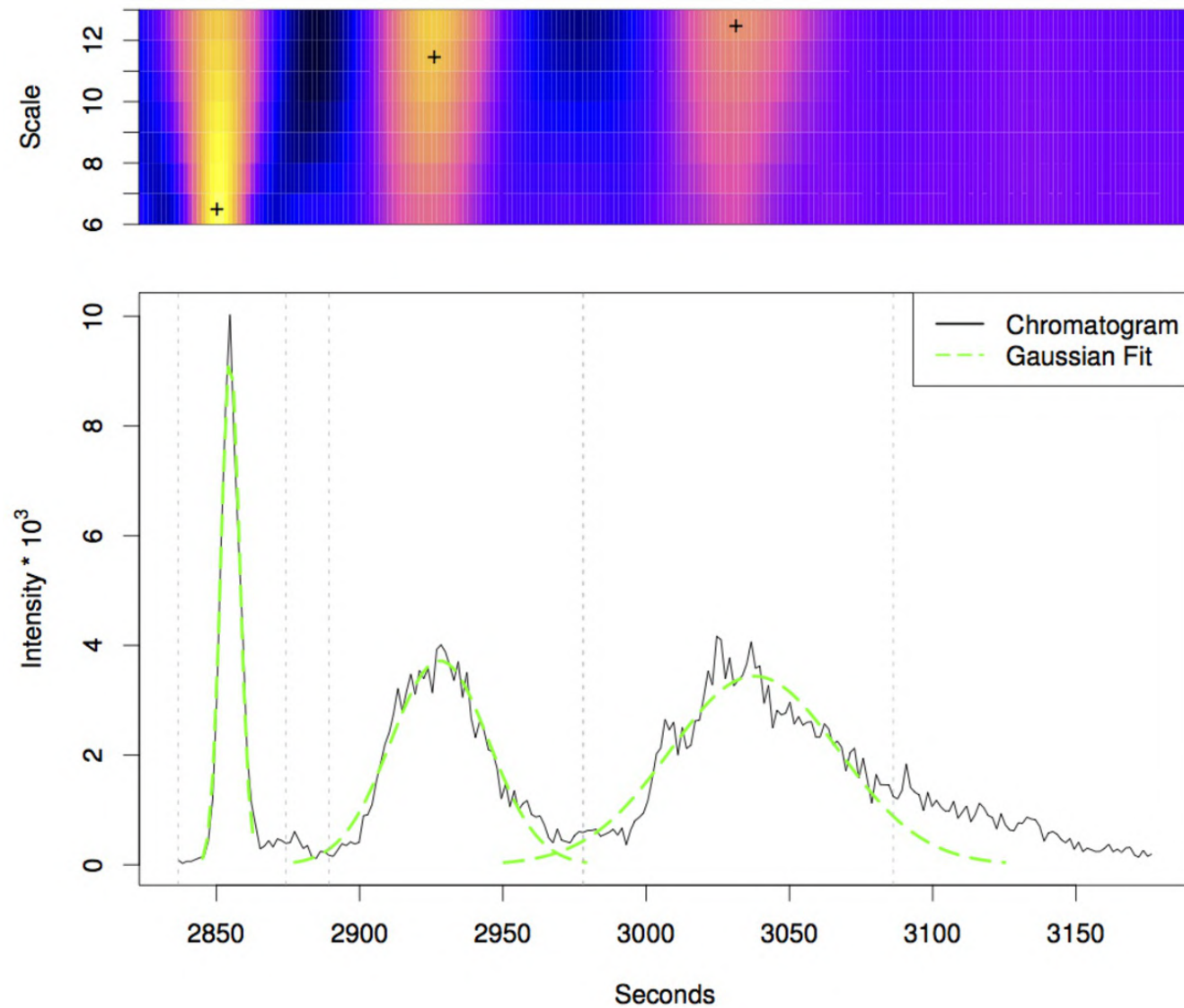
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# Chromatographic peak detection

- **centWave** [Tautenhahn et al. BMC Bioinformatics, 2008]:
- Allows detection of peaks with different rt widths.



