

# Package ‘SRADB’

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**Type** Package

**Title** A compilation of metadata from NCBI SRA and tools

**Version** 1.32.0

**Date** 2016-04-28

**Depends** RSQLite, graph, RCurl

**Imports** GEOquery

**Suggests** Rgraphviz

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**Maintainer** Jack Zhu <zhujack@mail.nih.gov>

**Description** The Sequence Read Archive (SRA) is the largest public repository of sequencing data from the next generation of sequencing platforms including Roche 454 GS System, Illumina Genome Analyzer, Applied Biosystems SOLiD System, Helicos Heliscope, and others. However, finding data of interest can be challenging using current tools. SRADB is an attempt to make access to the metadata associated with submission, study, sample, experiment and run much more feasible. This is accomplished by parsing all the NCBI SRA metadata into a SQLite database that can be stored and queried locally. Fulltext search in the package make querying metadata very flexible and powerful. fastq and sra files can be downloaded for doing alignment locally. Beside ftp protocol, the SRADB has functions supporting fastp protocol (ascp from Aspera Connect) for faster downloading large data files over long distance. The SQLite database is updated regularly as new data is added to SRA and can be downloaded at will for the most up-to-date metadata.

**License** Artistic-2.0

**LazyLoad** yes

**URL** <http://gbnci.abcc.ncifcrf.gov/sra/>

**BugReports** <https://github.com/seandavi/SRADB/issues/new>

**biocViews** Infrastructure, Sequencing, DataImport

**NeedsCompilation** no

**R topics documented:**

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|               |   |
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| SRADB-package | <i>Query NCBI SRA metadata within R or from a local SQLite database</i> |
|---------------|---|

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**Description**

The Sequence Read Archive (SRA) represents largest public repository of sequencing data from the next generation of sequencing platforms including Roche 454 GS System, Illumina Genome Analyzer, Applied Biosystems SOLiD System, Helicos Heliscope, and others. However, finding data of interest can be challenging using current tools. SRADB is an attempt to make access to the metadata associated with submission, study, sample, experiment and run much more feasible. This is accomplished by parsing all the NCBI SRA metadata into a SQLite database that can be stored and queried locally. SRADB is simply a thin wrapper around the SQLite database along with associated tools and documentation. Fulltext search in the package make querying metadata very flexible and powerful. SRA data files (sra or sra-lite) can be downloaded for doing alignment locally. Available BAM files in local or in the Meltzerlab sraDB can be loaded into IGV for visualization easily. The SQLite database is updated regularly as new data is added to SRA and can be downloaded at will for the most up-to-date metadata.

**Details**

```

Package:    SRADB
Type:      Package
Version:   1.9.2
Date:      2012-02-13
License:   What license is it under?
LazyLoad:  yes

```

**Author(s)**

Jack Zhu and Sean Davis

Maintainer: Jack Zhu <zhujack@mail.nih.gov>

**References**

<http://gbnci.abcc.ncifcrf.gov/backup/SRAMetadb.sqlite.gz> <http://dl.dropbox.com/u/51653511/SRAMetadb.sqlite.gz>

**Examples**

```

if(file.exists('SRAMetadb.sqlite')) {

  library(SRADB)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)

  ## Get column descriptions
  a <- colDescriptions(sra_con=sra_con)[1:5,]

  ## Convert SRA experiment accessions to other types
  b <- sraConvert( in_acc=c(" SRR000137", "SRR000138 "), out_type=c('sample'), sra_con=sra_con )

  ## Fulltext search SRA meta data using SQLite fts3 module
  rs <- getSRA (search_terms ='breas* NEAR/2 can*', out_types=c('run','study'), sra_con=sra_con)
  rs <- getSRA (search_terms ='breast', out_types=c('run','study'), sra_con=sra_con)
  rs <- getSRA (search_terms ='breas* can*', out_types=c('study'), sra_con=sra_con)
  rs <- getSRA (search_terms ='MCF7 OR "MCF-7"', out_types=c('sample'), sra_con=sra_con)
  rs <- getSRA (search_terms ='study_title: breas* can*', out_types=c('run','study'), sra_con=sra_con)
  rs <- getSRA (search_terms ='study_title: breas* can*', out_types=c('run','study'), sra_con=sra_con, acc_onl

  ## List fastq file ftp or fasp addresses associated with "SRX000122"
  listSRAfile (in_acc = c("SRX000122"), sra_con = sra_con, fileType = 'sra')
  listSRAfile (in_acc = c("SRX000122"), sra_con = sra_con, fileType = 'sra', srcType='fasp')

  ## Get file size and date from NCBI ftp site for available fastq files associated with "SRS012041","SRS000290"
  ## Not run:
  getSRAinfo (in_acc=c("SRS012041","SRS000290"), sra_con=sra_con, sraType='sra')

  ## End(Not run)

  ## Download sra files from NCBI SRA using ftp protocol:
  getSRAfile( in_acc = c("SRR000648","SRR000657"), sra_con = sra_con, destDir = getwd(), fileType = 'sra' )

