

# Package ‘Guitar’

April 14, 2017

**Type** Package

**Title** Guitar

**Version** 1.12.0

**Date** 2016-7-14

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**Description** The package is designed for visualization of RNA-related genomic features with respect to the landmarks of RNA transcripts, i.e., transcription starting site, start codon, stop codon and transcription ending site.

**License** GPL-2

**Depends** Rsamtools, GenomicFeatures, rtracklayer, GenomicAlignments, GenomicRanges, ggplot2, grid, IRanges

**biocViews** Sequencing, SplicedAlignment, Alignment, DataImport, RNASeq, MethylSeq, QualityControl, Transcription, Coverage

**NeedsCompilation** no

## R topics documented:

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Guitar-package

*Guitar*

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**Description**

RNA Landmarks Guided Transcriptomic View of RNA-related Genomic Features.

**Details**

The package is designed for transcriptomic visualization of RNA-related genomic features represented with genome-based coordinates with respect to the landmarks of RNA transcripts, i.e., transcription starting site, start codon, stop codon and transcription ending site.

**Author(s)**

Jia Meng <jia.meng@hotmail.com>

**References**

~~ Literature or other references for background information ~~

**Examples**

```
# read genomic features
narrowPeak <- system.file(
  "extdata", "m6A_hg19_1000peaks_mac2.narrowPeak",
  package="Guitar")

# bam imported as GAlignments
m6A_Bcell <- narrowPeaktoGRanges(narrowPeak)

# generate a list of genomic features
m6A_Bcell_1 <- m6A_Bcell[1:300]
m6A_Bcell_2 <- m6A_Bcell[301:600]
m6A_Bcell_3 <- m6A_Bcell[601:900]
feature_hg19 <- list(m6A_Bcell_1, m6A_Bcell_2, m6A_Bcell_3)
names(feature_hg19) <- c("m6A_1", "m6A_2", "m6A_3")

# Make Guitar coordiantes
txdb_file <- system.file("extdata", "hg19_toy.sqlite",
  package="Guitar")
txdb <- loadDb(txdb_file)
gc_txdb <- makeGuitarCoordsFromTxDb(txdb, noBins = 10)

# Plot
GuitarPlot(feature_hg19,
  GuitarCoordsFromTxDb = gc_txdb)
```

---

BED12toGRangesList     *BED12toGRangesList*

---

**Description**

read bed12 format into R as GRangesList object

**Usage**

```
BED12toGRangesList(filepath, header)
```

**Arguments**

filepath	the path where the bed12 file is located
header	whether the bed12 file has header or not, default: FALSE

**Details**

The function read bed12 into R as GRangesList object, with the introns spliced out, making it different from other functions such as import.bed.

**Value**

The returned GRangesList object has the same number of GRangesLists as the number of rows in the bed12 file.

**Author(s)**

Jia Meng <jia.meng@hotmail.com>

**References**

For more information about the bed format, please refer to: <https://genome.ucsc.edu/FAQ/FAQformat.html#format1>

**Examples**

```
bed12=system.file("extdata", "m6A_mm10_exomePeak_1000peaks_bed12.bed", package="Guitar")
m6A_HepG2 <- BED12toGRangesList(bed12)
```

---

combinedGuitarPlot      *combinedGuitarPlot*

---

### Description

combine multiple GuitarPlots, especially when they are from different species and cannot directly handled with GuitarPlot function together.

### Usage

```
combinedGuitarPlot(ct, comLength = c(0.136,0.459, 0.405))
```

### Arguments

ct	Count of overlapping features, which is can be the output from GuitarPlot function. See example.
comLength	a vector of 3 elements,indicating the length of 5'UTR, CDS and 3'UTR in the figure. Default: c(0.136,0.459, 0.405), which is counted from the default setting of hg19 UCSC gene annotation.

### Details

combine multiple GuitarPlots, especially when they are from different species.

### Value

A figure showing the transcriptomic distribution of the genomic features will be generated. Post-editing with Adobe Illustrator or other graphic software is recommended.

