Package 'methylCC'

December 26, 2024

Title Estimate the cell composition of whole blood in DNA methylation samples

Version 1.21.0

Imports Biobase, GenomicRanges, IRanges, S4Vectors, dplyr, magrittr, minfi, bsseq, quadprog, plyranges, stats, utils, bumphunter, genefilter, methods, IlluminaHumanMethylation450kmanifest, IlluminaHumanMethylation450kanno.ilmn12.hg19

Depends R (>= 3.6), FlowSorted.Blood.450k

Suggests rmarkdown, knitr, testthat (>= 2.1.0), BiocGenerics, BiocStyle, tidyr, ggplot2

Description A tool to estimate the cell composition of DNA methylation whole blood sample measured on any platform technology (microarray and sequencing).

biocViews Microarray, Sequencing, DNAMethylation, MethylationArray, MethylSeq, WholeGenome

VignetteBuilder knitr

RoxygenNote 6.1.1

Encoding UTF-8

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BugReports https://github.com/stephaniehicks/methylCC/

URL https://github.com/stephaniehicks/methylCC/

git_url https://git.bioconductor.org/packages/methylCC

git_branch devel

git_last_commit 109b2f6

git_last_commit_date 2024-10-29

Repository Bioconductor 3.21

Date/Publication 2024-12-26

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Contents

`.extract_raw_data` *Extract raw data*

Description

Extract the methylation values and GRanges objects

Usage

```
.extract_raw_data(object)
```

Arguments

`object`	an object can be a `RGChannelSet`, `GenomicMethylSet` or `BSseq` object

Value

A list preprocessed objects from the `RGChannelSet`, `GenomicMethylSet` or `BSseq` objects to be used in `.preprocess_estimatecc()`.

`.find_dmrs` *Finding differentially methylated regions*

Description

This function uses the `FlowSorted.Blood.450k` whole blood reference methylomes with six cell types to identify differentially methylated regions.

Usage

```
.find_dmrs(verbose = TRUE, gr_target = NULL, include_cpgs = FALSE,
  include_dmrs = TRUE, num_cpgs = 50, num_regions = 50,
  bumphunter_beta_cutoff = 0.2, dmr_up_cutoff = 0.5,
  dmr_down_cutoff = 0.4, dmr_pval_cutoff = 1e-11,
  cpg_pval_cutoff = 1e-08, cpg_up_dm_cutoff = 0,
  cpg_down_dm_cutoff = 0, pairwise_comparison = FALSE,
  mset_train_flow_sort = NULL)
```

Arguments

`verbose`	TRUE/FALSE argument specifying if verbose messages should be returned or not. Default is TRUE.
`gr_target`	Default is NULL. However, the user can provide a GRanges object from the `object` in `estimatecc`. Before starting the procedure to find differentially methylated regions, the intersection of the `gr_target` and GRanges object from the reference methylomes (`FlowSorted.Blood.450k`).
`include_cpgs`	TRUE/FALSE. Should individual CpGs be returned. Default is FALSE.
`include_dmrs`	TRUE/FALSE. Should differentially methylated regions be returned. Default is TRUE. User can turn this to FALSE and search for only CpGs.
`num_cpgs`	The max number of CpGs to return for each cell type. Default is 50.
`num_regions`	The max number of DMRs to return for each cell type. Default is 50.
`bumphunter_beta_cutoff`	The `cutoff` threshold in `bumphunter()` in the `bumphunter` package.
`dmr_up_cutoff`	A cutoff threshold for identifying DMRs that are methylated in one cell type, but not in the other cell types.
`dmr_down_cutoff`	A cutoff threshold for identifying DMRs that are not methylated in one cell type, but methylated in the other cell types.
`dmr_pval_cutoff`	A cutoff threshold for the p-values when identifying DMRs that are methylated in one cell type, but not in the other cell types (or vice versa).
`cpg_pval_cutoff`	A cutoff threshold for the p-values when identifying differentially methylated CpGs that are methylated in one cell type, but not in the other cell types (or vice versa).

