Package 'mpwR'

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Title Standardized Comparison of Workflows in Mass Spectrometry-Based Bottom-Up Proteomics

Version 0.1.5.1

Description Useful functions to analyze proteomic workflows including number of identifications, data completeness, missed cleavages, quantitative and retention time precision etc. Various software outputs are supported such as 'ProteomeDiscoverer', 'Spectronaut', 'DIA-NN' and 'MaxQuant'.

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Imports comprehenr, data.table, dplyr, flowTraceR, forcats, ggplot2, magrittr, plotly, purrr, stats, stringr, tibble, tidyr, UpSetR

Suggests flextable, knitr, rmarkdown, testthat (>= 3.0.0), utils

VignetteBuilder knitr

Config/testthat/edition 3

Encoding UTF-8

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NeedsCompilation no

Author Oliver Kardell [aut, cre]

Maintainer Oliver Kardell <Okdll@gmx.net>

Repository CRAN

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Contents

`create_example` *Create example data*

Description

Example data for ProteomeDiscoverer, Spectronaut, DIA-NN and MaxQuant.

Usage

```
create_example()
```

Details

Example data is generated for each software for testing functions of mpwR. Each column is created in a randomized fashion. Connections between columns are not necessarily valid. E.g. column of Precursor Charges might not reflect charges of Precuror.IDs column.

Value

This function returns list with example data. Each list entry has filename and software information as well as a corresponding data set.

Author(s)

Oliver Kardell

Examples

```
data <- create_example()
```

get_CV_LFQ_pep *Peptide-level: Quantitative precision*

Description

Calculate quantitative precision on peptide-level

Usage

```
get_CV_LFQ_pep(input_list)
```

Arguments

input_list	A list with data frames and respective quantitative peptide information.

Details

For each submitted data the coefficient of variation is calculated on peptide-level for LFQ intensities. Only full profiles are included.

Value

This function returns the original submitted data of the input_list including a new output column:

- CV_Peptide_LFQ_mpwR - coefficient of variation in percentage.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(stringr)
library(magrittr)
library(tibble)

# Example data
set.seed(123)
data <- list(
  Spectronaut = list(
     filename = "C",
     software = "Spectronaut",
     data = list(
        "Spectronaut" = tibble::tibble(
           Run_mpwR = rep(c("A","B"), times = 5),
           Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 2),
           Stripped.Sequence_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
           Peptide.IDs_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
           ProteinGroup.IDs_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
```

```
            Retention.time_mpwR = sample(1:20, 10),
            Peptide_LFQ_mpwR = sample(1:30, 10),
            ProteinGroup_LFQ_mpwR = sample(1:30, 10))
      )
  )
)

# Result
output <- get_CV_LFQ_pep(
  input_list = data
)
```

`get_CV_LFQ_pg` *Proteingroup-level: Quantitative precision*

Description

Calculate quantitative precision on proteingroup-level

Usage

```
get_CV_LFQ_pg(input_list)
```

Arguments

`input_list`	A list with data frames and respective quantitative proteingroup information.

Details

For each submitted data the coefficient of variation is calculated on proteingroup-level for LFQ intensities. Only full profiles are included.

Value

This function returns the original submitted data of the `input_list` including a new output column:

- CV_ProteinGroup_LFQ_mpwR - coefficient of variation in percentage.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(stringr)
library(magrittr)
library(tibble)

# Example data
set.seed(123)
data <- list(
  Spectronaut = list(
     filename = "C",
     software = "Spectronaut",
     data = list(
        "Spectronaut" = tibble::tibble(
           Run_mpwR = rep(c("A","B"), times = 5),
           Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 2),
           Peptide.IDs_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
           ProteinGroup.IDs_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
           Retention.time_mpwR = sample(1:20, 10),
           Peptide_LFQ_mpwR = sample(1:30, 10),
           ProteinGroup_LFQ_mpwR = sample(1:30, 10))
     )
  )
)

# Result
output <- get_CV_LFQ_pg(
  input_list = data
)
```

---

get_CV_RT *Retention time precision*

Description

Calculate retention time precision

Usage

```
get_CV_RT(input_list)
```

Arguments

input_list	A list with data frames and respective retention time information.

Details

For each submitted data the coefficient of variation is calculated on precursor-level for retention time. Only full profiles are included.