```

```

## Download fastq files from EBI using ftp protocol:
getSRAfile( in_acc, sra_con, destDir = getwd(), fileType = 'fastq', srcType = 'ftp', makeDirectory = FALSE,

## Download fastq files from EBI ftp site using fasp protocol:
## Not run:
ascpCMD <- 'ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty'
getSRAfile( in_acc, sra_con, fileType = 'fastq', srcType = 'fasp', ascpCMD = ascpCMD )
## End(Not run)

## Start IGV from R if no IGV running
## Not run: startIGV(memory='mm')

## load BAM files to IGV
## Not run:
exampleBams = file.path(system.file('extdata',package='SRADB'), dir(system.file('extdata',package='SRADB')
sock <- IGVsocket()
IGVload(sock,exampleBams)

## End(Not run)
## Change the IGV genome
## Not run:
IGVgenome(sock,genome='hg18')

## End(Not run)
## Go to a specified region in IGV
## Not run:
IGVgoto(sock,'chr1:1-10000')
IGVgoto(sock,'TP53')

## End(Not run)

## Make a snapshot of the current IGV window
## Not run:
IGVsnapshot(sock)
dir()

## End(Not run)

## create a graphNEL object from SRA accessions, which are full text search results of terms 'primary thyroid
g <- sraGraph('primary thyroid cell line', sra_con)

## Not run:
library(Rgraphviz)
attrs <- getDefaultAttrs(list(node=list(fillcolor='lightblue', shape='ellipse')))
plot(g, attrs=attrs)

## End(Not run)

dbDisconnect(sra_con)

} else {
  print("use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file
and then rerun the example")
}

```

---

ascpR

*Fasp file downloading using the ascp command line program*


---

## Description

This function downloads files by fasp protocol using Aspera's ascp command line program, which is include in Aspera Connect software (<http://www.asperasoft.com/>).

## Usage

```
ascpR( ascpCMD, ascpSource, destDir = getwd() )
```

## Arguments

|            |   |
|------------|---|
| ascpCMD    | ascp main commands, which should be constructed by a user according to the actual installation of Aspera Connect in the system, with proper options to be used. Example commands: "ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty" (Linux) or ""'/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera Connect.app/Contents/Resources/asperaweb_id_dsa.putty'" (Mac OS X). More about ascp please see the help ('ascp -h' in a shell). |
| ascpSource | character vector of fasp file sources for the ascp command, e.g. era-fasp@fasp.sra.ebi.ac.uk:vol1/fastq (EBI), anonftp@ftp-trace.ncbi.nlm.nih.gov:/sra/sra-instant/reads/ByExp/sra/SRX/SRX000/SRX0001 (NCBI).   |
| destDir    | destination directory to save downloaded files.   |

## Details

The function takes advatage of Aspera's fasp transport technology (<http://www.asperasoft.com/>), which provides high-speed transferring large files over the Internet. Due to complexity with options with ascp and installation difference between different systems, this funciton asks users to supply main ascp comands. Users who are not familiar with ascp command line program should have IT support personnel to install the software and constrect main ascp comands.

## Value

A data.frame containing all matched SRA accessions and ftp or fasp addresses.

## Author(s)

Jack Zhu <[zhujack@mail.nih.gov](mailto:zhujack@mail.nih.gov)>

## References

<http://www.asperasoft.com/>

## See Also

[ascpSRA](#), [getSRAfile](#), [getFASTQinfo](#), [getSRAinfo](#)

**Examples**

```

if( file.exists('SRAMetadb.sqlite') ) {
  ## Not run:
  library( SRAdb )
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)
  rs <- getFASTQinfo (in_acc=c("SRR000648","SRR000657"), srcType='fasp')

  ascpSource <- rs$fasp
  ascpCMD <- 'ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty'
  ## common ascpCMD in Mac OS X:
  #ascpCMD = "'/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera

  ascpR( ascpCMD, ascpSource, destDir = getwd() )
  dbDisconnect( sra_con )

  ## End(Not run)
} else {
  print( "Use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example" )
}

```

---

ascpSRA

*Fasp SRA data file downloading using the ascp command line program*


---

**Description**

This function downloads SRA data files (fastq, sra ) by fasp protocol using Aspera's ascp command line program, which is included in Aspera Connect software (<http://www.asperasoft.com/>).

**Usage**

```
ascpSRA ( in_acc, sra_con, ascpCMD, fileType = 'sra', destDir = getwd() )
```

**Arguments**

|          |  |
|----------|--|
| in_acc   | character vector of SRA accessions, which should be in same SRA data type, either submission, study, sample, experiment or run.  |
| sra_con  | connection to the SRAMetadb SQLite database.   |
| ascpCMD  | ascp main commands, which should be constructed by a user according to the actual installation of Aspera Connect in the system, with proper options to be used. Example commands: "ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty" (Linux) or "'/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera Connect.app/Contents/Resources/asperaweb_id_dsa.putty'" (Mac OS X). More about ascp please see the help ('ascp -h' in a shell). |
| fileType | type of SRA data files, which should be "sra", or "fastq" ('litesra' has phased out).  |
| destDir  | destination directory to save downloaded files.  |

**Details**

This function will get fasp file sources first using function [listSRAfile](#) and then download data files using function [ascpR](#).