### Examples

```
# read genomic features
narrowPeak <- system.file(
  "extdata", "m6A_hg19_1000peaks_mac2.narrowPeak",
  package="Guitar")

# bam imported as GAlignments
m6A_Bcell <- narrowPeaktoGRanges(narrowPeak)

# generate a list of genomic features
m6A_Bcell_1 <- m6A_Bcell[1:300]
m6A_Bcell_2 <- m6A_Bcell[301:600]
m6A_Bcell_3 <- m6A_Bcell[601:900]
m6A_Bcell_4 <- m6A_Bcell[201:900]

# Make Guitar coordiantes
txdb_file <- system.file("extdata", "hg19_toy.sqlite",
  package="Guitar")
txdb <- loadDb(txdb_file)
gc_txdb <- makeGuitarCoordsFromTxDb(txdb,noBins =10)

# Guitar plot 1
```

```
feature1_hg19 <- list(m6A_Bcell_1, m6A_Bcell_2)
names(feature1_hg19) <- c("m6A_1", "m6A_2")
ct1 <- GuitarPlot(feature1_hg19, returnCount = TRUE,
                  GuitarCoordsFromTxDb = gc_txdb)

# Guitar plot 2
feature2_hg19 <- list(m6A_Bcell_3, m6A_Bcell_4)
names(feature2_hg19) <- c("m6A_3", "m6A_4")
ct2 <- GuitarPlot(feature2_hg19, returnCount = TRUE,
                  GuitarCoordsFromTxDb = gc_txdb)

# combine two Gutiar Plot
ct <- rbind(ct1,ct2)
combinedGuitarPlot(ct)
```

---

getNeighborhood	<i>getNeighborhood</i>
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## Description

This function extract the neighborhood DNA regions of a GRangesList object. The neighborhood DNA regions represent the promoter region (5' end) and complementary DNA region on the 3' end, which we call "Tail" here. The 5' end neighborhood DNA can often be useful as a kind of negative control region that is not associated with a clear biological function.

This function is different from the "promoters" function available from GenomicRanges R package, which is designed to extract only the promoter region (5' side DNA), but not the 3' side DNA region.

## Usage

```
getNeighborhood(comp, side = 5, Width = 1000)
```

## Arguments

comp	a GRangesList object, whose 5' or 3' side DNA neighborhood will be extracted.
side	either 5 or 3. When it is 5, the promoter region of the GRangesList object will be extracted; if it is 3, the region on the 3' side complementary to the promoter region will be extracted.
Width	The width of the neighborhood region will be extracted. default: 1000

## Details

All inner list elements of the input GRangesList object must have the same strand

## Value

A GRangesList, of the same length as the input, will be returned. You may use unlist to convert it into a GRanges object.

## Author(s)

Jia Meng <jia.meng@hotmail.com>

**Examples**

```

gr1 <-
  GRanges(seqnames = "chr2", ranges = IRanges(3000, 6000),
          strand = "+", score = 5L, GC = 0.45)
gr2 <-
  GRanges(seqnames = c("chr1", "chr1"),
          ranges = IRanges(c(7000,13000), width = 3),
          strand = c("+"), score = 3:4, GC = c(0.3, 0.5))
grl <- GRangesList("gr1" = gr1, "gr2" = gr2)
promoter <- getNeighborhood(grl,5)
promoter
promoter_GRanges <- unlist(promoter)
promoter_GRanges

```

GuitarPlot

*GuitarPlot***Description**

Plot the transcriptomic distribution of genomic features

**Usage**

```

GuitarPlot(gfeatures, GuitarCoordsFromTxDb = NA,
           txdb = NA, genome = NA,
           noBins = 10, saveToPDFprefix = NA,
           returnCount = FALSE, includeNeighborDNA = FALSE,
           maximalFeatureAmbiguity=5, rescaleComponent=TRUE, fill=FALSE, adjust=1)