Value

This function returns the original submitted data of the `input_list` including a new output column:

- CV_Retention.time_mpwR - coefficient of variation in percentage.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(tibble)

# Example data
set.seed(123)
data <- list(
  Spectronaut = list(
     filename = "C",
     software = "Spectronaut",
     data = list(
        "Spectronaut" = tibble::tibble(
           Run_mpwR = rep(c("A","B"), times = 5),
           Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 2),
           Peptide.IDs_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
           ProteinGroup.IDs_mpwR = rep(c("A", "B", "C", "D", "E"), each = 2),
           Retention.time_mpwR = sample(1:20, 10),
           Peptide_LFQ_mpwR = sample(1:30, 10),
           ProteinGroup_LFQ_mpwR = sample(1:30, 10))
     )
  )
)

# Result
output <- get_CV_RT(
  input_list = data
)
```

---

get_DC_Report *Data Completeness Report*

Description

Generates a data completeness report from precursor to proteingroup-level

Usage

```
get_DC_Report(input_list, metric = c("absolute", "percentage"))
```

Arguments

input_list	A list with data frames and respective level information.
metric	"absolute" for absolute numbers or "percentage" for displaying percentages. Default is absolute.

Details

For each submitted data a data completeness report is generated highlighting missing values on precursor-, peptide-, protein- and proteingroup-level.

Value

This function returns a list. For each analysis a respective data frame including missing value information per level is stored in the generated list.

- Analysis - analysis name.
- Nr.Missing.Values - number of missing values.
- Precursor.IDs - number of precursor identification per missing value entry - absolute or in percentage.
- Peptide.IDs - number of peptide identification per missing value entry - absolute or in percentage.
- Protein.IDs - number of protein identification per missing value entry - absolute or in percentage.
- ProteinGroup.IDs - number of proteingroup identification per missing value entry - absolute or in percentage.
- Profile - categorical entries: "unique", "sparse", "shared with at least 50%" or "complete".

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(tibble)
library(stringr)

# Example data
data <- list(
DIANN = list(
  filename = "B",
  software = "DIA-NN",
  data = list(
     "DIA-NN" = tibble::tibble(
        Run_mpwR = rep(c("A","B"), times = 10),
        Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4),
        Protein.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4),
        Peptide.IDs_mpwR = rep(c("A", "A", "B", "B", "C"), each = 4),
```

```
        ProteinGroup.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4)
      )
    )
  )
)

# Result
output <- get_DC_Report(
  input_list = data,
  metric = "absolute"
)
```

`get_ID_Report` *Report for identifications*

Description

Generates a report for identifications

Usage

```
get_ID_Report(input_list)
```

Arguments

`input_list`	A list with data frames and respective level information.

Details

For each submitted data a report with achieved number of identifications is generated on precursor-, peptide-, protein- and proteingroup-level.

Value

This function returns a list. For each analysis a respective data frame including number of identifications per run is stored in the generated list.

- Analysis - analysis name.
- Run - run information.
- Precursor.IDs - number of precursor identification.
- Peptide.IDs - number of peptide identification.
- Protein.IDs - number of protein identification.
- ProteinGroup.IDs - number of proteingroup identification.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(tibble)
library(stringr)

# Example data
data <- list(
DIANN = list(
  filename = "B",
  software = "DIA-NN",
  data = list(
     "DIA-NN" = tibble::tibble(
        Run_mpwR = rep(c("A","B"), times = 10),
        Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4),
        Protein.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4),
        Peptide.IDs_mpwR = rep(c("A", "A", "B", "B", "C"), each = 4),
        ProteinGroup.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4)
     )
  )
)
)

# Result
output <- get_ID_Report(
  input_list = data
)
```

`get_MC_Report` *Report about Missed Cleavages*

Description

Generates report with information about number of missed cleavages

Usage

```
get_MC_Report(input_list, metric = c("absolute", "percentage"))
```

Arguments

`input_list`	A list with data frames and respective missed cleavage information.
`metric`	`"absolute"` for absolute numbers or `"percentage"` for displaying percentages. Default is absolute.

Details

For each submitted data a report is generated with information about the number of missed cleavages.

Value

This function returns a list. For each analysis a respective data frame including information of missed cleavages is stored in the generated list.

- Analysis - analysis name.
- Missed.Cleavage - categorical entry with number of missed cleavages.
- mc_count - number of missed cleavages per categorical missed cleavage entry - absolute or in percentage.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(tibble)
library(magrittr)
library(stringr)

# Example data
data <- list(
Spectronaut = list(
  filename = "C",
  software = "Spectronaut",
  data = list(
     "Spectronaut" = tibble::tibble(
        Stripped.Sequence_mpwR = c("A", "B", "C", "D", "E"),
        Missed.Cleavage_mpwR = c(0, 1, 1, 2, 2)
     )
  )
)
)

# Result
output <- get_MC_Report(
  input_list = data,
  metric = "absolute"
)
```