**Value**

A data.frame of all matched SRA accessions and ftp or fasp file addresses.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>

**References**

<http://www.asperasoft.com/>

**See Also**

[ascpR](#), [listSRAfile](#), [getSRAfile](#), [getFASTQinfo](#), [getSRAinfo](#)

**Examples**

```
if( file.exists('SRAMetadb.sqlite') ) {
  ## Not run:
  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'

  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)
  in_acc <- c("SRR000648","SRR000657")
  ascpCMD <- 'ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty'
  ## common ascpCMD for a system with Mac OS X:
  #ascpCMD <- "'/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera

  sraFiles <- ascpSRA( in_acc, sra_con, ascpCMD, fileType = 'sra', destDir=getwd() )
  dbDisconnect(sra_con)

  ## End(Not run)
} else {
  print( "Use ascpSRAdbFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example" )
}
```

---

colDescriptions

*Get column descriptions of SRAMetadb.sqlite*

---

**Description**

Get column descriptions of SRAMetadb.sqlite, including table, field, field data type, description and default values

**Usage**

```
colDescriptions( sra_con )
```

**Arguments**

sra\_con                      Connection of the SRAMetadb SQLite database

**Value**

A seven-column data.frame including table\_name, field\_name, type, description, value\_list.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov> and Sean Davis <sdavis2@mail.nih.gov>

**Examples**

```
if(file.exists('SRAdb.sqlite')) {  
  
  library(SRAdb)  
  sra_dbname <- 'SRAdb.sqlite'  
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)  
  
  ## Get column descriptions  
  a <- colDescriptions(sra_con=sra_con)[1:5,]  
  
} else {  
  print("use getSRAdbFile() to get a copy of the SRAdb.sqlite file and then rerun the example")  
}
```

---

entityGraph

*Create a new graphNEL object from an input entity matrix or data.frame*

---

**Description**

This function will create a new graphNEL object from an input entity matrix or data.frame

**Usage**

```
entityGraph(df)
```

**Arguments**

df                    A matrix or data.frame

**Details**

A graphNEL object with edgemode='directed' is created from input data.frame and the [plot](#) function will draw a graph

**Value**

A graphNEL object with edgemode='directed'

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov> and Sean Davis <sdavis2@mail.nih.gov>



**See Also**

[getSRA](#), [sraConvert](#), [sraGraph](#)

**Examples**

```
if(file.exists('SRAMetadb.sqlite')) {

  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)

  ## create a graphNEL object from SRA accessions, which are full text search results of terms 'primary thyroid
  acc <- getSRA (search_terms = 'primary thyroid cell line', out_types=c('sra'), sra_con=sra_con, acc_only=TR
  g <- entityGraph(acc)
  ## Not run:
  library(Rgraphviz)
  attrs <- getDefaultAttrs(list(node=list(fillcolor='lightblue', shape='ellipse')))
  plot(g, attrs= attrs)

  ## End(Not run)

} else {
  print("use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file
and then rerun the example")
}
```

---

getFASTQfile

*Download SRA fastq files from EBI ENA through ftp or fasp*


---

**Description**

This function downloads SRA fastq data files through ftp or fasp from EBI ENA site for a given list of SRA accessions.

**Usage**

```
getFASTQfile( in_acc, sra_con, destDir = getwd(), srcType = 'ftp', makeDirectory = FALSE, method =
```

**Arguments**

|                            |  |
|----------------------------|--|
| <code>in_acc</code>        | character vector of SRA accessions that could be be in one or more SRA sata types: study, sample, experiment and/or run.   |
| <code>sra_con</code>       | connection to the SRAMetadb SQLite database  |
| <code>destDir</code>       | destination directory to save downloaded fastq files   |
| <code>srcType</code>       | type of transfer protocol, which should be "ftp" or "fasp".  |
| <code>makeDirectory</code> | logical, TRUE or FALSE. If TRUE and baseDir does not exists, storedir will be created to save downloaded files, otherwise downloaded fastq files will be saved to current directory. |
| <code>method</code>        | character vector of length 1, passed to the identically named argument of <a href="#">download.file</a> .  |

ascpCMD            ascp main commands, which should be constructed by a user according to the actual installation of Aspera Connect in the system, with proper options to be used. Example commands: "ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb\_id\_dsa.putty" (Linux) or ""/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera Connect.app/Contents/Resources/asperaweb\_id\_dsa.putty'" (Mac OS X). More about ascp please see the help ('ascp -h' in a shell).

### Details

The function first gets ftp/fasp addresses of SRA fastq files using function [getFASTQinfo](#) for a given list of input SRA accessions; then downloads the fastq files through ftp or fasp.

### Warning

Downloading SRA fastq files through ftp over long distance could take long time and should consider using using 'fasp'.

### Author(s)

Jack Zhu <zhujack@mail.nih.gov>

### See Also

[getFASTQinfo](#), [getSRAfile](#), [ascpR](#)

### Examples

```
if(file.exists('SRAMetadb.sqlite')) {
  ## Not run:
  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect( dbDriver("SQLite"), sra_dbname )

  ## Download fastq files from EBI ENA through ftp
  getFASTQfile( in_acc = c("SRR000648", "SRR000657"), sra_con, destDir = getwd(), srcType = 'ftp', ascpCMD = NULL )

  ## Download fastq files from EBI ENA through fasp
  ascpCMD <- 'ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty'
  ## common ascpCMD for a system with Mac OS X:
  #ascpCMD <- ""/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera
  getFASTQfile( in_acc = c("SRR000648", "SRR000657"), sra_con, srcType='fasp', ascpCMD=ascpCMD )

  dbDisconnect( sra_con )

  ## End(Not run)
} else {
  print("use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example")
}
```

---

|              |   |
|--------------|---|
| getFASTQinfo | <i>Get SRA fastq file information and associated meta data from EBI ENA</i> |
|--------------|---|

---

### Description

This function gets SRA fastq file information and essential associated meta data from EBI ENA web site ( [http://www.ebi.ac.uk/ena/data/view/reports/sra/fastq\\_files/](http://www.ebi.ac.uk/ena/data/view/reports/sra/fastq_files/) ) for SRA accessions given.