`cpg_up_dm_cutoff`
: A cutoff threshold for identifying differentially methylated CpGs that are methylated in one cell type, but not in the other cell types.

`cpg_down_dm_cutoff`
: A cutoff threshold for identifying differentially methylated CpGs that are not methylated in one cell type, but are methylated in the other cell types.

`pairwise_comparison`
: TRUE/FAlSE of whether all pairwise comparisons (e.g. methylated in Granulocytes and Monocytes, but not methylated in other cell types). Default if FALSE.

`mset_train_flow_sort`
: Default is NULL. However, a user can provide a `MethylSet` object after processing the `FlowSorted.Blood.450k` dataset. The default normalization is `preprocessIllumina()`.

Value

A list of data frames and GRanges objects.

`.initializeMLEs` *.initializeMLEs*

Description

Helper functions to initialize MLEs in `estimatecc()`.

Usage

```
.initializeMLEs(init_param_method, n, K, Ys, Zs, a0init, a1init, sig0init,
  sig1init, tauinit)
```

Arguments

`init_param_method`
: method to initialize parameter estimates. Choose between "random" (randomly sample) or "known_regions" (uses unmethyalted and methylated regions that were identified based on Reinus et al. (2012) cell sorted data.). Defaults to "random".

`n`
: Number of samples

`K`
: Number of cell types

`Ys`
: observed methylation levels in samples provided by user of dimension R x n

`Zs`
: Cell type specific regions of dimension R x K

`a0init`
: Default NULL. Initial mean methylation level in unmethylated regions

`a1init`
: Default NULL. Initial mean methylation level in methylated regions

`sig0init`
: Default NULL. Initial var methylation level in unmethylated regions

`sig1init`
: Default NULL. Initial var methylation level in methylated regions

`tauinit`
: Default NULL. Initial var for measurement error

Value

A list of MLE estimates to be used in `estimatecc()`.

`.initialize_theta` *.initialize_theta*

Description

Creates a container with initial theta parameter estimates

Usage

```
.initialize_theta(n, K, alpha0 = NULL, alpha1 = NULL, sig0 = NULL,
  sig1 = NULL, tau = NULL)
```

Arguments

`n`	Number of samples
`K`	Number of cell types
`alpha0`	Default NULL. Initial mean methylation level in unmethylated regions
`alpha1`	Default NULL. Initial mean methylation level in methylated regions
`sig0`	Default NULL. Initial var methylation level in unmethylated regions
`sig1`	Default NULL. Initial var methylation level in methylated regions
`tau`	Default NULL. Initial var for measurement error

Value

A data frame with initial parameter estimates to be used in `.initializeMLEs()`.

`.methylcc_engine` *.methylcc_engine*

Description

Helper function for estimatecc

Usage

```
.methylcc_engine(Ys, Zs, current_pi_mle, current_theta, epsilon, max_iter)
```

Arguments

Ys	observed methylation levels in samples provided by user of dimension R x n
Zs	Cell type specific regions of dimension R x K
current_pi_mle	cell composition MLE estimates of dimension K x n
current_theta	other parameter estimates in EM algorithm
epsilon	Add here.
max_iter	Add here.

Value

A list of MLE estimates that is used in `estimatecc()`.

.methylcc_estep *Expectation step*

Description

Expectation step in EM algorithm for methylCC

Usage

```
.methylcc_estep(Ys, Zs, current_pi_mle, current_theta, meth_status = 0)
```

Arguments

Ys	observed methylation levels in samples provided by user of dimension R x n
Zs	Cell type specific regions of dimension R x K
current_pi_mle	cell composition MLE estimates of dimension K x n
current_theta	other parameter estimates in EM algorithm
meth_status	Indicator function corresponding to regions that are unmethylated (meth_status=0) or methylated (meth_status=1)

Value

List of expected value of the first two moments of the random effects (or the E-Step in the EM algorithm) used in `.methylcc_engine()`

.methylcc_mstep *Maximization step*

Description

Maximization step in EM Algorithm for methylCC

Usage

```
.methylcc_mstep(Ys, Zs, current_pi_mle, current_theta, estep0, estep1)
```