---

get_summary_Report *Summary report*

Description

Generates a summary report

Usage

```
get_summary_Report(
  input_list,
  CV_RT_th_hold = 5,
  CV_LFQ_Pep_th_hold = 20,
  CV_LFQ_PG_th_hold = 20
)
```

Arguments

input_list	A list with data frames including ID, DC, MC, LFQ and RT information.
CV_RT_th_hold	Numeric. User-specified threshold for CV value of retention time precision. Default is 5.
CV_LFQ_Pep_th_hold	Numeric. User-specified threshold for CV value of quantitative precision. Default is 20.
CV_LFQ_PG_th_hold	Numeric. User-specified threshold for CV value of quantitative precision. Default is 20.

Details

For each submitted data a summary report including information about achieved identifications (ID), data completeness (DC), missed cleavages (MC), and both quantitative (LFQ) and retention time (RT) precision is generated.

Value

This function returns a list. For each analysis a respective data frame is stored in the list with the following information:

- Analysis - analysis name.
- "Median ProteinGroup.IDs abs." - median number of proteingroup identifications.
- "Median Protein.IDs abs." - median number of protein identifications.
- "Median Peptide.IDs abs." - median number of peptide identifications.
- "Median Precursor.IDs abs." - median number of precursor identifications.
- "Full profile - Precursor.IDs abs." - number of precursor identifications for full profiles.
- "Full profile - Peptide.IDs abs." - number of peptide identifications for full profiles.
- "Full profile - Protein.IDs abs." - number of protein identifications for full profiles.
- "Full profile - ProteinGroup.IDs abs." - number of proteingroup identifications for full profiles.
- "Full profile - Precursor.IDs %" - number of precursor identifications for full profiles in percentage.
- "Full profile - Peptide.IDs %" - number of peptide identifications for full profiles in percentage.
- "Full profile - Protein.IDs %" - number of protein identifications for full profiles in percentage.

- "Full profile - ProteinGroup.IDs %" - number of proteinGroup identifications for full profiles in percentage.
- "Precursor.IDs abs. with a CV Retention time < X %" - number of precursor identifications with a CV value for retention time precision under user-specified threshold X.
- "Proteingroup.IDs abs. with a CV LFQ < X %" - number of proteingroup identifications with a CV value for quantitative precision under user-specified threshold X.
- "Peptide.IDs abs. with a CV LFQ < X %" - number of peptide identifications with a CV value for quantitative precision under user-specified threshold X.
- "Peptide IDs with zero missed cleavages abs." - number of peptide identifications with zero missed cleavages.
- "Peptide IDs with zero missed cleavages %" - number of peptide identifications with zero missed cleavages in percentage.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(tibble)

# Example data
data <- list(
DIANN = list(
 filename = "B",
 software = "DIA-NN",
 data = list(
   "DIA-NN" = tibble::tibble(
     "Run_mpwR" = c("R01", "R01", "R02", "R03", "R01"),
     "Precursor.IDs_mpwR" = c("A1", "A1", "A1", "A1", "B2"),
     "Retention.time_mpwR" = c(3, 3.5, 4, 5, 4),
     "ProteinGroup_LFQ_mpwR" = c(3, 4, 5, 4, 4),
     "Peptide.IDs_mpwR" = c("A", "A", "A", "A", "B"),
     "Protein.IDs_mpwR" = c("A", "A", "A", "A", "B"),
     "ProteinGroup.IDs_mpwR" = c("A", "A", "A", "A", "B"),
     "Stripped.Sequence_mpwR" = c("ABCR", "AKCR", "ABKCK", "ARKAR", "ABCDR")
   )
 )
)
)

# Result
output <- get_summary_Report(
  input_list = data
)
```

`get_Upset_list` *Generate Upset list*

Description

Generate a list as input for Upset plot

Usage