### Usage

```
getFASTQinfo( in_acc, sra_con, srcType = 'ftp' )
```

### Arguments

|         |  |
|---------|--|
| in_acc  | character vector of SRA accessions that could be be in one or more SRA sata types: study, sample, experiment and/or run. |
| sra_con | Connection to the SRAMetadb SQLite database  |
| srcType | option for listing either 'ftp' or 'fasp' addresses. The default is 'ftp'.   |

### Details

EBI ENA web site ( [http://www.ebi.ac.uk/ena/data/view/reports/sra/fastq\\_files/](http://www.ebi.ac.uk/ena/data/view/reports/sra/fastq_files/) ) is the souce for parsing infromation from, which is updated and verified daily. Ftp or fasp addresses got from this functon can be used in either [getFASTQfile](#) or [getSRAfile](#) to download the files.

### Value

A data.frame of ftp/fasp inftomation ( addresses, file size, read number, etc) and associated meta data ( study, sample, experiment, run, organism, instrument.platform, instrument.model, library.name, library.layout, library.source, library.selection, run.read.count, run.base.count, etc. ).

### Author(s)

Jack Zhu <[zhujack@mail.nih.gov](mailto:zhujack@mail.nih.gov)>

### See Also

[getFASTQfile](#), [listSRAfile](#), [getSRAfile](#)

### Examples

```
if(file.exists('SRAMetadb.sqlite')) {
  ## Not run:
  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)
  getFASTQinfo( in_acc = c("SRR000648","SRR000657"), sra_con, srcType = 'ftp' )
  getFASTQinfo( in_acc = c("SRR000648","SRR000657"), sra_con, srcType = 'fasp' )

  ## End(Not run)
} else {
  print("Use getSRADBFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example")
}
```

---

`getSRA`*Fulltext search SRA meta data using SQLite fts3 module*

---

### Description

This function does Fulltext search on any SRA fields in any SRA data types with Fulltext capacity in the SQLite and returns SRA records

### Usage

```
getSRA(search_terms, out_types=c('sra', 'submission', 'study', 'experiment', 'sample', 'run'), sra_co
```

### Arguments

|                           |  |
|---------------------------|--|
| <code>search_terms</code> | Free text search terms constructed according to SQLite query syntax defined here: <a href="http://www.sqlite.org/fts3.html#section_1_3">http://www.sqlite.org/fts3.html#section_1_3</a>  |
| <code>out_types</code>    | Character vector of the following SRA data types: 'sra', 'submission', 'study', 'sample', 'experiment', 'run'. Note: if 'sra' is within out_types, the out_types will be set to c('submission', 'study', 'sample', 'experiment') |
| <code>sra_con</code>      | Connection to the SRAMetadb SQLite database  |
| <code>acc_only</code>     | logical, if TRUE, the function will return SRA accession for each out_types  |

### Details

Queries performed by this function could be Phrase queries, e.g. `""lin* app*""`, or NEAR queries, e.g. `""ACID compliant" NEAR/2 sqlite'`, or with the Enhanced Query Syntax. Check Full Text Search section on the SQLite site for details. if `'acc_only=TRUE'`, a data.frame containing only SRA accessions will be returned, which can be used as input for [sraGraph](#).

### Value

A data.frame containing all returned SRA records with fields defined by out\_types.

If `acc_only=FALSE`, a data.frame of matched accessions of out\_types will be returned.

### Author(s)

Jack Zhu <[zhujack@mail.nih.gov](mailto:zhujack@mail.nih.gov)>

### References

<http://www.sqlite.org/>

### See Also

[sraConvert](#)

**Examples**

```

if(file.exists('SRAMetadb.sqlite')) {

  library(SRADb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)

  ## Fulltext search SRA meta data using SQLite fts3 module:
  # find all records with words of 'breast' and 'cancer' in a filed and there could be one to many words between
  rs <- getSRA (search_terms ='breast cancer', out_types=c('run','study'), sra_con=sra_con)

  # find all records with exact phrase of 'breast cancer' in a filed:
  rs <- getSRA (search_terms ="breast cancer", out_types=c('run','study'), sra_con=sra_con)

  # find records with words beginning with 'braes' and 'can', and the distance between them is equal or less than
  rs <- getSRA (search_terms ='breas* NEAR/2 can*', out_types=c('run','study'), sra_con=sra_con)

  # the same as above except that only one space between the two words
  rs <- getSRA (search_terms ="breas* can*", out_types=c('study'), sra_con=sra_con)

  # find records with 'MCF7' or 'MCF-7' - adding double quote to avoid the SQLite to break down 'MCF-7' to 'MCF'
  rs <- getSRA (search_terms ='MCF7 OR "MCF-7"', out_types=c('sample'), sra_con=sra_con)

  # the same as above, but only search the field of 'study_title':
  rs <- getSRA (search_terms ='study_title: brea* can*', out_types=c('run','study'), sra_con=sra_con)

  # the same as above, but only search the field of 'study_title' and return only accessions:
  rs <- getSRA (search_terms ='study_title: brea* can*', out_types=c('run','study'), sra_con=sra_con, acc_only=TRUE)

} else {
  print("use getSRADBFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example")
}

```

---

|              |   |
|--------------|---|
| getSRADBFile | <i>Download and unzip last version of SRAMetadb.sqlite.gz from the server</i> |
|--------------|---|

---

**Description**

This function is the standard method for downloading and unzipping the most recent SRAMetadb SQLite file from the server.

