

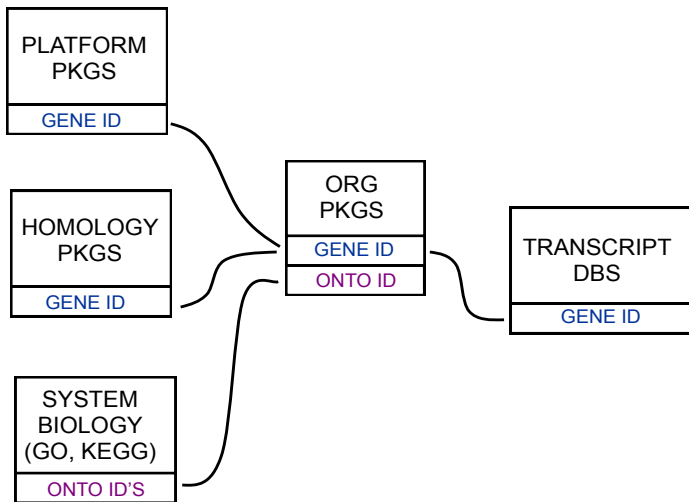
Using the *GenomicFeatures* package

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Bioconductor Annotation Packages: a bigger picture



Bioconductor Sequence Annotations

The GenomicFeatures package

- ▶ Transcript information stored in SQLite databases
- ▶ These databases attempt to represent the relational nature of splicing data correctly
- ▶ Transcript information exposed to user via the *GenomicRanges* infrastructure
- ▶ Parsers are available to construct these databases from biomaRt or the UCSC genome browser. Also there is a generic parser for custom jobs.

GenomicFeatures transcript sources

Constructors

makeTranscriptDbFromBiomart, makeTranscriptDbFromUCSC

```
> library(GenomicFeatures)
> nrow(supportedUCSCtables())
```

```
[1] 24
```

```
> head(supportedUCSCtables(), 10)
```

	track	subtrack
knownGene	UCSC Genes	<NA>
knownGeneOld3	Old UCSC Genes	<NA>
wgEncodeGencodeManualV3	Gencode Genes	Genecode Manual
wgEncodeGencodeAutoV3	Gencode Genes	Genecode Auto
wgEncodeGencodePolyaV3	Gencode Genes	Genecode PolyA
ccdsGene	CCDS	<NA>
refGene	RefSeq Genes	<NA>
xenoRefGene	Other RefSeq	<NA>
vegaGene	Vega Genes	Vega Protein Genes
vegaPseudoGene	Vega Genes	Vega Pseudogenes

TranscriptDb basics

Making a *TranscriptDb* object

```
> mm9KG <-  
+   makeTranscriptDbFromUCSC(genome = "mm9",  
+                             tablename = "knownGene")
```

Saving and Loading

```
> saveFeatures(mm9KG, file="mm9_knownGene.sqlite")  
  
> txdb <-  
+   loadFeatures(system.file("extdata", "mm9_knownGene.sqlite",  
+                             package = "SeattleIntro2010"))
```

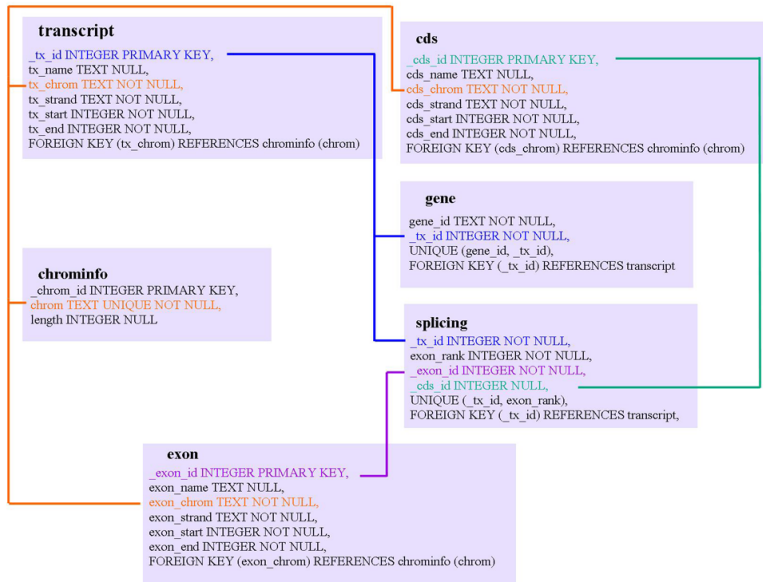
TranscriptDb class

```
> txdb
```

```
TranscriptDb object:
```

```
| Db type: TranscriptDb  
| Data source: UCSC  
| Genome: mm9  
| UCSC Table: knownGene  
| Type of Gene ID: Entrez Gene ID  
| Full dataset: yes  
| transcript_nrow: 49409  
| exon_nrow: 237551  
| cds_nrow: 204831  
| Db created by: GenomicFeatures package from Bioconductor  
| Creation time: 2010-12-07 20:14:52 -0800 (Tue, 07 Dec 2010)  
| GenomicFeatures version at creation time: 1.2.3  
| RSQLite version at creation time: 0.9-4  
| DBSCHEMAVERSION: 1.0
```