```
get_Upset_list(
  input_list,
  level = c("Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs"),
  percentage_runs = 100,
  flowTraceR = FALSE,
  remove_traceR_unknownMods = FALSE
)
```

Arguments

`input_list`	A list with data frames and respective level information.
`level`	Character string. Choose between "Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs". Default is "Precursor.IDs".
`percentage_runs`	Number. Percentage of appearance in runs. 100 means: Identification is present in 100% of runs. Default is 100.
`flowTraceR`	Logical. If FALSE no level conversion is applied. Useful for inter-software comparisons. Default is FALSE.
`remove_traceR_unknownMods`	Logical. If FALSE no unknown Modifications are filtered out. Only applies if flowTraceR is set to TRUE. Default is FALSE.

Details

An input is generated for Upset plotting for either precursor-, peptide-, protein- or proteingroup-level. For inter-software comparisons flowTraceR is integrated.

Value

This function returns a list for each analysis with respective level information.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(tibble)
library(magrittr)
library(stringr)

# Example data
data <- list(
DIANN = list(
 filename = "B",
 software = "DIA-NN",
 data = list(
   "DIA-NN" = tibble::tibble(
     Run_mpwR = rep(c("A","B"), times = 10),
     Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4),
     Protein.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4),
     Peptide.IDs_mpwR = rep(c("A", "A", "B", "B", "C"), each = 4),
     ProteinGroup.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 4)
   )
 )
),
Spectronaut = list(
 filename = "C",
 software = "Spectronaut",
 data = list(
   "Spectronaut" = tibble::tibble(
     Run_mpwR = rep(c("A","B"), times = 15),
     Precursor.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 6),
     Peptide.IDs_mpwR = rep(c("A", "A", "B", "B", "C"), each = 6),
     ProteinGroup.IDs_mpwR = rep(c("A2", "A3", "B2", "B3", "C1"), each = 6)
   )
 )
)
)

# Result
output <- get_Upset_list(
  input_list = data,
  level = "Precursor.IDs"
)
```

`load_experimental_design` *Load and Prepare the input data with experimental design file*

Description

Based on submitted experimental design file the input data will be imported, renamed and default filtering will be applied. An experimental design template is available with write_experimental_design.

Usage

```
load_experimental_design(path)
```

Arguments

path	Path to folder with experimental design file. Within exp_design.csv the user needs to specify the analysis name, software and path to analysis folder. Also specific default suffixes are required: for MaxQuant: _evidence, _peptides, _proteinGroups; for PD - R-friendly headers enabled: _PSMs, _Proteins, _PeptideGroups, _ProteinGroups; for DIA-NN, Spectronaut and Generic: _Report

Details

Function for easily importing the default software outputs and preparing for downstream analysis with mpwR from multiple analysis folders. As default for MaxQuant "Reverse", "Potential contaminants" and "Only identified by site" are filtered out. As default for PD only "High" confidence identifications are included and for Found in Sample column(s) also only "High" identifications. Contaminants are filtered out. As default for Spectronaut only EG.Identified equals TRUE are included.

Value

A list - each list entry has filename and software info as well as stored data.

Author(s)

Oliver Kardell

Examples

```
## Not run:
#get template with write_experimental_design and adjust inputs
write_experimental_design("DIRECTORY_TO_FILE")

#load in data
files <- load_experimental_design(path = "DIRECTORY_TO_FILE/your_exp_design.csv")

## End(Not run)
```

---

plot_CV_density *Density plot*

Description

Plot cumulative density for precision results

Usage

```
plot_CV_density(
  input_list,
  xaxes_limit = 50,
  cv_col = c("RT", "Pep_quant", "PG_quant")
)
```

Arguments

input_list	A list with data frames and respective information on quantitative or retention time precision.
xaxes_limit	Numeric. Limit of x-axes in plot.
cv_col	Character string. Choose between "RT", "Pep_quant", "PG_quant" for corresponding precision category. Default is RT for retention time precision. Pep_quant equals quantitative precision on peptide-level. PG_quant equals quantitative precision on proteingroup-level.

Details

Quantitative or retention time precision are plotted as cumulative density.