Arguments

Ys	observed methylation levels in samples provided by user of dimension R x n
Zs	Cell type specific regions of dimension R x K
current_pi_mle	cell composition MLE estimates of dimension K x n
current_theta	other parameter estimates in EM algorithm
estep0	Results from expectation step for unmethylated regions
estep1	Results from expectation step for methylated regions

Value

A list of the updated MLEs (or the M-Step in the EM algorithm) used in `.methylcc_engine()`

.pick_target_positions *Pick target positions*

Description

Pick probes from target data using the indices in `dmp_regions`

Usage

```
.pick_target_positions(target_granges, target_object = NULL,
  target_cvg = NULL, dmp_regions)
```

Arguments

target_granges	add more here.
target_object	an optional argument which contains the meta-data for `target_granges`. If `target_granges` already contains the meta-data, do not need to supply `target_object`.
target_cvg	coverage reads for the target object
dmp_regions	differentially methylated regions

Value

A list of GRanges objects to be used in `.preprocess_estimatecc()`

`.preprocess_estimatecc` *.preprocess_estimatecc*

Description

This function preprocesses the data before the `estimatecc()` function

Usage

```
.preprocess_estimatecc(object, verbose = TRUE,
  init_param_method = "random",
  celltype_specific_dmrs = celltype_specific_dmrs)
```

Arguments

`object`	an object can be a `RGChannelSet`, `GenomicMethylSet` or `BSseq` object
`verbose`	TRUE/FALSE argument specifying if verbose messages should be returned or not. Default is TRUE.
`init_param_method`	method to initialize parameter estimates. Choose between "random" (randomly sample) or "known_regions" (uses unmethyalted and methylated regions that were identified based on Reinus et al. (2012) cell sorted data.). Defaults to "random".
`celltype_specific_dmrs`	cell type specific differentially methylated regions (DMRs).

Value

A list of object to be used in estimatecc

`.splitit` *.splitit*

Description

helper function to split along a variable

Usage

```
.splitit(x)
```

Arguments

x	a vector

Value

A list to be used in `find_dmrs()`

.WFun *Helper function to take the product of Z and cell composition estimates*

Description

Helper function which is the product of Z and pi_mle

Usage

```
.WFun(Zs, pi_mle)
```

Arguments

Zs	Cell type specific regions of dimension R x K
pi_mle	cell composition MLE estimates

Value

A list of output after taking the product of Z and cell composition mle estimates to be used in `.methylcc_estep()`.

---

cell_counts *Generic function that returns the cell composition estimates*

Description

Given a estimatecc object, this function returns the cell composition estimates

Accessors for the 'cell_counts' slot of a estimatecc object.

Usage

```
cell_counts(object)

## S4 method for signature 'estimatecc'
cell_counts(object)
```

Arguments

object	an object of class `estimatecc`.

Value

Returns the cell composition estimates

Examples

```
# This is a reduced version of the FlowSorted.Blood.450k
# dataset available by using BiocManager::install("FlowSorted.Blood.450k),
# but for purposes of the example, we use the smaller version
# and we set \code{demo=TRUE}. For any case outside of this example for
# the package, you should set \code{demo=FALSE} (the default).

dir <- system.file("data", package="methylCC")
files <- file.path(dir, "FlowSorted.Blood.450k.sub.RData")
if(file.exists(files)){
    load(file = files)

    set.seed(12345)
    est <- estimatecc(object = FlowSorted.Blood.450k.sub, demo = TRUE)
    cell_counts(est)
 }
```

`estimatecc` *Estimate cell composition from DNAm data*

Description

Estimate cell composition from DNAm data

Usage

```
estimatecc(object, find_dmrs_object = NULL, verbose = TRUE,
  epsilon = 0.01, max_iter = 100, take_intersection = FALSE,
  include_cpgs = FALSE, include_dmrs = TRUE,
  init_param_method = "random", a0init = NULL, a1init = NULL,
  sig0init = NULL, sig1init = NULL, tauinit = NULL, demo = FALSE)
```