**Usage**

```

getSRADBFile(destdir = getwd(), destfile = "SRAMetadb.sqlite.gz",
             method)

```

**Arguments**

|          |  |
|----------|--|
| destdir  | The destination directory of the downloaded file   |
| destfile | The filename of the downloaded file. This filename should end in ".gz" as the unzipping assumes that is the case |
| method   | Character vector of length 1, passed to the identically named argument of <a href="#">download.file</a> .        |

**Value**

Prints some diagnostic information to the screen.  
Returns the local filename for use later.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>, Sean Davis <sdavis2@mail.nih.gov>

**Examples**

```
## Not run: geometadbfile <- getSRadbFile()
```

---

|                         |   |
|-------------------------|---|
| <code>getSRAfile</code> | <i>Download SRA data file through ftp or fasp</i> |
|-------------------------|---|

---

**Description**

This function downloads sra data files associated with input SRA accessions from NCBI SRA or downloads fastq files from EBI ENA through ftp or fasp protocol.

**Usage**

```
getSRAfile( in_acc, sra_con, destDir = getwd(), fileType = 'sra', srcType = 'ftp', makeDirectory =
```

**Arguments**

|                            |  |
|----------------------------|--|
| <code>in_acc</code>        | character vector of SRA accessions, which should be in same SRA data type, either submission, study, sample, experiment or run.  |
| <code>sra_con</code>       | Connection to the SRAMetadb SQLite database  |
| <code>destDir</code>       | destination directory to save downloaded files.  |
| <code>fileType</code>      | type of SRA data files, which should be "sra", or "fastq" ('litesra' has phased out).  |
| <code>srcType</code>       | type of transfer protocol, which should be "ftp" or "fasp".  |
| <code>makeDirectory</code> | logical, TRUE or FALSE. If TRUE and baseDir does not exists, storedir will be created to save downloaded files, otherwise downloaded fastq files will be saved to current directory.   |
| <code>method</code>        | Character vector of length 1, passed to the identically named argument of <a href="#">download.file</a> .  |
| <code>ascpCMD</code>       | ascp main commands, which should be constructed by a user according to the actual installation of Aspera Connect in the system, with proper options to be used. Example commands: "ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty" (Linux) or ""/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -i '/Applications/Aspera Connect.app/Contents/Resources/asperaweb_id_dsa.putty'" (Mac OS X). More about ascp please see the help ('ascp -h' in a shell). |

**Details**

The function first gets ftp/fasp addresses of SRA data files with function [getSRAinfo](#) for a given list of input SRA accessions; then downloads the SRA data files through ftp or fasp. The sra or sra-lite data files are downloaded from NCBI SRA and the fastq files are downloaded from EBI ENA.

**Warning**

Downloading SRA data files through ftp over long distance could take long time and should consider using using 'fasp'.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>

**See Also**

[listSRAfile](#), [getSRAinfo](#), [getFASTQinfo](#), [getFASTQfile](#)

**Examples**

```
if( file.exists( 'SRAMetadb.sqlite' ) ) {

library(SRAdb)
sra_dbname <- 'SRAMetadb.sqlite'
sra_con <- dbConnect( dbDriver("SQLite"), sra_dbname )

## Not run:
## Download sra files from NCBI SRA using ftp protocol:
getSRAfile( in_acc = c("SRR000648","SRR000657"), sra_con = sra_con, destDir = getwd(), fileType = 'sra' )

## Convert NCBI SRA format (.sra) data to fastq:
## Download SRA Toolkit: http://trace.ncbi.nlm.nih.gov/Traces/sra/sra.cgi?cmd=show&f=software&m=software&s=
## Run fastq-dump to
## system ("fastq-dump SRR000648.sra")

## Download fastq files from EBI using ftp protocol:
getSRAfile( in_acc, sra_con, destDir = getwd(), fileType = 'fastq', srcType = 'ftp', makeDirectory = FALSE, m

## Download fastq files from EBI ftp site using fasp protocol:
ascpCMD <- 'ascp -QT -l 300m -i /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty'
getSRAfile( in_acc, sra_con, fileType = 'fastq', srcType = 'fasp', ascpCMD = ascpCMD )

## End(Not run)

dbDisconnect( sra_con )
} else {
print( "use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example" )
}
```

---

getSRAinfo

*Get SRA data file information from NCBI SRA*

---

**Description**

This function gets SRA .sra file information from NCBI SRA ftp site for a given list SRA accessions.

**Usage**

```
getSRAinfo( in_acc, sra_con, sraType = 'sra' )
```

**Arguments**

|         |   |
|---------|---|
| in_acc  | character vector of SRA accessions, which should be in same SRA data type, either submission, study, sample, experiment or run. |
| sra_con | connection to the SRAMetadb SQLite database   |
| sraType | type of SRA data files, which should be 'sra' ('litesra' has phased out).   |

**Details**

The function first gets ftp addresses of sra or sra-lite data files with function [listSRAfile](#) and then get file size and date from NCBI SRA ftp sites.

