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Using cellHTS2 for cell based assays

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A LIFE OF SCIENCE

- analyze high throughput cell-based assays with low complexity readouts
e.g. plate reader assays, luminescence assays...
- from raw data to annotated hit list
- data preprocessing, normalization
- data quality assessment
- replicate scoring, annotation
- analysis audit trail

- single color assays
- multi color assays
- activator or inhibitor type assays
- dual way assays
- independent of the instrument, as long as the output is ASCII text
- not coupled to a particular screening library, or to particular organisms

Assumes that every well contains the same type of reagents:

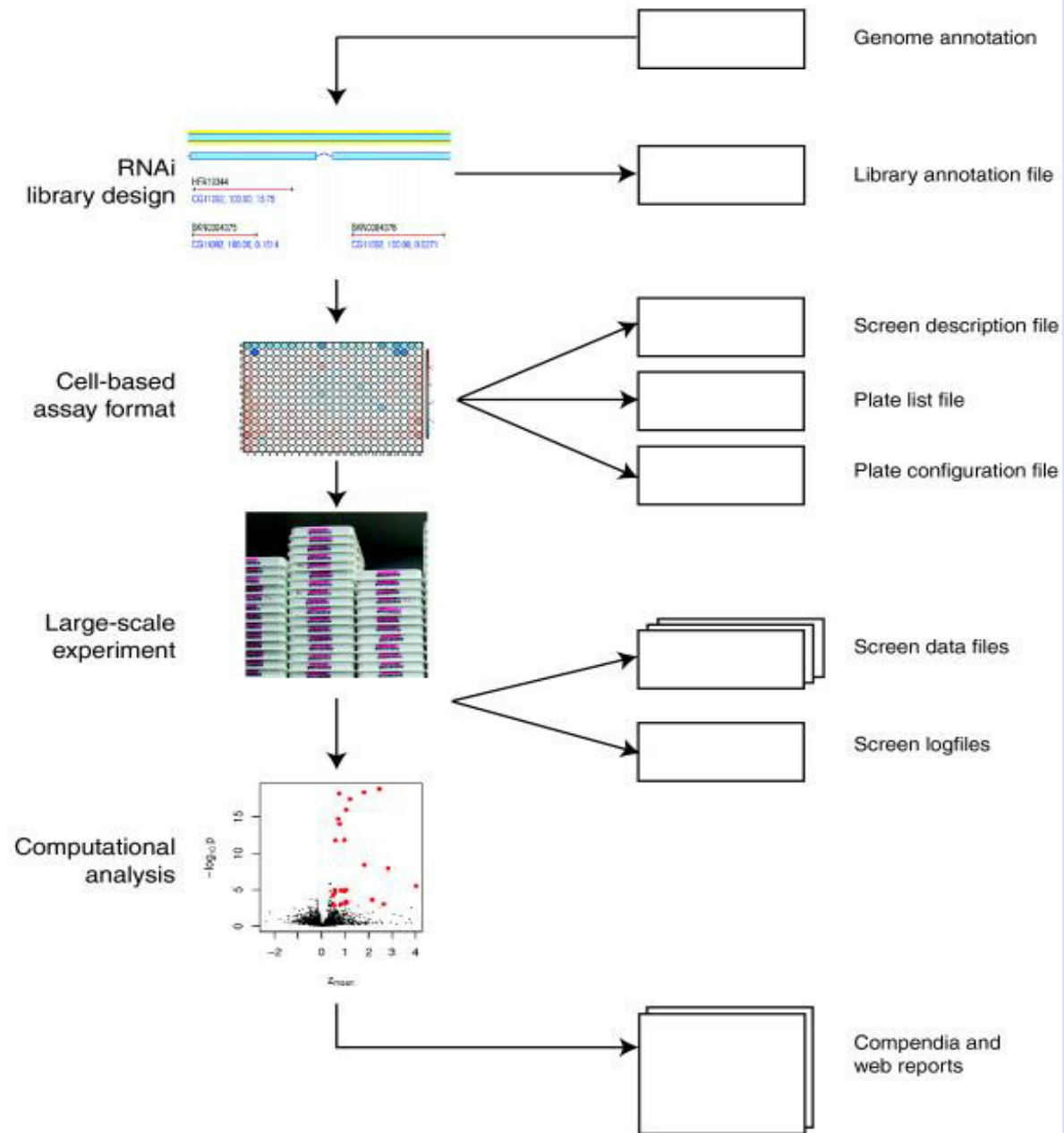
- the same controls (can be circumvented but becomes very awkward...)