```

**Arguments**

<code>gfeatures</code>	genomic features, which can be <code>GRanges</code> , <code>GRangesList</code> , <code>GAlignment</code> .
<code>GuitarCoordsFromTxDb</code>	Guitar coordiantes generated from <code>makeGuitarCoordsFromTxDb</code> object
<code>txdb</code>	If <code>GuitarCoordsFromTxDb</code> is not provided, you may optionally provide a <code>TranscriptDb</code> object, and the Guitar coordiante will be generated accordingly from it.
<code>genome</code>	If neither <code>GuitarCoordsFromTxDb</code> nortxdb is not provided, you may optionally provide genome assembly number, gene annotation will be automatically downloaded from UCSC, and the Guitar coordiante will be generated accordingly from it.
<code>noBins</code>	The resolution of the analysis. Default: 100
<code>saveToPDFprefix</code>	Whether the figure generated should be saved into a PDF file.
<code>returnCount</code>	Whether the count information should be returned. This information is from <code>countOverlaps</code> between the provided <code>gfeatures</code> and Guitar coordiantes. It is not recommended to use this information.
<code>includeNeighborDNA</code>	Whether neighborhood DNA regions should be included in the <code>GuitarPlot</code> .

maximalFeatureAmbiguity	Features that overlap with more than this number of transcripts will not be used in the analysis. If a genomic feature overlaps with multiple transcripts but the number doesn't exceed this threshold, its weight will be evenly divided. Default: 5
rescaleComponent	Adjustment of the bandwidth. Default: 1.
fill	Whether calculate the relative location-standardized density or not. If set "TRUE", the density of tracks will be standardized with respect to each location. Default: FALSE
adjust	Whether calculate the relative location-standardized density or not. If set "TRUE", the density of tracks will be standardized with respect to each location. Default: 1

### Details

This function plots the transcriptomic distribution of genomic features. It is designed for a fast usage of the Guitar package without the needs to go into the details.

### Value

A figure showing the transcriptomic distribution of the genomic features will be generated. Post-editing with Adobe Illustrator or other graphic software is recommended.

### Examples

```
# read genomic features
narrowPeak <- system.file(
  "extdata", "m6A_hg19_1000peaks_mac2.narrowPeak",
  package="Guitar")

# bam imported as GAlignments
m6A_Bcell <- narrowPeaktoGRanges(narrowPeak)

# generate a list of genomic features
m6A_Bcell_1 <- m6A_Bcell[1:300]
m6A_Bcell_2 <- m6A_Bcell[301:600]
m6A_Bcell_3 <- m6A_Bcell[601:900]
feature_hg19 <- list(m6A_Bcell_1, m6A_Bcell_2, m6A_Bcell_3)
names(feature_hg19) <- c("m6A_1", "m6A_2", "m6A_3")

# Make Guitar coordiantes
txdb_file <- system.file("extdata", "hg19_toy.sqlite",
  package="Guitar")
txdb <- loadDb(txdb_file)
gc_txdb <- makeGuitarCoordsFromTxDb(txdb, noBins = 10)

# Plot
GuitarPlot(feature_hg19,
  GuitarCoordsFromTxDb = gc_txdb)
```

---

`makeGuitarCoordsFromGRangesList`*makeGuitarCoordsFromGRangesList*

---

## Description

make Guitar Coordinates From a GRangesList object

## Usage

```
makeGuitarCoordsFromGRangesList(comp, noBins = 100, collapseGene = FALSE, width = 51)
```

## Arguments

<code>comp</code>	A GRangesList object
<code>noBins</code>	The number of sections the "transcript" of the GRangesList object will be equally divided into. This is the resolution of the Guitar coordinates. The larger noBins is, the clearer the visualization will be; however, more computation time and memory resource will be required. You may want to set a smaller number when using a slow computer. Default: 100.
<code>collapseGene</code>	Whether merge the Guitar coordinates from different transcripts. Default: FALSE.
<code>width</code>	The width of each check points. Default: 51

## Value

Guitar coordinates will be generated, which provides a fast reference between genomic coordinates and the transcriptomic coordinates. By default, the returned value should be in GRanges format; if `collapseGene = TRUE`, GRangesList object will be returned.