Value

This function returns a density plot.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(dplyr)
library(comprehenr)
library(tibble)

# Example data
set.seed(123)
data <- list(
  "A" = tibble::tibble(
    Analysis_mpwR = rep("A", times = 10),
    CV_Retention.time_mpwR = sample(1:20, 10),
    CV_Peptide_LFQ_mpwR = sample(1:30, 10),
    CV_ProteinGroup_LFQ_mpwR = sample(1:30, 10)),
 "B" = tibble::tibble(
     Analysis_mpwR = rep("B", times = 10),
     CV_Retention.time_mpwR = sample(1:20, 10),
     CV_Peptide_LFQ_mpwR = sample(1:30, 10),
     CV_ProteinGroup_LFQ_mpwR = sample(1:30, 10))
)
```

```
# Plot
plot_CV_density(
  input_list = data,
  cv_col = "Pep_quant"
)
```

`plot_DC_barplot` *Individual Barplots - Data Completeness*

Description

Plot number of identifications per missing values for each analysis.

Usage

```
plot_DC_barplot(
  input_list,
  level = c("Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs"),
  label = c("absolute", "percentage")
)
```

Arguments

`input_list`	A list with data frames and respective level information.
`level`	Character string. Choose between "Precursor.IDs", "Peptide.IDs", "Protein.IDs" or "ProteinGroup.IDs" for corresponding level. Default is "Precursor.IDs".
`label`	Character string. Choose between "absolute" or "percentage". Default is "absolute".

Details

For each submitted individual analysis a detailed barplot is generated with information about the number of achieved identifications per missing values.

Value

This function returns a list with a barplot for each analysis.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(magrittr)
library(comprehenr)
library(tibble)

# Example data
data <- list(
 "A" = tibble::tibble(
   Analysis = c("A", "A", "A"),
   Nr.Missing.Values = c(2, 1, 0),
   Precursor.IDs = c(50, 200, 4500),
   Peptide.IDs = c(30, 190, 3000),
   Protein.IDs = c(20, 40, 600),
   ProteinGroup.IDs = c(15, 30, 450),
   Profile = c("unique", "shared with at least 50%", "complete")
 ),
 "B" = tibble::tibble(
   Analysis = c("B", "B", "B"),
   Nr.Missing.Values = c(2, 1, 0),
   Precursor.IDs = c(50, 180, 4600),
   Peptide.IDs = c(50, 170, 3200),
   Protein.IDs = c(20, 40, 500),
   ProteinGroup.IDs = c(15, 30, 400),
   Profile = c("unique", "shared with at least 50%", "complete")
 )
)

# Plot
plot_DC_barplot(
  input_list = data,
  level = "Precursor.IDs",
  label = "absolute"
)
```

---

plot_DC_stacked_barplot

Summary Barplot - Data Completeness

Description

Plot number of identifications per missing values as stacked barplot.

Usage

```
plot_DC_stacked_barplot(
  input_list,
  level = c("Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs"),
  label = c("absolute", "percentage")
)
```

Arguments

input_list	A list with data frames and respective level information.
level	Character string. Choose between "Precursor.IDs", "Peptide.IDs", "Protein.IDs" or "ProteinGroup.IDs" for corresponding level. Default is "Precursor.IDs".
label	Character string. Choose between "absolute" or "percentage". Default is "absolute".

Details

The analyses are summarized in a stacked barplot displaying information about the number of achieved identifications per missing values.

Value

This function returns a stacked barplot.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(magrittr)
library(dplyr)
library(tibble)

# Example data
data <- list(
 "A" = tibble::tibble(
   Analysis = c("A", "A", "A"),
   Nr.Missing.Values = c(2, 1, 0),
   Precursor.IDs = c(50, 200, 4500),
   Peptide.IDs = c(30, 190, 3000),
   Protein.IDs = c(20, 40, 600),
   ProteinGroup.IDs = c(15, 30, 450),
   Profile = c("unique", "shared with at least 50%", "complete")
 ),
 "B" = tibble::tibble(
   Analysis = c("B", "B", "B"),
   Nr.Missing.Values = c(2, 1, 0),
   Precursor.IDs = c(50, 180, 4600),
   Peptide.IDs = c(50, 170, 3200),
   Protein.IDs = c(20, 40, 500),
   ProteinGroup.IDs = c(15, 30, 400),
   Profile = c("unique", "shared with at least 50%", "complete")
 )
)

# Plot
plot_DC_stacked_barplot(
```

```
  input_list = data,
  level = "Precursor.IDs",
  label = "absolute"
)
```

`plot_ID_barplot` *Individual Barplots - Identifications*

Description

Plot number of achieved identifications per analysis.

Usage