- the same library

The screenshot shows a web browser window with the following elements:

- Browser Address Bar:** <http://www.pubmedcentral.nih.gov/articlerender.fcgi?tool=pubmed&pubmedid=>
- Page Navigation:** "Analysis of cell-based RNAi s..." and "Analysis of cell-based R..." tabs.
- Logos:** PubMed Central, BioMed Central, and Genome Biology.
- Journal List:** "Journal List > Genome Biol > v.7(7); 2006"
- Article Information:** "Genome Biol. 2006; 7(7): R66. PMCID: PMC1779553. Published online 2006 July 25. doi: 10.1186/gb-2006-7-7-r66. Copyright © 2006 Boutros et al.; licensee BioMed Central Ltd."
- Title:** "Analysis of cell-based RNAi screens"
- Authors:** "Reviewed by Michael Boutros,¹ Lígia P Brás,^{2,3} and Wolfgang Huber^{1,2}"
- Footnotes:**
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- Open Access License:** "This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited."
- Cited by:** "This article has been cited by other articles in PMC."
- Section Header:** "Abstract"
- Abstract Content:** "RNA interference (RNAi) screening is a powerful technology for functional"
- Page-Footer:** <http://www.pubmedcentral.nih.gov/articlerender.fcgi?tool=pubmed&pubmedid=16869968#supplementary-material-sec>

cellHTS2: structure



Numeric values x_{ijk}

i = wells (e.g. 20,000)

j = different reporters (e.g. 2)

k = different assays (e.g. 5)

Metadata about wells

p_i = plate in which is well i

r_i = row (within plate) of well i

c_i = column (within plate) of well i

siRNA sequence, target gene,

Metadata about reporters

Fluc, Rluc, ...