TranscriptDb schema



Ungrouped transcript-related information

Extractors

transcripts, exons, cds

```
> tx <- transcripts(txdb)
```

```
> length(tx)
```

```
[1] 49409
```

```
> head(tx, 5)
```

GRanges with 5 ranges and 2 elementMetadata values

	seqnames	ranges	strand	tx_id	tx_name
	<Rle>	<IRanges>	<Rle>	<integer>	<character>
[1]	chr1	[4797974, 4832908]	+	14	uc007afg.1
[2]	chr1	[4797974, 4836816]	+	15	uc007afh.1
[3]	chr1	[4847895, 4887985]	+	16	uc007afi.1
[4]	chr1	[4848119, 4880877]	+	17	uc007afj.1
[5]	chr1	[5073254, 5089858]	+	20	uc007afm.1

seqlengths

chr1	chr2 ...	chrX_random	chrY_random
197195432	181748087 ...	1785075	58682461

Grouped transcript-related information

Extractors

transcriptsBy, exonsBy, cdsBy, intronsByTranscript,
fiveUTRsByTranscript, threeUTRsByTranscript

```
> txExons <- exonsBy(txdb, by="tx")  
> txIntrons <- intronsByTranscript(txdb)  
> txExons[6]
```

GRangesList of length 1

\$6

GRanges with 2 ranges and 3 elementMetadata values

	seqnames	ranges	strand	exon_id	exon_name
	<Rle>	<IRanges>	<Rle>	<integer>	<character>
[1]	chr1	[4483181, 4483816]	-	15	NA
[2]	chr1	[4481009, 4482749]	-	14	NA

exon_rank

	<integer>
[1]	1
[2]	2

seqlengths

Standard GRanges accessors can be used

Accessors

names, length, elementMetaData, width, ranges, start, end

Examples:

```
> head(start(tx))
```

```
[1] 4797974 4797974 4847895 4848119 5073254 5073254
```

```
> head(ranges(txExons), n=1)
```

```
CompressedIRangesList of length 1
```

```
$`1`
```

```
IRanges of length 2
```

	start	end	width
[1]	3203520	3205713	2194
[2]	3195985	3197398	1414

```
> head(elementMetadata(tx), n=2)
```

```
DataFrame with 2 rows and 2 columns
```

tx_id	tx_name
<integer>	<character>

GenomicFeatures exercise 1

Load the transcriptDB object above and then gather the annotations for transcripts as grouped by their genes.

GenomicFeatures exercise 1 solution

```
> library(GenomicFeatures)
> txdb <-
+   loadFeatures(system.file("extdata", "mm9_knownGene.sqlite",
+                             package = "SeattleIntro2010"))
> myTranscripts <- transcriptsBy(txdb, by="gene")
```

Overlapping with transcripts

Methods

`findOverlaps`, `countOverlaps`, `match`, `%in%`, `subsetByOverlaps`

Usage and help:

```
> findOverlaps(query, subject, maxgap = 0L, minoverlap = 1L,  
+             type = c("any", "start", "end"),  
+             select = c("all", "first"))
```

Example

```
> grngs <- GRanges("chr1", gaps(ranges(txIntrons[[7]])), "-")  
> countOverlaps(grngs, tx)
```

```
[1] 5 4 4
```

Using Annotations and Genomic Ranges

- ▶ This slide is to set up for some examples that follow where we want to ask some questions about chrIII and chrV
- ▶ First we just need to determine how long the chromosomes are
- ▶ Then we create a GRanges object to describe the data we want to get
- ▶ Finally we just get a file handle for the actual data

Example

```
> library(GenomicFeatures)
> txdb <- loadFeatures(system.file("extdata", "sacCer2_sgdGene.sqlite",
+                                package = "SeattleIntro2010"))
> t <- transcripts(txdb)
> seqlengths(t)
> ## length of chr3 and 5 is 316617 and 576869
> chr3Len <- seqlengths(t)["chrIII"]
> chr5Len <- seqlengths(t)["chrV"]
> ## Define a GRanges object that describes the sequences we want to re
> gr <- GRanges(seqnames = Rle(c("chrIII", "chrV")),
+               ranges = IRanges(1, width=c(chr3Len, chr5Len)),
+               seqlengths = c(chr3Len, chr5Len))
```

Using Annotations and Genomic Ranges

- ▶ Use `readGappedAlignments` to read in the data
- ▶ Use the `which` parameter to specify `GRanges` that we want in our data.