# Chromatographic peak detection

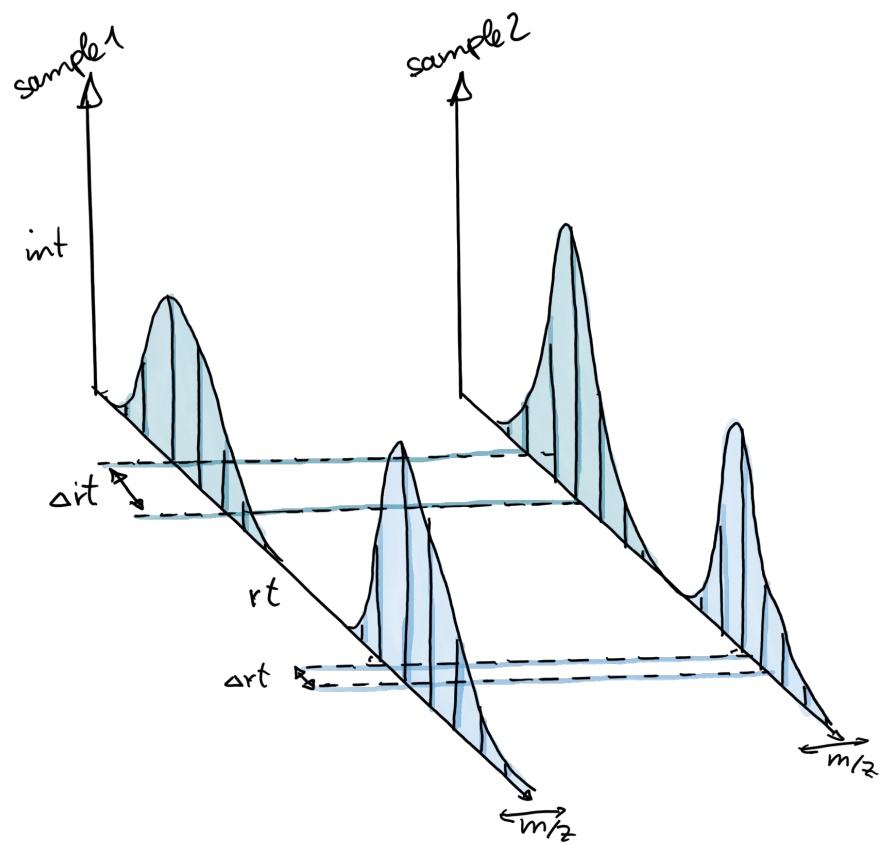
- `MSnbase`: data import with `readMSData`.
- `xcms`: peak detection with `findChromPeaks` and algorithm-specific parameter object.

```
cwp <- CentWaveParam(peakwidth = c(2, 10), snthresh = 5)
data <- findChromPeaks(data, param = cwp)
head(chromPeaks(data), n = 3)
```

```
##           mz      mzmin  mzmax      rt rtmin  rtmax      into      intb
## CP001 114.0907 114.0899 114.0929  1.954 0.280  3.907 1559.829 1555.923
## CP002 114.0913 114.0884 114.0929  5.860 4.465  8.650 1890.221 1885.757
## CP003 114.0914 114.0899 114.0929 10.882 8.650 13.114 1950.953 1946.210
##           maxo  sn  sample
## CP001 584.9510 584      1
## CP002 601.8881 601      1
## CP003 691.9580 691      1
```