Metadata about assays k

replicate number

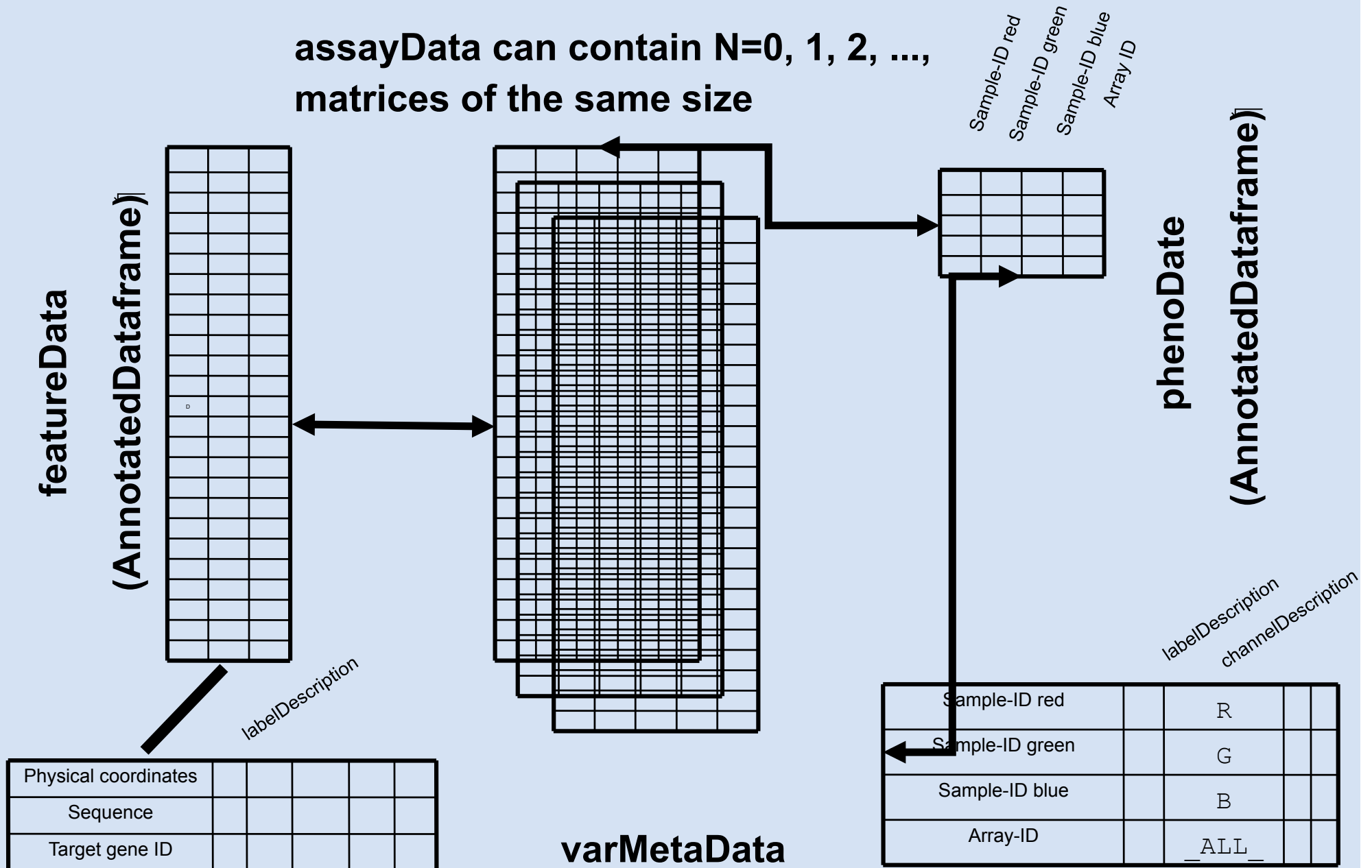
different variants of the assay (e.g. Wnt1_LRP6_Frz8, β -Catenin,

LRP6 Δ E1-4, Wnt1_R-Spondin3, Wnt3a)

date it was done

Data structure: NChannelSet

assayData can contain N=0, 1, 2, ...,
matrices of the same size



- reads data from most established instruments, but the can handle arbitrary data formats through user-defined import functions
- automatically detects plate layout
- raw data files need to be specified using a plate list file

columns Filename, Plate, Replicate, (Batch), ...

- can read collated data from a single file

- 1) Configuration
controls, screen design, flagging
- 2) Annotation
features, e.g. target genes
- 3) Normalization
between and within plates, data transformation
- 4) Replicate scoring and summarization
standardization and reduction to a single value per feature (e.g. z-scores)

human readable audit trail in form of an interactive HTML document can be produced at each of these steps

- screen level: screen description file (MIAME)
- plate level: plate configuration file

regular expressions as annotation rules

Plate	Well	Content
*	*	sample
*	A0[1-2]	other
*	B01	neg
*	B02	pos

visualization of plate layouts

configurationAsScreenPlot()