Example

```
> ## The most convenient way
> grGappedBam <- readGappedAlignments(fl, index=fl,
+                                     which=gr)
> grbam <- grg(grGappedBam)
> seqlengths(grbam) <- c(chr3Len, chr5Len)
> ## I saved this for you previously like this:
> save(grbam, file= "bamReadsChr3andChr5.rda")
```

Practical examples: Ura3 and TIM9

Load in the Data and Annotations

Example

```
> load(system.file("data", "bamReadsChr3andChr5.rda",  
+                  package = "SeattleIntro2010"))  
> ## 1st get the annotations for all exons grouped by gene  
> library(GenomicFeatures)  
> txdb <- loadFeatures(system.file("extdata", "sacCer2_sgdGene.sqlite",  
+                                 package = "SeattleIntro2010"))  
> txs <- transcriptsBy(txdb, by="gene")
```

GenomicFeatures exercise 2

Why did we use `transcriptsBy` instead of `transcripts`?

GenomicFeatures exercise 2 solution

Because we want to consider the ranges associated with each gene, and the `transcripts` function would not distinguish between two actual transcripts that were associated with a single gene.

Practical examples: Ura3 and TIM9

Subset the annotations down to just two genes so that we can overlap with just those ranges.

Example

```
> ## Check if the gene IDs "YEL021W" & "YEL020W-A" are present
> c("YEL021W", "YEL020W-A") %in% names(txs)
> ## Lets extract only two genes
> ind = names(txs) %in% c("YEL021W", "YEL020W-A")
> txpair = txs[ind]
> ## drop strand info. (so we will match both + and - strands)
> sgrbam <- grbam
> strand(sgrbam) <- rep("*", length(strand(sgrbam)))
```

Practical examples: Ura3 and TIM9

Use findOverlaps to see which things match

Example

```
> OL <- findOverlaps(sgrbam, txpair)
> head(matchMatrix(OL))
> ind <- matchMatrix(OL)[,"query"]
> ## Subset to just the data that overlapped
> subData <- grbam[ind]
> seqlengths(subData) <- chr5Len
```

Practical examples: Ura3 and TIM9

We can also do this process in reverse (starting with ranges)

Example

```
> ##starting with the data object
> subData
> ## call findOverlaps()
> OLrev <- findOverlaps(subData, txs)
> matchMatrix(OLrev)
> ## Then extract the genes that overlapped
> ind <- unique(matchMatrix(OLrev)[,"subject"])
> txs[ind]
```

GenomicFeatures exercise 3

Use the code below to load the results from the DE data from Martins talk earlier in the day. Use your gene centric annotation skills to find any gene names for the top 6 gene IDs, and then use your new skills to find the ranges that correspond to these genes.

```
> load(system.file("data", "nbTopTable.rda",  
+                  package = "SeattleIntro2010"))
```


GenomicFeatures exercise 3 solution

```
> ## 1st lets see if any of these genes is even named
> ids = head(nbTopTable)[,1]
> library(org.Sc.sgd.db)
> mget(ids, org.Sc.sgdGENENAME, ifnotfound=NA)
> ## Now lets get the ranges
> ind = names(txs) %in% ids
> txSub = txs[ind]
```

Visualizing the Data

- ▶ Working with `rtracklayer` often involves wrapping things by using the `GRangesForUCSCGenome` wrapper
- ▶ Then you can launch a genome browser from R

Example

```
> library(rtracklayer)
> ## construct a range that describes where you want to look
> range <- GRangesForUCSCGenome(genome="sacCer2", chrom="chrV",
+                               ranges=ranges(subData),
+                               strand = strand(subData))
> # browseGenome(range = range)
> NA
```

Visualizing the Data

- ▶ You can also save files as .bed .gff or .wig files
- ▶ Those files can then be imported into the genome browser

Example

```
> ## You can export your tracks (bed, gff and wig are supported)
> export( range, "GeneRange.bed")
> ## We also saved this for you
> loadedBedFile <- system.file("extdata", "GeneRange.bed",
+                               package = "SeattleIntro2010")
> loadedBedFile
> ## Can upload this into genome browser
```

Visualizing the Data

Lets look at another gene (the last one had two genes that overlapped)

Example

```
> ind = names(txs) %in% "YEL022W"  
> tx = txs[ind]  
> OL <- findOverlaps(grbam, tx)  
> ind <- matchMatrix(OL)[,"query"]  
> subDat <- grbam[ind]
```

Visualizing the Data

`coverage` returns an `rle` with values computed to fit within the entire chromosome. Therefore we have to adjust for that.