# Alignment

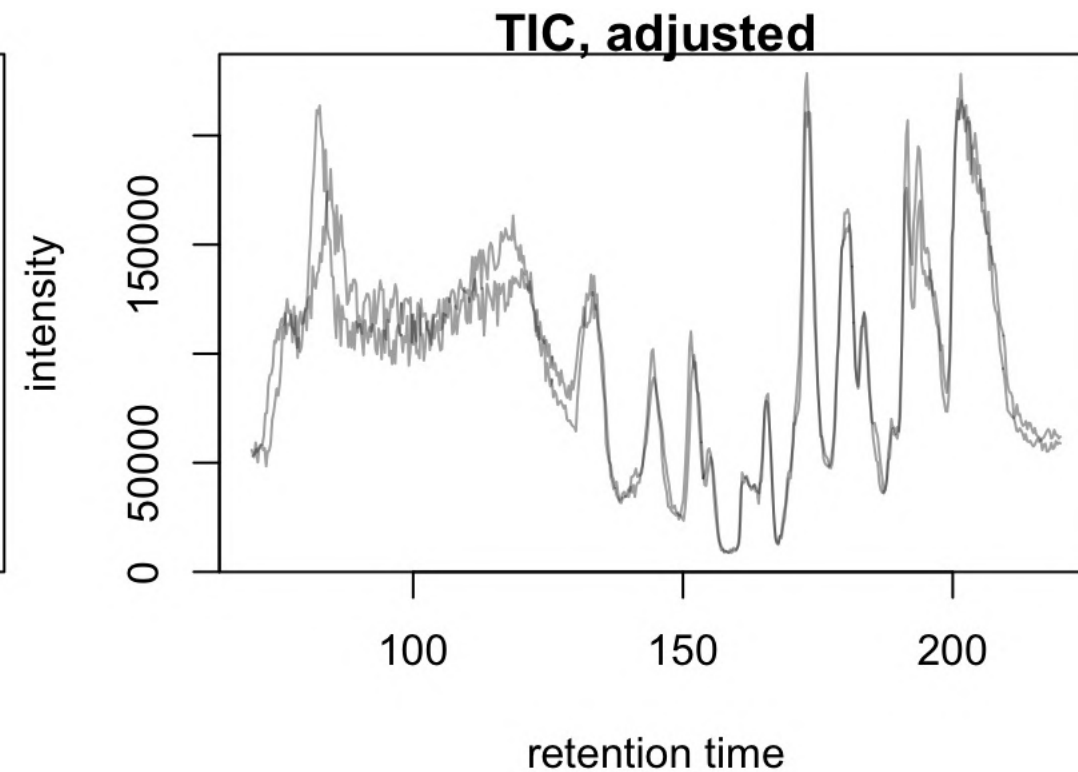
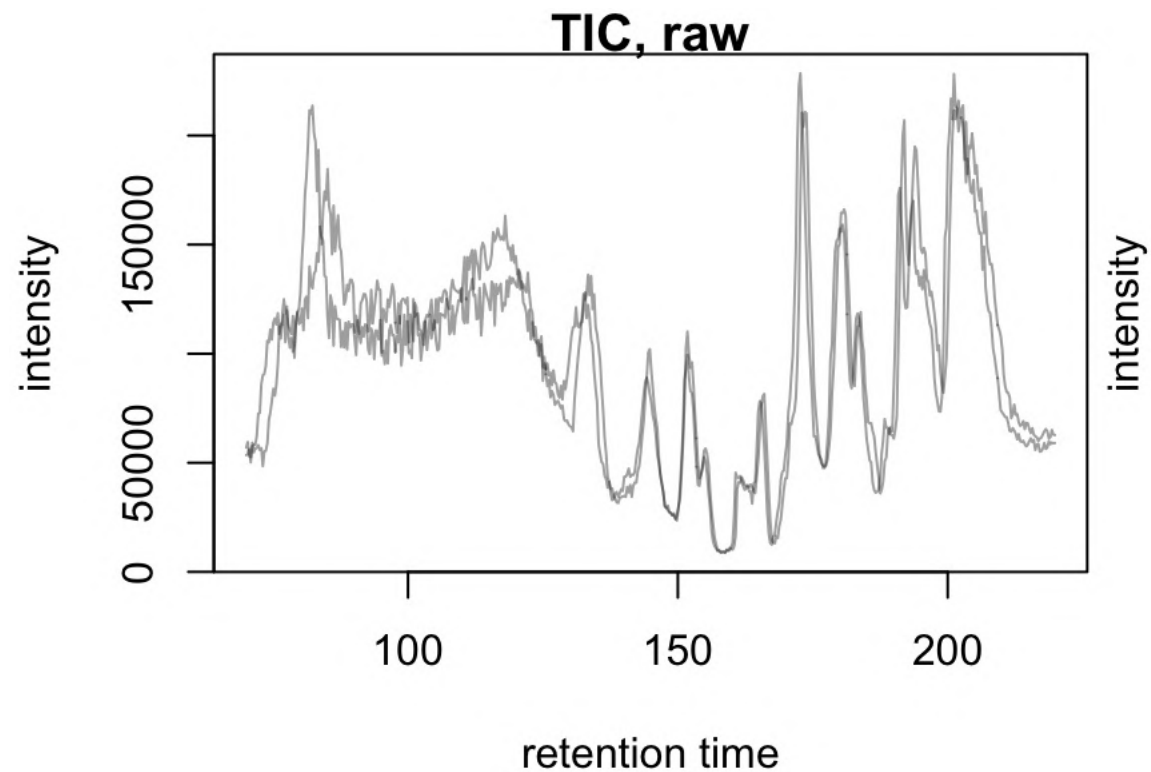
- **Aim:** adjust differences in retention times between samples.
- Same analyte elutes at slightly different time between measurements.



- **Why?** Age of column, temperature ...