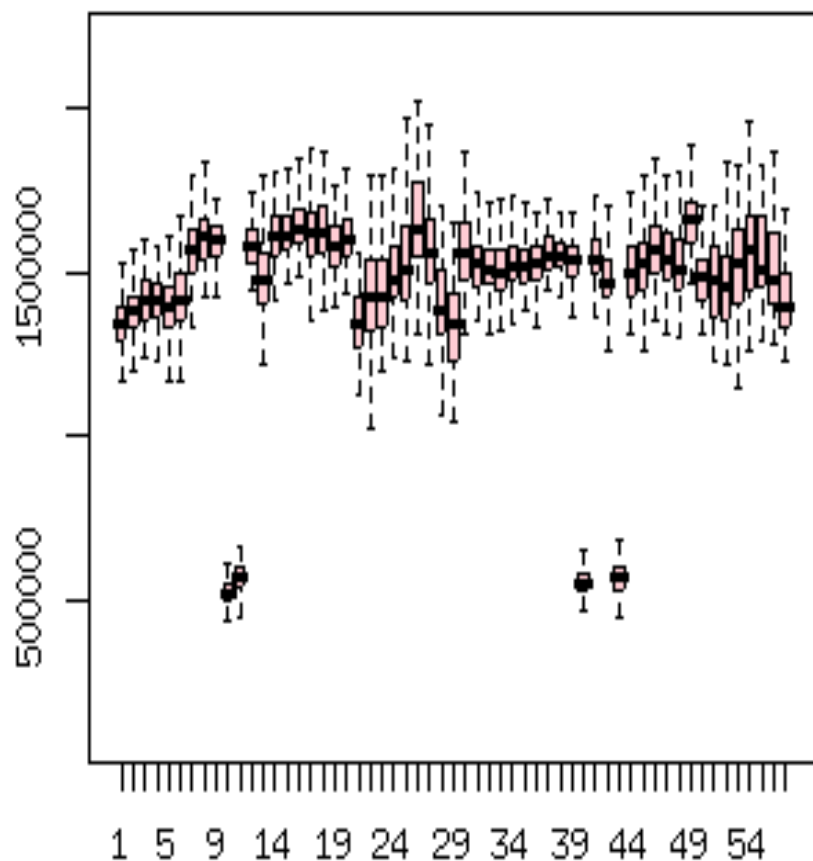
- flagging: screen log file

Annotation: function *annotate()*

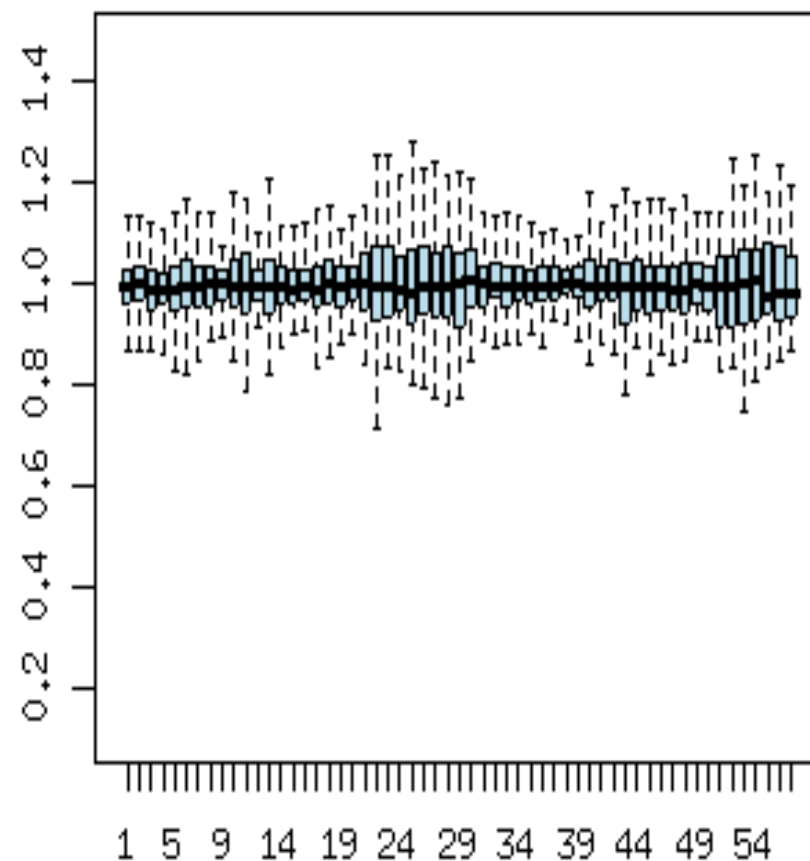
- description of features in annotation file
- mandatory columns: Plate, Well, GeneID
- optional column GeneSymbol: Human-readable name