```
plot_ID_barplot(
  input_list,
  level = c("Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs")
)
```

Arguments

`input_list`	A list with data frames and respective level information.
`level`	Character string. Choose between "Precursor.IDs", "Peptide.IDs", "Protein.IDs" or "ProteinGroup.IDs" for corresponding level. Default is "Precursor.IDs".

Details

For each submitted individual analysis a detailed barplot is generated with information about the number of achieved identifications per run.

Value

This function returns a list with a barplot for each analysis.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(magrittr)
library(comprehenr)
library(tibble)

# Example data
data <- list(
 "A" = tibble::tibble(
   Analysis = c("A", "A", "A"),
```

```
    Run = c("R01", "R02", "R03"),
    Precursor.IDs = c(4800, 4799, 4809),
    Peptide.IDs = c(3194, 3200, 3185),
    Protein.IDs = c(538, 542, 538),
    ProteinGroup.IDs = c(487, 490, 486)
  ),
  "B" = tibble::tibble(
    Analysis = c("B", "B", "B"),
    Run = c("R01", "R02", "R03"),
    Precursor.IDs = c(4597, 4602, 4585),
    Peptide.IDs = c(3194, 3200, 3185),
    Protein.IDs = c(538, 542, 538),
    ProteinGroup.IDs = c(487, 490, 486)
  )
)

# Plot
plot_ID_barplot(
  input_list = data,
  level = "Precursor.IDs"
)
```

`plot_ID_boxplot` *Summary Boxplot - Identifications*

Description

Plot summary of number of identifications in boxplot.

Usage

```
plot_ID_boxplot(
  input_list,
  level = c("Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs")
)
```

Arguments

`input_list`	A list with data frames and respective level information.
`level`	Character string. Choose between "Precursor.IDs", "Peptide.IDs", "Protein.IDs" or "ProteinGroup.IDs" for corresponding level. Default is "Precursor.IDs".

Details

The analyses are summarized in a boxplot displaying information about the number of achieved identifications.

Value

This function returns a boxplot.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(magrittr)
library(dplyr)
library(tibble)

# Example data
data <- list(
 "A" = tibble::tibble(
   Analysis = c("A", "A", "A"),
   Run = c("R01", "R02", "R03"),
   Precursor.IDs = c(7000, 6100, 4809),
   Peptide.IDs = c(3194, 3200, 3185),
   Protein.IDs = c(538, 542, 538),
   ProteinGroup.IDs = c(487, 490, 486)
 ),
 "B" = tibble::tibble(
   Analysis = c("B", "B", "B"),
   Run = c("R01", "R02", "R03"),
   Precursor.IDs = c(3000, 3500, 4585),
   Peptide.IDs = c(3194, 3200, 3185),
   Protein.IDs = c(538, 542, 538),
   ProteinGroup.IDs = c(487, 490, 486)
 )
)

# Plot
plot_ID_boxplot(
  input_list = data,
  level = "Precursor.IDs"
)
```

`plot_MC_barplot` *Individual Barplots - Missed Cleavages*

Description

Plot number of missed cleavages for each analysis.

Usage

```
plot_MC_barplot(input_list, label = c("absolute", "percentage"))
```

Arguments

input_list	A list with data frames and respective information about missed cleavages.
label	Character string. Choose between "absolute" or "percentage". Default is "absolute".

Details

For each submitted individual analysis a detailed barplot is generated with information about the number of missed cleavages.

Value

This function returns a list with a barplot for each analysis.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(comprehenr)
library(tibble)

# Example data
data <- list(
 "A" = tibble::tibble(
   Analysis = c("A", "A", "A", "A", "A"),
   Missed.Cleavage = c("0", "1", "2", "3", "No R/K cleavage site"),
   mc_count = c("2513", "368", "23", "38", "10")
 ),
 "B" = tibble::tibble(
   Analysis = c("B", "B", "B", "B", "B"),
   Missed.Cleavage = c("0", "1", "2", "3", "No R/K cleavage site"),
   mc_count = c("2300", "368", "23", "38", "10")
 )
)

# Plot
plot_MC_barplot(
  input_list = data,
  label = "absolute"
)
```

plot_MC_stacked_barplot *Summary Barplot - Missed Cleavages*

Description

Plot number of missed cleavages as stacked barplot.

Usage

```
plot_MC_stacked_barplot(input_list, label = c("absolute", "percentage"))
```

Arguments

input_list	A list with data frames and respective information about missed cleavages.
label	Character string. Choose between "absolute" or "percentage". Default is "absolute".

Details

The analyses are summarized in a stacked barplot displaying information about the number of missed cleavages.