# Alignment

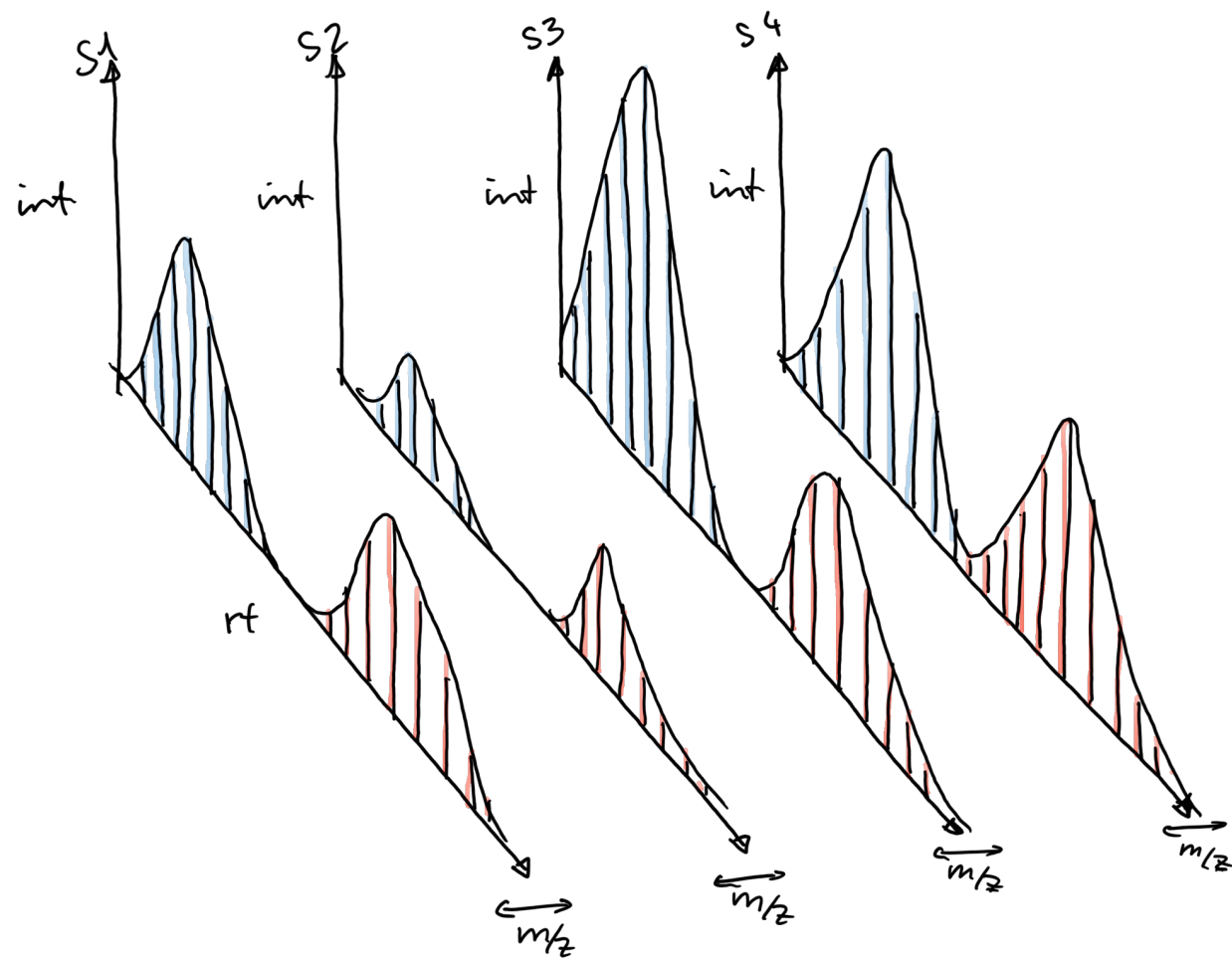
- Many algorithms available [Smith et al. Brief Bioinformatics 2013]
- `xcms`: `adjustRtime` function with `PeakGroupsParam` [Smith et al. Anal. chem. 2006] or `ObiwarParam` [Prince et al. Anal. chem. 2006].