**Value**

A data.frame of ftp addresses of SRA data files, and file size and date along with input SRA accessions.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>

**See Also**

[listSRAfile](#), [getSRAfile](#)

**Examples**

```
if(file.exists('SRAMetadb.sqlite')) {
  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)

  ## Get file size and date from NCBI ftp site for available fastq files associated with "SRS012041", "SRS000290"
  # getSRAinfo (in_acc=c("SRS012041", "SRS000290"), sra_con=sra_con, sraType='sra')

} else {
  print(" use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example")
}
```

---

IGVclear

*Clear IGV tracks loaded.*

---

**Description**

Clear IGV tracks loaded in the current IGV.

**Usage**

```
IGVclear(sock)
```

**Arguments**

sock                    A socket connection to IGV.



**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#), [IGVload](#), [IGVgoto](#)

**Examples**

```
## Not run:
## Create a file list from example bam files in the package
exampleBams = file.path(system.file('extdata',package='SRADB'),
  dir(system.file('extdata',package='SRADB'),pattern='bam$'))

##Create a socket connection to IGV
sock <- IGVsocket()
## Load the bam files into IGV
IGVload(sock, exampleBams)

## Clear loaded tracks in the current IGV
IGVclear(sock)

## End(Not run)
```

---

IGVcollapse

*Collapse tracks in the IGV*

---

**Description**

Using the remote command port of IGV, this function collapses tracks in the IGV.

**Usage**

```
IGVcollapse(sock)
```

**Arguments**

sock                    A socket connection to IGV.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#), [IGVload](#)

**Examples**

```
## Not run:  
sock <- IGVsocket()  
IGVcollapse(sock)  
  
## End(Not run)
```

---

IGVgenome

*Set the IGV genome.*

---

**Description**

Set the IGV genome via the remote command port.

**Usage**

```
IGVgenome(sock, genome="hg18")
```

**Arguments**

|        |  |
|--------|--|
| sock   | A socket connection to IGV.                        |
| genome | String representing a genome that IGV knows about. |

**Author(s)**

Sean Davis <sdavis2@mail.nih.gov>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#)

**Examples**

```
## Not run:  
sock <- IGVsocket()  
IGVgenome(sock, genome='hg18')  
  
## End(Not run)
```

---

IGVgoto

*Go to a specified region in IGV.*

---

### Description

Using the remote command port of IGV, go to a specified region.

### Usage

```
IGVgoto(sock, region)
```

### Arguments

|        |   |
|--------|---|
| sock   | A socket connection to IGV.   |
| region | Scrolls to a locus. Use any text that is valid in the IGV search box. |

### Author(s)

Sean Davis <sdavis2@mail.nih.gov>

### References

<http://www.broadinstitute.org/igv/PortCommands>

### See Also

[startIGV](#), [IGVload](#)

### Examples

```
## Not run:
sock <- IGVsocket()
IGVgoto(sock, 'chr1:1-10000')
IGVgoto(sock, 'TP53')

## End(Not run)
```

---

IGVload

*Load data into IGV via remote port call.*

---

### Description

Loads data via a remote call to IGV.

### Usage

```
IGVload(sock, files)
```

**Arguments**

|       |  |
|-------|--|
| sock  | A socket connection to IGV.  |
| files | Character vector of one or more filenames with full path or urls to load. Among supported file types are BAM and IGV session file, for other file types please check IGV web site: <a href="http://www.broadinstitute.org/igv/ControlIGV">http://www.broadinstitute.org/igv/ControlIGV</a> . |

**Author(s)**

Sean Davis <[sdavis2@mail.nih.gov](mailto:sdavis2@mail.nih.gov)>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#), [IGVgoto](#)

**Examples**

```
## Not run:
## Create a file list from example bam files in the package
exampleBams = file.path(system.file('extdata',package='SRadb'),
  dir(system.file('extdata',package='SRadb'),pattern='bam$'))

## Create a socket connection to IGV
sock <- IGVsocket()
## Load the bam files into IGV
IGVload(sock, exampleBams)

## End(Not run)
```

---

IGVsession

*Create an IGV session file*

---

**Description**

This function will create an IGV session file

**Usage**

```
IGVsession(files, sessionFile, genome='hg18', VisibleAttribute='', destdir=getwd())
```

**Arguments**

|                  |   |
|------------------|---|
| files            | Character vector of one or more filenames or urls to load - required.                                   |
| sessionFile      | String representing session file name - required  |
| genome           | String representing a genome that IGV knows about.  |
| VisibleAttribute | Character vector of one or more IGV Visible Attributes to annotate data tracks to be loaded - optional. |
| destdir          | Path where to save the IGV session file.  |

**Details**

While the current state of an IGV session can be saved to a named session file that can be opened to restore the IGV session later on, a IGV session file can be manually or programmatically created to achieve more efficient data loading and better control of IGV. IGVsession function was developed to create such IGV session files. For details please check IGV web site: <http://www.broadinstitute.org/igv/ControlIGV>

**Value**

An IGV session file with full file path.