Normalization: between plates

Raw, replicate 1, channel 1



Normalized, replicate 1, channel 1



From which data points:

- **Based on the intensities of the controls**
if they work uniformly well across all plates
- **Based on the intensities of the samples**
invoke assumptions such as "most genes have no effect", or "same distribution of effect sizes"

Which estimator:

mean vs median vs shorth

standard deviation vs MAD vs IQR

**No universally optimal answer, it depends on the data.
In the best case, it doesn't matter.**

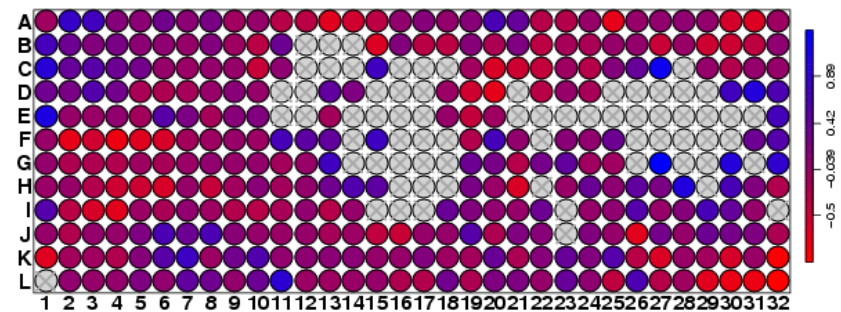
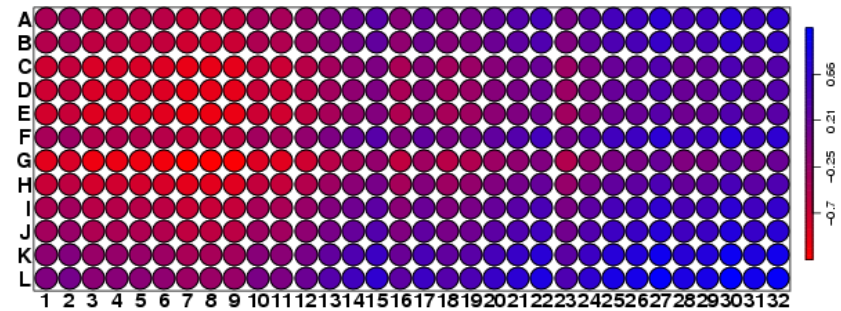
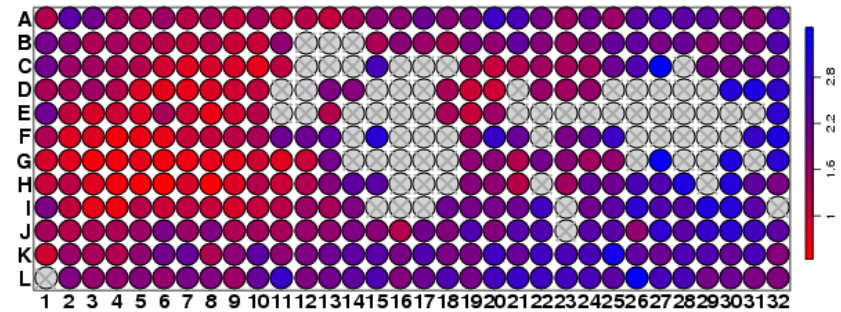
- by estimator of location for each plate:
mean, median, shorth, percent control
- data transformation (optional):
log or linear scale
- Variance adjustment (optional):
by plate, by batch, by experiment
- function *normalizePlates()*

Normalization: *spatial within plates*

B-score:
two-way median polish

r^{th} row
 c^{th} column
 i^{th} plate

Malo et al., Nat. Biotech. 2006



- *scoreReplicates()*
z-score, normalized percent inhibition
- *summarizeReplicates()*
e.g. mean, max, min
- *summarizeChannels()*

HTML report: function *writeReport()*

report

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