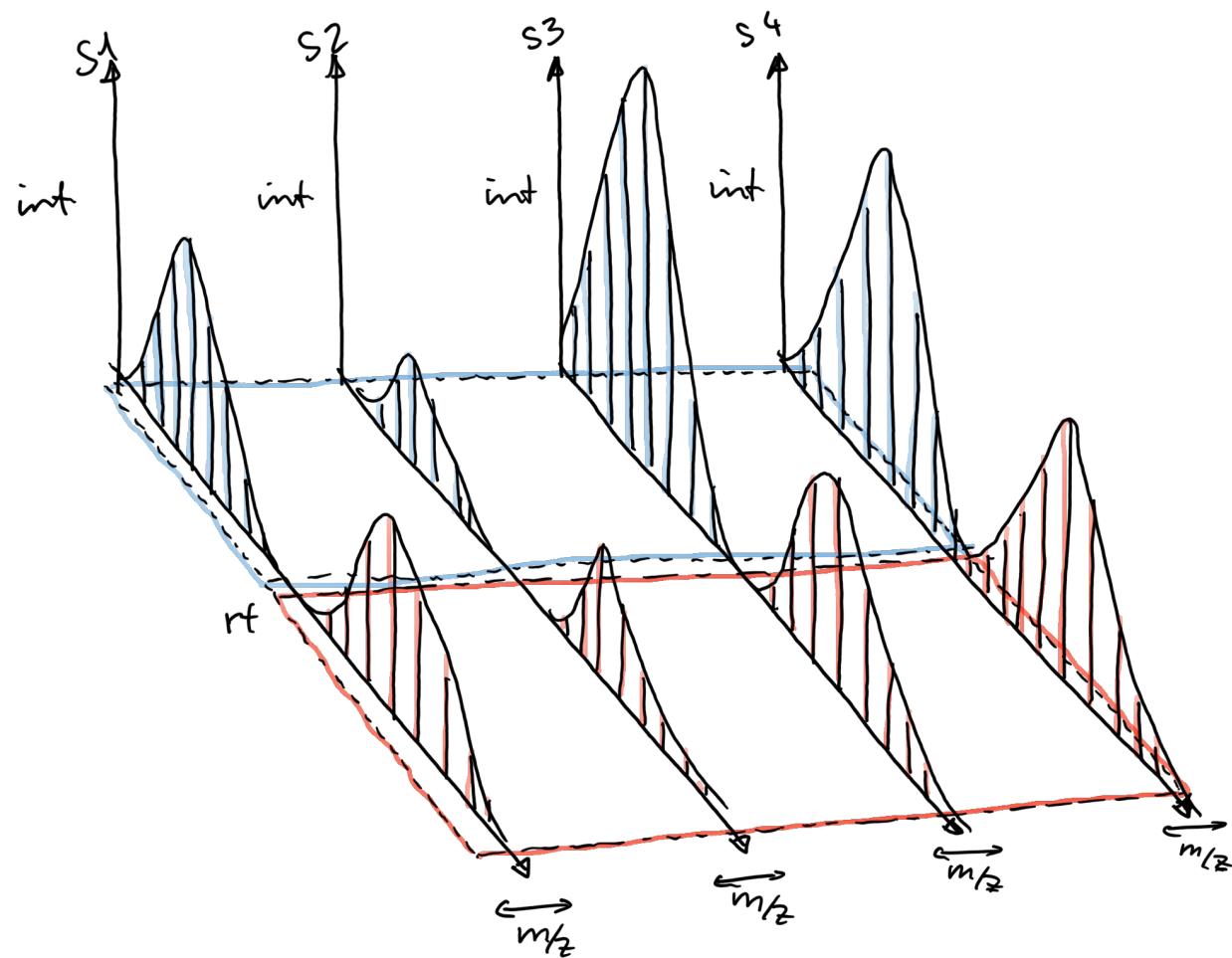
# Correspondence

- **Aim:** group peaks representing same ion species across samples.
- **Result:** matrix of abundances, rows *features*, columns samples.