**Author(s)**

Jack Zhu <zhujack@mail.nih.gov>

**See Also**

[IGVload](#), [IGVgenome](#), [IGVgoto](#)

**Examples**

```
library(SRadb)
exampleBams = file.path(system.file('extdata', package='SRadb'),
  dir(system.file('extdata', package='SRadb'), pattern='bam$'))
exampleSessionFile <- IGVsession(exampleBams, 'exampleBams.xml');
## Not run:
## Start IGV within R. You only need one IGV instance with listen port 60151 open.
startIGV()

## Create a socket connection to IGV
sock <- IGVsocket()
## Wait until IGV fully launched and make sure the listen port for IGV is open (If not configured in IGV, follow
IGVload(sock, exampleSessionFile)

## End(Not run)
```

---

IGVsnapshot

*Make a file snapshot of the current IGV screen.*

---

**Description**

From the IGV documentation: "Saves a snapshot of the IGV window to an image file. If filename is omitted, writes a .png file with a filename generated based on the locus. If filename is specified, the filename extension determines the image file format, which must be .png or .eps."

**Usage**

```
IGVsnapshot(sock, fname = "", dirname=getwd())
```

**Arguments**

|         |  |
|---------|--|
| sock    | A socket connection to IGV.  |
| fname   | The filename to save. Alternatively, if not specified, IGV will create a filename based on the locus being viewed. |
| dirname | The directory name as a string for where to save the snapshot file.  |

**Author(s)**

Sean Davis <sdavis2@mail.nih.gov>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#)

**Examples**

```
## Not run:
## Create a snapshot of the current IGV window, which is usually the first launched IGV with listen port 6015
sock <- IGVsocket()
IGVsnapshot(sock)
dir()

## End(Not run)
```

---

IGVsocket

*Create a Socket Connection to IGV.*

---

**Description**

Create a Socket Connection to IGV by a specified port and host.

**Usage**

```
IGVsocket(host='localhost', port=60151)
```

**Arguments**

|      |   |
|------|---|
| host | The name of remote host where IGV is running. |
| port | The port to connect to/listen on.             |

**Author(s)**

Sean Davis <sdavis2@mail.nih.gov>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#), [IGVgoto](#)

**Examples**

```
## Not run:
## Create a socket connection to IGV
sock <- IGVsocket()

## End(Not run)
```

---

IGVsort

*Sort an alignment track by the specified option.*

---

**Description**

Using the remote command port of IGV, Sorts an alignment track by the specified option. Recognized values for the option parameter are: base, position, strand, quality, sample, and readGroup.

**Usage**

```
IGVsort(sock, option)
```

**Arguments**

|        |   |
|--------|---|
| sock   | A socket connection to IGV.   |
| option | Recognized values for the option parameter are: base, position, strand, quality, sample, and readGroup. |

**Author(s)**

Jack Zhu<zhujack@mail.nih.gov>

**References**

<http://www.broadinstitute.org/igv/PortCommands>

**See Also**

[startIGV](#), [IGVload](#)

**Examples**

```
## Not run:
sock <- IGVsocket()
IGVsort(sock, 'position')
IGVsort(sock, 'base')
IGVsort(sock, 'sample')

## End(Not run)
```

---

|             |   |
|-------------|---|
| listSRAfile | <i>List sra, sra-lite or fastq data file names associated with input SRA accessions</i> |
|-------------|---|

---

### Description

This function lists all sra, sra-lite or fastq data files associated with input SRA accessions

### Usage

```
listSRAfile( in_acc, sra_con, fileType = 'sra', srcType = 'ftp' )
```

### Arguments

|          |   |
|----------|---|
| in_acc   | character vector of SRA accessions, which should be in same SRA data type, either submission, study, sample, experiment or run. |
| sra_con  | connection to the SRAMetadb SQLite database   |
| fileType | types of SRA data files, which should be 'sra' or 'fastq'. ('litesra' has phased out).  |
| srcType  | type of transfer protocol, which should be "ftp" or "fasp".   |

### Details

SRA fastq files are hosted at EBI ftp site (<ftp://ftp.sra.ebi.ac.uk/vol1/fastq/>) and .sra files are hosted at NCBI ftp site (<ftp://ftp-trace.ncbi.nlm.nih.gov/sra/sra-instant/reads/>). 'litesra' has phased out.

### Value

A data frame of matched SRA accessions and data file names with ftp or fasp addresses.

### Author(s)

Jack Zhu <[zhujack@mail.nih.gov](mailto:zhujack@mail.nih.gov)>

### See Also

[getSRAfile](#)

### Examples

```
if( file.exists('SRAMetadb.sqlite') ) {
  ## Not run:
  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)

  ## List ftp or fasp addresses of sra files associated with "SRX000122"
  listSRAfile (in_acc = c("SRX000122"), sra_con = sra_con, fileType = 'sra')
  listSRAfile (in_acc = c("SRX000122"), sra_con = sra_con, fileType = 'sra', srcType='fasp')

  ## End(Not run)
} else {
```



```
print("use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file and then rerun the example")
}
```

---

sraConvert *Cross-reference between GEO data types*

---

## Description

A common task is to find all the SRA entities of one type associated with another SRA entity (eg., find all SRA samples associated with SRA study 'SRP001990'). This function provides a very fast mapping between entity types to facilitate queries of this type.