Arguments

`object`	an object can be a `RGChannelSet`, `GenomicMethylSet` or `BSseq` object
`find_dmrs_object`	If the user would like to supply different differentially methylated regions, they can use the output from the `find_dmrs` function to supply different regions to `estimatecc`.
`verbose`	TRUE/FALSE argument specifying if verbose messages should be returned or not. Default is TRUE.
`epsilon`	Threshold for EM algorithm to check for convergence. Default is 0.01.

max_iter	Maximum number of iterations for EM algorithm. Default is 100 iterations.
take_intersection	TRUE/FALSE asking if only the CpGs included in object should be used to find DMRs. Default is FALSE.
include_cpgs	TRUE/FALSE. Should individual CpGs be returned. Default is FALSE.
include_dmrs	TRUE/FALSE. Should differentially methylated regions be returned. Default is TRUE.
init_param_method	method to initialize parameter estimates. Choose between "random" (randomly sample) or "known_regions" (uses unmethyalted and methylated regions that were identified based on Reinus et al. (2012) cell sorted data.). Defaults to "random".
a0init	Default NULL. Initial mean methylation level in unmethylated regions
a1init	Default NULL. Initial mean methylation level in methylated regions
sig0init	Default NULL. Initial var methylation level in unmethylated regions
sig1init	Default NULL. Initial var methylation level in methylated regions
tauinit	Default NULL. Initial var for measurement error
demo	TRUE/FALSE. Should the function be used in demo mode to shorten examples in package. Defaults to FALSE.

Value

A object of the class estimatecc that contains information about the cell composition estimation (in the summary slot) and the cell composition estimates themselves (in the cell_counts slot).

Examples

```
# This is a reduced version of the FlowSorted.Blood.450k
# dataset available by using BiocManager::install("FlowSorted.Blood.450k),
# but for purposes of the example, we use the smaller version
# and we set \code{demo=TRUE}. For any case outside of this example for
# the package, you should set \code{demo=FALSE} (the default).

dir <- system.file("data", package="methylCC")
files <- file.path(dir, "FlowSorted.Blood.450k.sub.RData")
if(file.exists(files)){
    load(file = files)

    set.seed(12345)
    est <- estimatecc(object = FlowSorted.Blood.450k.sub, demo = TRUE)
    cell_counts(est)
 }
```

estimatecc-class *the estimatecc class*

Description

Objects of this class store all the values needed information to work with a estimatecc object

Value

`summary` returns the summary information about the cell composition estimate procedure and `cell_counts` returns the cell composition estimates

Slots

`summary` information about the samples and regions used to estimate cell composition

`cell_counts` cell composition estimates

Examples

```
# This is a reduced version of the FlowSorted.Blood.450k
# dataset available by using BiocManager::install("FlowSorted.Blood.450k),
# but for purposes of the example, we use the smaller version
# and we set \code{demo=TRUE}. For any case outside of this example for
# the package, you should set \code{demo=FALSE} (the default).

dir <- system.file("data", package="methylCC")
files <- file.path(dir, "FlowSorted.Blood.450k.sub.RData")
if(file.exists(files)){
    load(file = files)

    set.seed(12345)
    est <- estimatecc(object = FlowSorted.Blood.450k.sub, demo = TRUE)
    cell_counts(est)
 }
```

FlowSorted.Blood.450k.sub *A reduced size of the FlowSorted.Blood.450k dataset*

Description

A reduced size of the `FlowSorted.Blood.450k` dataset

The object was created using the script in /inst and located in the /data folder.

Format

A RGset object with 2e5 rows (probes) and 6 columns (whole blood samples).

offMethRegions *Unmethylated regions for all celltypes*

Description

This is the script used to create the offMethRegions data set. The purpose is use in the estimate_cc() function.

The object was created using the script in /inst and located in the /data folder.

Format

add more here.

onMethRegions *Methylated regions for all celltypes*

Description

This is the script used to create the onMethRegions data set. The purpose is use in the estimate_cc() function.

The object was created using the script in /inst and located in the /data folder.

Format

add more here.

Index