Example

```
> ## a shift solution
> cov <- coverage(subData, shift=list(chrV=1-min(start(subData))))
> x <- cov[["chrV"]]
> ## or a slice solution
> x <- slice(coverage(subData), 1L)[["chrV"]][[1]]
```

Visualizing the Data

- ▶ This just defines a plotting function so we can look at the data
- ▶ Both the start and the runValues of the rle have to be plotted

Example

```
> plotCoverage <- function(x, xlab = "Position", ylab = "Coverage",  
+ ...) {  
+   plot(c(start(x)), c(runValue(x)), type = "s", col = "blue",  
+        xlab = xlab, ylab = ylab, ...)  
+ }  
> ## then call out plot function  
> plotCoverage(x)
```

Visualizing the Data

Of course we also could have also plotted this using UCSC's genome browser

Example

```
> sd <- GRangesForUCSCGenome(genome="sacCer2", chrom="chrV",  
+                             ranges=ranges(subData),  
+                             strand = strand(subData))  
> export(sd, "subData.bed")
```

GenomicFeatures exercise 4

Use the data provided to look up data reads that map to LEU2(YCL018W) and NFS1 (YCL017C) from chromosome 3. Then either plot them or display the data in a browser. What do you notice about these two genes?

GenomicFeatures exercise 4 solution

```
> ## Get annots
> ind = names(txs) %in% c("YCL018W", "YCL017C")
> txpair = txs[ind]
> ## Overlap and subset
> OL <- findOverlaps(grbam, txpair)
> ind <- matchMatrix(OL[, "query"])
> subData <- grbam[ind]
> seqlengths(subData) <- chr3Len
> ## Save as a track:
> range <- GRangesForUCSCGenome(genome="sacCer2", chrom="chrIII"
+                               ranges=ranges(subData),
+                               strand = strand(subData))
> export( range, "GeneRange2.bed")
> ## or just plot
> x <- slice(coverage(subData), 1L)[["chrIII"]][[1]]
> plotCoverage(x)
```

Retrieving other data from rtracklayer

Next release watch for FeatureDb objects

But for now we can use rtracklayer

```
> ## create a session object
> session <- browserSession()
> ## then ucscGenomes will list the genomes
> head(ucscGenomes())
> ## Choose one, and assign it into your session object
> genome(session) <- "sacCer2"
> ## NOW you can list the trackNames
> head(trackNames(session))
```

Retrieving other data from rtracklayer

To complete the process, we must create a query and then call `getTable`

```
> ## this information is used to create a query object
> query <- ucscTableQuery(session, "oreganno")
> ## and for each track, you can list the tables it contains
> tableNames(query)
> ## This information can be used to refine/modify your query ob
> tableName(query) <- "oreganno"
> ## Which is all then used to getTable to retrieve the data
> result <- getTable(query)
> head(result)
```

Retrieving other data from rtracklayer

The final step is that you have to take whatever data you got from your query and put it into a GRanges object that can be used with findOverlaps etc.

```
> ranges <- GRanges(seqnames = result[["chrom"]],  
+                   ranges = IRanges(start = result[["chromStart"]],  
+                                     end = result[["chromEnd"]],  
+                   strand = result[["strand"]],  
+                   name = result[["name"]])
```

GenomicFeatures exercise 5

Use rtracklayer to get the track for ensembl genes for yeast. Then use this information along with the reads that mapped to the about the NFS1(YCL017C) gene to determine the coordinates of any ensembl genes are in the same location as the NFS1 gene.

GenomicFeatures exercise 5 solution

```
> query <- ucscTableQuery(session, "ensGene")
> tableNames(query)
> tableName(query) <- "ensGene"
> ensGene <- getTable(query)
> ## then make that into a GRanges object.
> ensRanges <- GRanges(seqnames = ensGene[["chrom"]],
+                       ranges = IRanges(start = ensGene[["txStart"]],
+                                         end = ensGene[["txEnd"]]),
+                       strand = ensGene[["strand"]],
+                       ensId = ensGene[["name"]])
>
> NA
```