## Author(s)

Jia Meng <jia.meng@hotmail.com>

## Examples

```
gr1 <- GRanges(seqnames = "chr2", ranges = IRanges(3000, 6000),
  strand = "+", score = 5L, GC = 0.45)
gr2 <- GRanges(seqnames = c("chr1", "chr1"),
  ranges = IRanges(c(7000,13000), width = 3),
  strand = c("+"), score = 3:4, GC = c(0.3, 0.5))
grl <- GRangesList("gr1" = gr1, "gr2" = gr2)
tc <- makeGuitarCoordsFromGRangesList(grl, noBins = 5)
tc
mcols(tc)
```



---

`makeGuitarCoordsFromTxDb`*makeGuitarCoordsFromTxDb*

---

## Description

Make a Guitar Coordinates from TranscriptDb object, i.e., making Guitar coordinates for 8 different components, including, 5'UTR, CDS, 3'UTR, lncRNA, Promoter and Tail of mRNA and lncRNA. Additional filters will discard transcripts that are too short or has too much ambiguous on Genome to increase the sensitivity of the analysis.

## Usage

```
makeGuitarCoordsFromTxDb(txdb, maximalAmbiguity = 3,  
    minimalComponentLength = 100, minimalNcRNALength = 300,  
    noBins = 100)
```

## Arguments

<code>txdb</code>	A transcriptDb object, which can be generated from <code>makeTxDbFromUCSC</code> or other functions.
<code>maximalAmbiguity</code>	If a transcript overlap with more number of transcripts than this number, this transcript will be used in the analysis. By filtering out a number of transcripts, this filter also decrease memory usage and computation time. Default: 3.
<code>minimalComponentLength</code>	The minimal length of the components (5'UTR, CDS, 3'UTR) of a mRNA. Unfortunately, some mRNAs do not all 3 components or some components can be too short and cannot provide effective resolution for the analysis. These mRNAs will be filtered out from the analysis. Default: 100
<code>minimalNcRNALength</code>	non-coding RNAs with length smaller than this value will not be used in the analysis.
<code>noBins</code>	The number of sections the "transcript" of the GRangesList object will be equally divided into. This is the resolution of the Guitar coordinates. The larger noBins is, the clearer the visualization will be; however, more computation time and memory resource will be required. You may want to set a smaller number when using a slow computer. Default: 100.

## Value

A Guitar coordinates (GRanges object) will be returned, with Transcript ID, the relative position of each GRanges on the RNA transcript, the interval (bp) between different coordinates on a transcript component: Front (DNA), Back (DNA), 5'UTR, CDS, 3'UTR, lncRNA.

## Author(s)

Jia Meng <jia.meng@hotmail.com>

**Examples**

```
txdb_file <- system.file("extdata", "hg19_knownGene_sample.sqlite",
                        package="GenomicFeatures")
txdb <- loadDb(txdb_file)
gc_txdb <- makeGuitarCoordsFromTxDb(txdb, noBins = 3)
gc_txdb
```

---

narrowPeaktoGRanges    *narrowPeaktoGRanges*

---

**Description**

read the narrowpeak format from MACS software into GRangesList object.

NAME\_peaks.narrowPeak is BED6+4 format file which contains the peak locations together with peak summit, pvalue and qvalue. You can load it to UCSC genome browser. Definition of some specific columns are: 5th: integer score for display 7th: fold-change 8th: -log10pvalue 9th: -log10qvalue 10th: relative summit position to peak start.

**Usage**

```
narrowPeaktoGRanges(file)
```

**Arguments**

file                    a string specifies where the narrow peak file is located.

**Value**

A GRanges object will be returned.

**References**

MACS2 software: <https://github.com/taoliu/MACS/tree/master/MACS2>

**Examples**

```
narrowPeak=system.file("extdata", "m6A_hg19_1000peaks_mac2.narrowPeak", package="Guitar")
m6A_Bcell <- narrowPeaktoGRanges(narrowPeak) # bam imported as GAlignments
m6A_Bcell
```

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