# Package 'Rbec'

November 6, 2024

Type Package			
<b>le</b> Rbec: a tool for analysis of amplicon sequencing data from synthetic microbial communities			
<b>Version</b> 1.14.0			
<b>Description</b> Rbec is a adapted version of DADA2 for analyzing amplicon sequencing data from synthetic communities (SynComs), where the reference sequences for each strain exists. Rbec can not only accurately profile the microbial compositions in SynComs, but also predict the contaminants in SynCom samples.			
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<pre>Imports Rcpp (&gt;= 1.0.6), dada2, ggplot2, readr, doParallel, foreach,     grDevices, stats, utils</pre>			
LinkingTo Rcpp			
RoxygenNote 7.1.1			
biocViews Sequencing, MicrobialStrain, Microbiome			
Suggests knitr, rmarkdown			
VignetteBuilder knitr			
git_url https://git.bioconductor.org/packages/Rbec			
git_branch RELEASE_3_20			
git_last_commit 5acc5f9			
git_last_commit_date 2024-10-29			
Repository Bioconductor 3.20			
Date/Publication 2024-11-05			
Author Pengfan Zhang [aut, cre]			
Maintainer Pengfan Zhang <pre>pzhang@mpipz.mpg.de&gt;</pre>			
Contents			
Contam_detect	2 3		
Index	5		

2 Contam\_detect

Contam\_detect

Reference-based error correction of amplicon sequencing data

## Description

This function is designed for predicting the contaminated samples

## Usage

```
Contam_detect(log_file, outdir, outlier_constant=1.5)
```

## **Arguments**

#### **Details**

Ruben Garrido-Oter's group, Plant-Microbe interaction, Max Planck Institute for Plant Breeding Research

#### Value

Returns a plot showing the distribution of percentage of corrected reads across the whole sample set and a summary file recording which samples might be contaminated

## Author(s)

Pengfan Zhang

## **Examples**

```
#log_file <- system.file("extdata", "rbec_test.list", package = "Rbec")
log_path <- list.files(paste(path.package("Rbec"),
   "extdata/contamination_test", sep="/"),
   recursive=TRUE, full.names=TRUE)
log_file <- tempfile()
writeLines(log_path, log_file)
Contam_detect(log_file, tempdir())</pre>
```

Rbec 3

# Description

This function corrects the amplicon sequencing data from synthetic communities where the reference sequences are known a priori

# Usage

Rbec(fastq, reference, outdir, threads=1, sampling\_size=5000, ascii=33, min\_cont\_obs\_abd=200, min\_

# Arguments

fastq	the path of the fastq file containg merged amplicon sequencing reads (Ns are not allowed in the reads)	
reference	the path of the unique reference sequences, each sequence must be in one line (Ns are not allowed in the sequences)	
outdir	the output directory, which should be created by the user	
threads	the number of threads used, default 1	
sampling_size	the sampling size for calculating the error matrix, default 5000	
ascii	ascii characters used to encode phred scores (33 or 64), default 33	
min_cont_obs_abd		
	the minimum oberseved abundace of unique tags for detecting contamination sequences, default 200	
min_cont_abd	the relative abundance of unique tgas for detecting contamination sequences that can't be corrected by any of the references, default $0.03$	
min_E	the minimum expectation of the Possion distribution for the identification of paralogues, default $0.05$	
min_P	the minimum P value threshold of the Possion distribution to correct a read, default $1\text{e-}40$	
ref_seeker	the method for finding the candidate error-producing reference sequence for a tag showing identical lowest K-mer distance to multiple references. 1 for the abundance-based method; 2 for the transition probability-based method, default 1.	
cn	the copy number table documenting the copy number of the marker gene in each strain. Rbec will normalize the strain abundance if the copy number is available	

## **Details**

Ruben Garrido-Oter's group, Plant-Microbe interaction, Max Planck Institute for Plant Breeding Research

4 Rbec

#### Value

lambda\_final.out the lambda value and pvalue of the Poisson distribution for each read error\_matrix\_final.out the error matrix in the final iteration

strain\_table.txt the strain composition of the sample

strain\_table\_normalized.txt the copy-number-normalized strain composition of the sample if the copy number table is provided

contamination\_seq.fna the potential sequences generated by contaminants

rbec.log percentage of corrected reads, which can be used to predict contaminated samples paralogue\_seq.fna paralogue sequences found in each strain except for the reference provided

## Author(s)

Pengfan Zhang

## **Examples**

```
fastq <- system.file("extdata", "test_raw_merged_reads.fastq.gz", package = "Rbec")
ref <- system.file("extdata", "test_ref.fasta", package = "Rbec")
Rbec(fastq=fastq, reference=ref, outdir=tempdir(), threads=1, sampling_size=500, ascii=33)</pre>
```

# Index

 ${\tt Contam\_detect, 2}$ 

Rbec, 3