## Usage

```
sraConvert(in_acc, out_type = c("sra", "submission", "study", "sample", "experiment", "run"), sra_con)
```

## Arguments

|          |  |
|----------|--|
| in_acc   | Character vector of SRA accessions and should be of same SRA data type, either one of SRA submission, SRA study, SRA sample, SRA experiment and SRA run'   |
| out_type | Character vector of the following SRA data types: 'sra', 'submission', 'study', 'sample', 'experiment', 'run'. If 'sra' is in out_type, out_type will be c("submission", "study", "sample", "experiment", "run") |
| sra_con  | Connection to the SRAMetadb SQLite database  |

## Value

A data.frame containing all matched SRA accessions.

## Author(s)

Jack Zhu <zhujack@mail.nih.gov>

## See Also

[getSRA](#), [listSRAfile](#), [getSRAinfo](#)

## Examples

```
if(file.exists('SRAMetadb.sqlite')) {
  library(SRAdb)
  sra_dbname <- 'SRAMetadb.sqlite'
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)

  ## Convert SRA experiment accessions to other types
  a <- sraConvert( in_acc=c(" SRR000137", "SRR000138 "), out_type=c('sample'), sra_con=sra_con )
  b <- sraConvert (in_acc=c("SRX000089"), sra_con=sra_con)

} else {
  print("use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file
and then rerun the example")
}
```

---

|          |   |
|----------|---|
| sraGraph | <i>Create a new graphNEL object of SRA accessions from SRA full text search</i> |
|----------|---|

---

### Description

This function will create a new graphNEL object from SRA accessions using function of [entityGraph](#) and SRA accessions are returned from SRA full text search using function of [getSRA](#)

### Usage

```
sraGraph(search_terms, sra_con)
```

### Arguments

|              |   |
|--------------|---|
| search_terms | Free text search terms constructed according to SQLite query syntax defined here: <a href="http://www.sqlite.org/fts3.html#section_1_3">http://www.sqlite.org/fts3.html#section_1_3</a> |
| sra_con      | Connection to the SRAMetadb SQLite database   |

### Details

This function is a wrapper of two functions: `acc <- getSRA(search_terms, out_types='sra', sra_con, acc_only=TRUE)` and `g <- entityGraph(acc)`. A graphNEL object with `edgemode='directed'` is created from input data.frame of SRA accessions and the [plot](#) function will draw a graph

### Value

A graphNEL object with `edgemode='directed'`

### Author(s)

Jack Zhu <[zhujack@mail.nih.gov](mailto:zhujack@mail.nih.gov)> and Sean Davis <[sdavis2@mail.nih.gov](mailto:sdavis2@mail.nih.gov)>

### See Also

[getSRA](#), [sraConvert](#), [entityGraph](#)

### Examples

```
if(file.exists('SRAMetadb.sqlite')) {  
  
  library(SRAdb)  
  library(Rgraphviz)  
  
  sra_dbname <- 'SRAMetadb.sqlite'  
  sra_con <- dbConnect(dbDriver("SQLite"), sra_dbname)  
  
  ## create a graphNEL object from SRA accessions, which are full text search results of terms 'primary thyroid  
  g <- sraGraph('primary thyroid cell line', sra_con)  
  attrs <- getDefaultAttrs(list(node=list(fillcolor='lightblue', shape='ellipse')))  
  plot(g, attrs=attrs)
```

```

## similiar search as the above, returned much larger data.frame and graph is too clouded
g <- sraGraph('Ewing Sarcoma', sra_con)
## Not run:
plot(g)

## End(Not run)
} else {
print("use getSRAdbFile() to get a copy of the SRAMetadb.sqlite file
and then rerun the example")
}

```

---

startIGV

---

*Start IGV from R with different amount maximum memory support*


---

## Description

This function is to start the Integrative Genomics Viewer (IGV) within R, which is a high-performance visualization tool for interactive exploration of large, integrated datasets. It supports a wide variety of data types including sequence alignments, microarrays, and genomic annotations. In the SRADB, functions of load2IGV and load2newIGV can be used to load BAM format of sequencing data into IGV conveniently.

## Usage

```
startIGV(memory = "mm", devel=FALSE)
```

## Arguments

|        |  |
|--------|--|
| memory | Maximum usable memory support for the IGV to be launched, which is defined as the following: 'mm' - 1.2 GB, 'lm' - 2 GB, 'hm' - 10 GB, '' - 750 MB |
| devel  | Start development version of IGV.  |

## Details

IGV with 1.2 GB maximum usable memory ('mm') is usually for 32-bit Windows; IGV with 2 GB maximum usable memory ('lm') is usually for 32-bit MacOS; IGV with 10 GB maximum usable memory is for large memory 64-bit java machines; IGV with 750 MB (") is sufficient for most applications. The IGV will be launched through Java Web Start. For details about how IGV is launched or have problems to launch it, please refer to this site: <http://www.broadinstitute.org/igv/StartIGV>. Note: if `IGVload` will be used to load BAM files to the new launched IGV, a connection port needs to be enabled in the IGV. This is how to enable connection port in the IGV: in IGV, go View->Preferences->Advanced->Enable port and check the checkbox.

## Author(s)

Jack Zhu

## References

<http://www.broadinstitute.org/igv/>

**See Also**

[IGVload](#), [IGVgoto](#), [IGVgenome](#)

**Examples**

```
## launch IGV with 1.2 GB maximum usable memory support
## Not run: startIGV("lm")
```

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