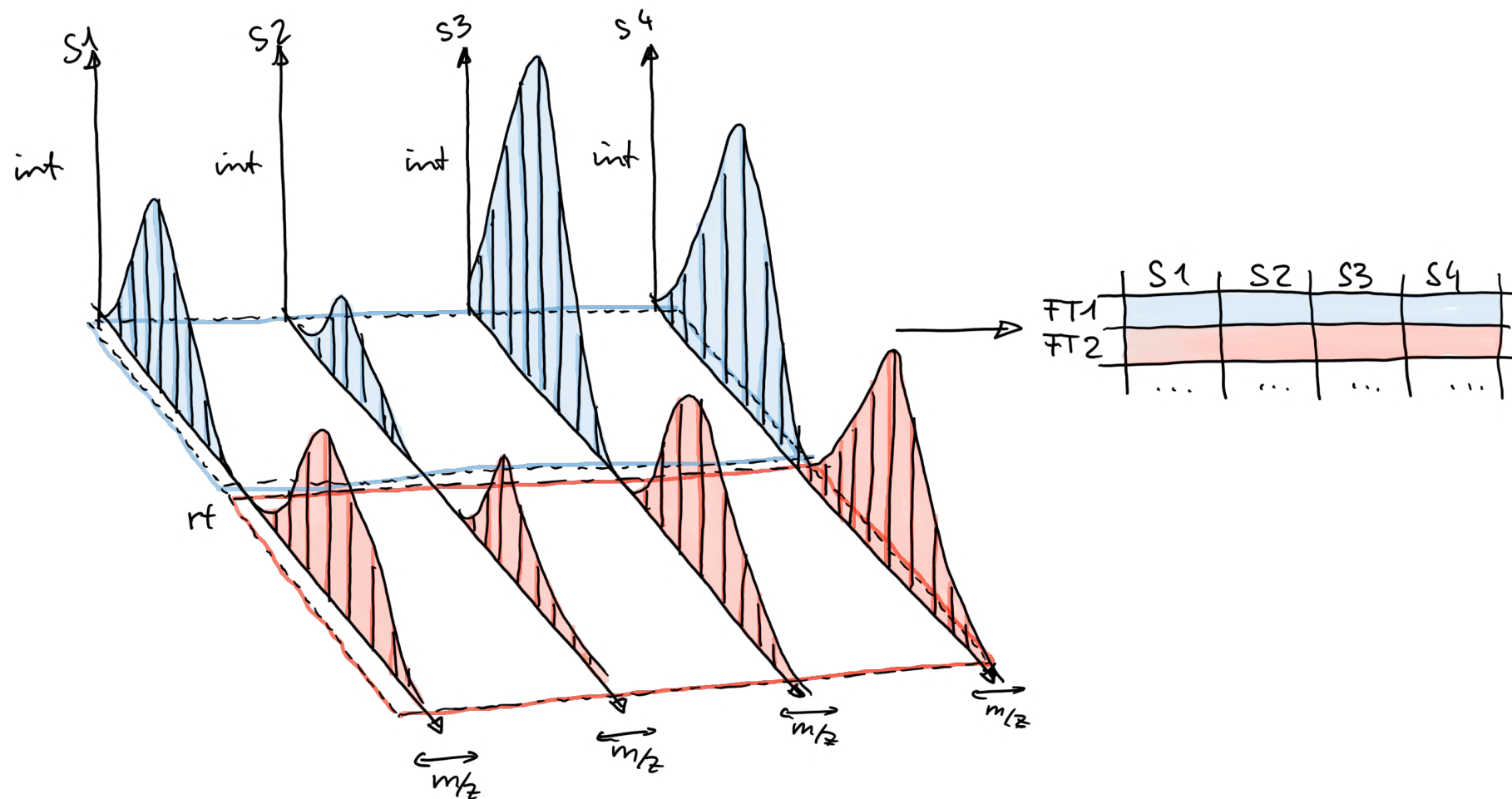
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