

Package ‘simpleSeg’

December 13, 2024

Type Package

Title A package to perform simple cell segmentation

Description Image segmentation is the process of identifying the borders of individual objects (in this case cells) within an image. This allows for the features of cells such as marker expression and morphology to be extracted, stored and analysed. simpleSeg provides functionality for user friendly, watershed based segmentation on multiplexed cellular images in R based on the intensity of user specified protein marker channels. simpleSeg can also be used for the normalization of single cell data obtained from multiple images.

Version 1.8.0

Date 2022-04-21

VignetteBuilder knitr

Encoding UTF-8

biocViews Classification, Survival, SingleCell, Normalization, Spatial

Imports BiocParallel, EBImage, terra, stats, spatstat.geom, S4Vectors, grDevices, SummarizedExperiment, methods, cytomapper

Suggests BiocStyle, testthat (>= 3.0.0), knitr, ggplot2

License GPL-3

RoxygenNote 7.2.3

Config/testthat/edition 3

BugReports <https://github.com/SydneyBioX/simpleSeg/issues>

URL <https://sydneybiox.github.io/simpleSeg/>

<https://github.com/SydneyBioX/simpleSeg>

git_url <https://git.bioconductor.org/packages/simpleSeg>

git_branch RELEASE_3_20

git_last_commit 272f5dc

git_last_commit_date 2024-10-29

Repository Bioconductor 3.20

Date/Publication 2024-12-12

Author Nicolas Canete [aut],
Alexander Nicholls [aut],
Ellis Patrick [aut, cre]

Maintainer Ellis Patrick <ellis.patrick@sydney.edu.au>

Contents

generateBPParam	2
normalizeCells	2
simpleSeg	3

Index	6
--------------	----------

generateBPParam	<i>Utility function to generate BPPARAM object.</i>
-----------------	---

Description

Utility function to generate BPPARAM object.

Usage

```
generateBPParam(cores = 1)
```

Arguments

cores Desired number of cores for BPPARAM object.

Value

A BPPARAM object.

normalizeCells	<i>Normalizes and transforms cell data in preparation for clustering (accepts dataframe, SingleCellExperiment and SpatialExperiment).</i>
----------------	---

Description

Normalizes and transforms cell data in preparation for clustering (accepts dataframe, SingleCellExperiment and SpatialExperiment).

Usage

```
normalizeCells(
  cells,
  markers = NULL,
  assayIn = NULL,
  assayOut = "norm",
  imageID = "imageID",
  transformation = NULL,
  method = NULL,
  cores = 1
)
```

Arguments

cells	A Dataframe of SingleCellExperiment or SpatialExperiment containing cells and features to be normalized/transformed
markers	A list containing the names of cell markers which will be normalized and/or transformed.
assayIn	If input is a SingleCellExperiment or SpatialExperiment with multiple assays, specify the assay to be normalized and/or transformed.
assayOut	If input is a SingleCellExperiment or SpatialExperiment, the new of the normalized data.
imageID	If input is a SingleCellExperiment or SpatialExperiment, this is the name of the image ID variable in order to stratify. cells correctly
transformation	The transformation/s to be performed, default is NULL, accepted values: 'asinh' and 'sqrt'.
method	The normalization method/s to be performed, default is NULL, accepted values: 'mean', 'minMax', 'trim99', 'PC1'.
cores	The number of cores for parallel processing.

Value

returns a dataframe with individual cells as rows and features as columns.

Examples

```
library(cytomapper)
data("pancreasSCE")
cells.normalized <- normalizeCells(
  cells = pancreasSCE,
  markers = c("CD99", "PIN", "CD8a", "CDH"),
  assayIn = "counts",
  assayOut = "normCounts",
  imageID = "ImageNb",
  transformation = "asinh",
  method = "trim99"
)
```

simpleSeg

Perform simple segmentation of multiplexed cellular images

Description

Perform simple segmentation of multiplexed cellular images

Usage

```
simpleSeg(
  image,
  nucleus,
  cellBody = "dilate",
  sizeSelection = 10,
```

```

smooth = 1,
transform = NULL,
watershed = "intensity",
tolerance = NULL,
ext = 1,
discSize = 3,
tissue = NULL,
pca = FALSE,
cores = 1
)

```

Arguments

image	An image or list of images or CytoImageList to be read into the function.
nucleus	The marker or list of markers corresponding to the nuclei.
cellBody	Method of cytoplasm identification. Can be 'none', 'dilate', 'discModel' or the name of a dedicated cytoplasm marker
sizeSelection	Minimum pixels for an object to be recognized as a cell and not noise.
smooth	The amount of Gaussian smoothing to be applied to the image/s
transform	A transformation or list of transformations and normalizations to be performed prior to nuclei or cytoplasm identification. Accepted vales: "sqrt", "asinh", "norm99", "maxThresh" and "tissueMask". Tissue mask may be used when the sample does not take up the entirety of the image (typically a circular sample inside the image. When tissue mask is specified the background noise present outside the sample area is removed).
watershed	Method used to perform watersheding. Accepted values: "intensity", "distance" or "combine".
tolerance	The minimum height of the object in the units of image intensity between its highest point (seed) and the point where it contacts another object (checked for every contact pixel). If the height is smaller than the tolerance, the object will be combined with one of its neighbors, which is the highest. Tolerance should be chosen according to the range of x. Default value is 1, which is a reasonable value if x comes from distmap.
ext	Radius of the neighborhood in pixels for the detection of neighboring objects. Higher value smooths out small objects.
discSize	The size of dilation around nuclei to create cell disc or capture cytoplasm
tissue	Channels to be used to create the tissue mask if specified in transforms.
pca	Whether to run PCA on aggregated nucleus markers in order to detect the cellular nuclei.
cores	The number of cores for parallel processing or a BPPARAM object

Value

A list of image masks

Examples

```

library(cytomapper)
data("pancreasImages")
masks <- simpleSeg(pancreasImages,

```

```
nucleus = "H3",  
cellBody = "discModel",  
sizeSelection = 8,  
smooth = 1.2,  
transform = "sqrt",  
watershed = "combine",  
tolerance = 1, ext = 1,  
discSize = 3,  
cores = 5  
)
```

Index

`generateBPParam`, [2](#)

`normalizeCells`, [2](#)

`simpleSeg`, [3](#)