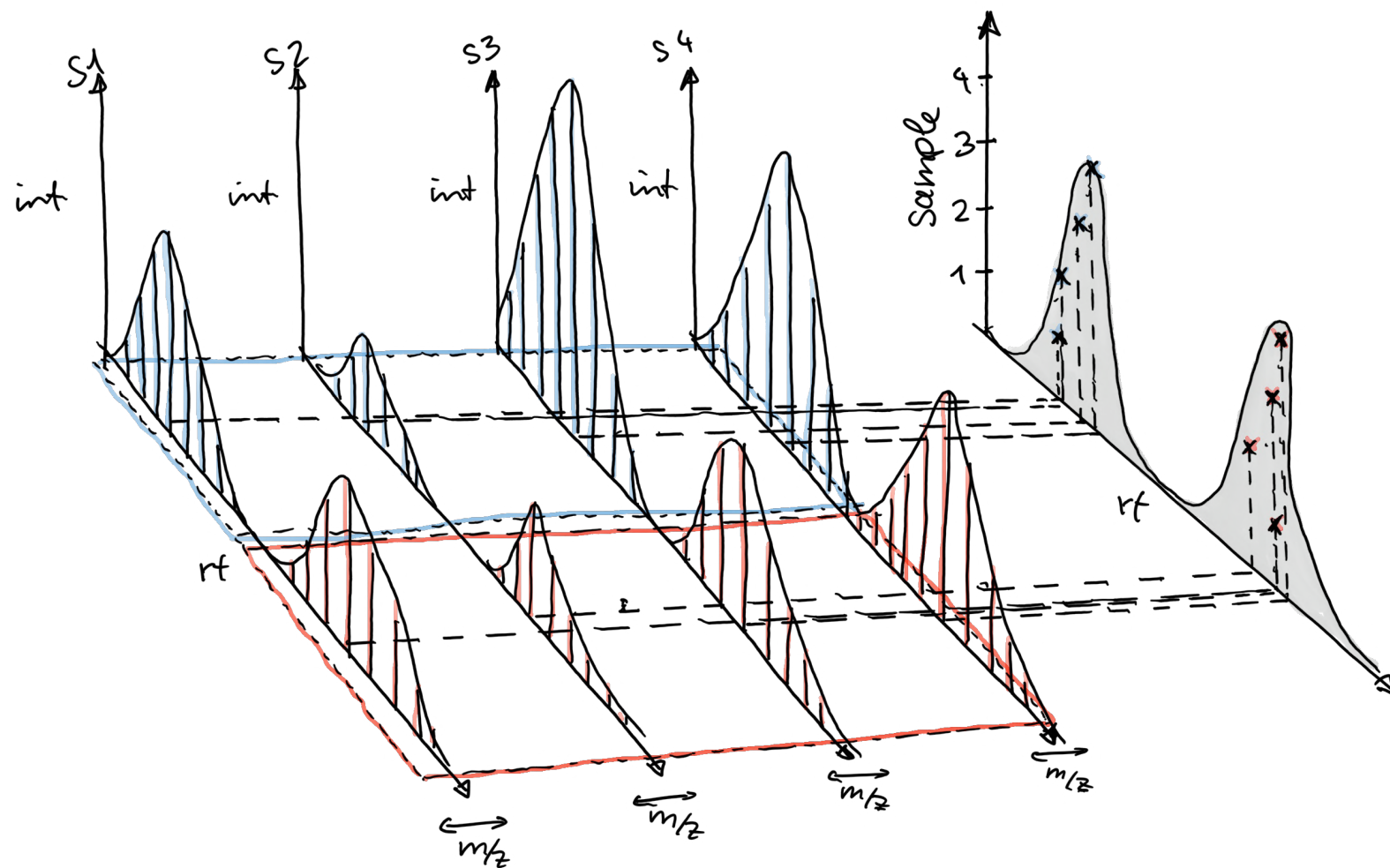


# Correspondence

- `xcms`: `groupChromPeaks` with `NearestPeaksParam` [Katajamaa et al. Bioinformatics 2006] and `PeakDensityParam` [Smith et al. Anal. chem. 2006].

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- *Peak density* approach (for a given  $m/z$  slice):
- Identify regions along  $rt$  with high peak density, group peaks.



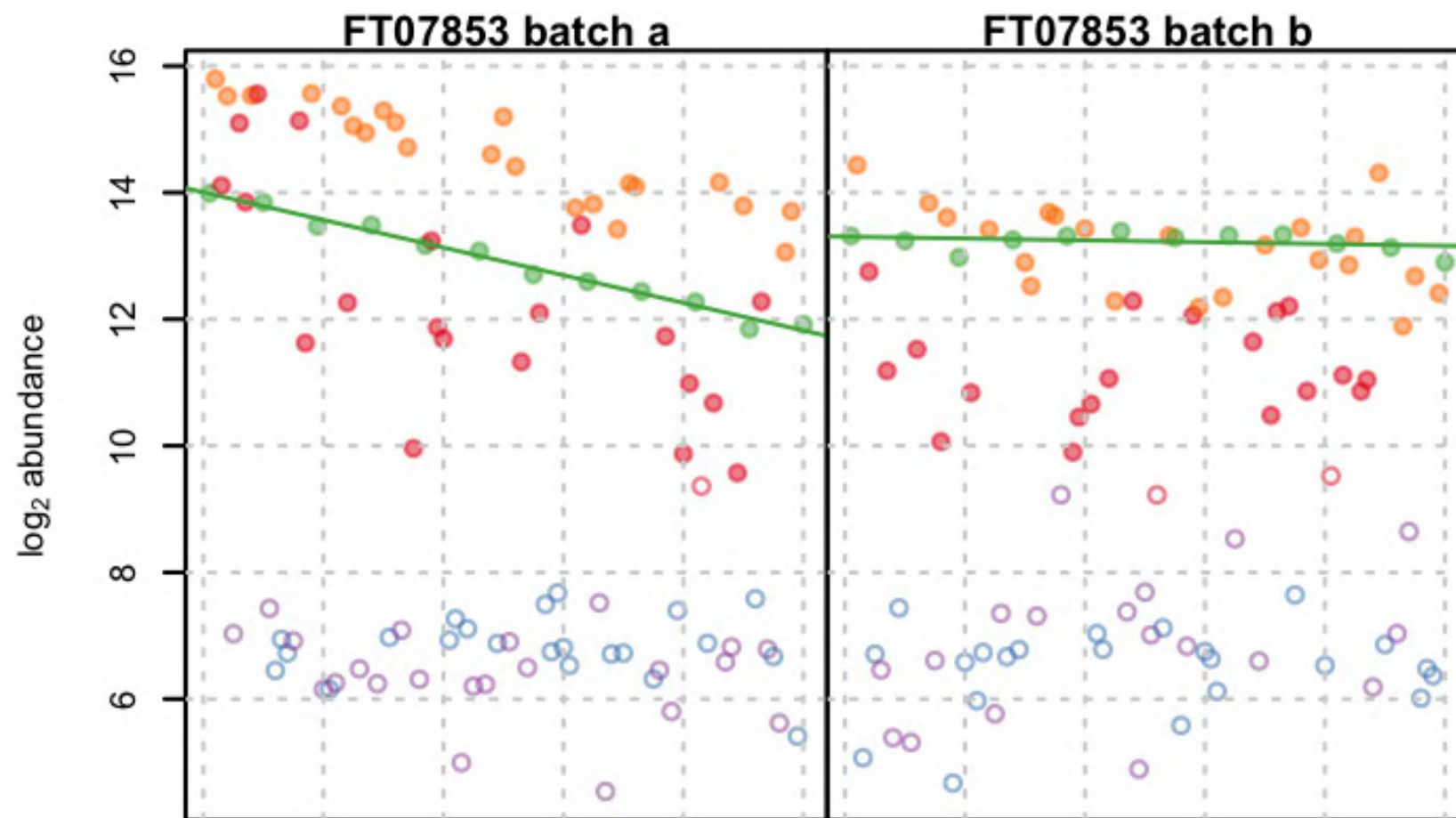
# Preprocessing result

- Numeric matrix with abundances.
- Normalization.
- Identification of features of interest.
- Annotation.

# Normalization

Account for:

- Sample-specific effects.
- Effects related to batch/measurement run.
- Injection order-dependent effects: specific to metabolite.



# Normalization

- Good practice for experimental design:
  - QC samples measured repeatedly.
  - Internal standards.
  - Replicates.
  - Measurement of study samples in randomized order.
- Popular normalization methods:
  - RUV [De Livera et al. Anal. Chem. 2015]
  - linear models [Wehrens et al. Metabolomics 2016]
  - linear and higher order models [Brunius et al. Metabolomics 2016].



# Annotation/Identification

- Feature != metabolite.

```
## DataFrame with 4 rows and 4 columns
##           mzmed           rtmed           POOL_1           POOL_2
##           <numeric>         <numeric>         <numeric>         <numeric>
## FT001 105.041814839707 167.961095453642 229.490739260736 3093.75184315684
## FT002 105.041653033614 157.083057856508 4762.39872227772 6601.45091358641
## FT003 105.069636149683 31.8108067962868 699.723986763237 1033.23232267732
## FT004 105.11027064078 63.7513630255991 20211.2633706294 15839.5504368189
```

- Feature characterized by m/z and retention time.

# Annotation based on mass matching

- m/z is **not** the mass.
- Mass of an  $[M+H]^+$  ion: m/z - mass of 1 hydrogen.
- Different ions from the same compound:  $[M+H]^+$ ,  $[M+Na]^+$ , ...
- Match mass against database.
  - The Human Metabolome Database (HMDB): <https://hmdb.ca>
  - Chemical Entities of Biological Interest: <https://www.ebi.ac.uk/chebi>
  - PubChem <https://pubchem.ncbi.nlm.nih.gov>
  - ...
- Will result in many hits.

# Improved Annotation

Annotate features based on m/z **and**:

- **retention time**: requires measurement of compound/standard on the same LC-MS setup.
- **MS2 spectrum**:
  - Requires LC-MS/MS data (DDA or DIA).
  - Reference spectrum has to be available in database.

# Afternoon metabolomics lab

- LC-MS data handling (MSnbase).
- LC-MS data preprocessing using xcms.