Value

This function returns a stacked barplot.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(dplyr)
library(tibble)

# Example data
data <- list(
 "A" = tibble::tibble(
   Analysis = c("A", "A", "A", "A", "A"),
   Missed.Cleavage = c("0", "1", "2", "3", "No R/K cleavage site"),
   mc_count = c("2513", "368", "23", "38", "10")
 ),
 "B" = tibble::tibble(
   Analysis = c("B", "B", "B", "B", "B"),
   Missed.Cleavage = c("0", "1", "2", "3", "No R/K cleavage site"),
   mc_count = c("2300", "368", "23", "38", "10")
 )
```

```
)

# Plot
plot_MC_stacked_barplot(
  input_list = data,
  label = "absolute"
)
```

`plot_radarchart` *Radar chart*

Description

Plot radar chart of summary statistics.

Usage

```
plot_radarchart(input_df)
```

Arguments

`input_df`	Data frame with summary information. Analysis column and at least one category column is required.

Details

Summary results are displayed via radar chart. Each analysis has its own trace.

Value

This function returns a radar chart as htmlwidget.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(plotly)
library(tibble)

# Example data
data <- tibble::tibble(
 Analysis = c("A", "B"),
 "Median ProteinGroup.IDs [abs.]" = c(5, 10),
 "Median Protein.IDs [abs.]" = c(5, 10),
 "Median Peptide.IDs [abs.]" = c(5, 10),
 "Median Precursor.IDs [abs.]" = c(5, 10),
 "Full profile - Precursor.IDs [abs.]" = c(5, 10),
```

```
 "Full profile - Peptide.IDs [abs.]" = c(5, 10),
 "Full profile - Protein.IDs [abs.]" = c(5, 10),
 "Full profile - ProteinGroup.IDs [abs.]" = c(5, 10),
 "Full profile - Precursor.IDs [%]" = c(5, 10),
 "Full profile - Peptide.IDs [%]" = c(5, 10),
 "Full profile - Protein.IDs [%]" = c(5, 10),
 "Full profile - ProteinGroup.IDs [%]" = c(5, 10),
 "Precursor.IDs [abs.] with a CV Retention time < 5 [%]" = c(5, 10),
 "Proteingroup.IDs [abs.] with a CV LFQ < 20 [%]" = c(NA, 10),
 "Peptide.IDs [abs.] with a CV LFQ < 20 [%]" = c(NA, 10),
 "Peptide IDs with zero missed cleavages [abs.]" = c(5, 10),
 "Peptide IDs with zero missed cleavages [%]" = c(5, 10)
)

# Plot
plot_radarchart(
  input_df = data
)
```

`plot_Upset` *Upset Plot*

Description

Plot intersections of analyses for different levels.

Usage

```
plot_Upset(
  input_list,
  label = c("Precursor.IDs", "Peptide.IDs", "Protein.IDs", "ProteinGroup.IDs"),
  nr_intersections = 10,
  highlight_overlap = FALSE
)
```

Arguments

`input_list`	A list with data frames and respective level information.
`label`	Character string. Choose between "Precursor.IDs", "Peptide.IDs", "Protein.IDs" or "ProteinGroup.IDs" for corresponding level. Default is "Precursor.IDs".
`nr_intersections`	Numeric. Maximum number of intersections shown in plot. Default is 10.
`highlight_overlap`	Logical. If TRUE, overlapping intersections is highlighted in yellow. Default is FALSE. If TRUE, overlapping intersections need to be in plot!

Details

Identifications per level of each analysis are compared and possible intersections visualized.

Value

This function returns a Upset plot.

Author(s)

Oliver Kardell

Examples

```
# Load libraries
library(UpSetR)
library(tibble)

# Example data
data <- list(
 "A" = c("A", "B", "C", "D"),
 "B" = c("A", "B", "C", "F"),
 "C" = c("A", "B", "G", "E")
)

# Plot
plot_Upset(
  input_list = data,
  label = "Peptide.IDs"
)
```

---

prepare_mpwR *Load and Prepare the input data*

Description

Input data will be imported, renamed and default filtering will be applied

Usage

```
prepare_mpwR(path, diann_addon_pg_qval = 0.01, diann_addon_prec_qval = 0.01)
```

Arguments

path	Path to folder where the input data is stored - only input data. No subfolders or other files. Analysis name as prefix + for MaxQuant: _evidence, _peptides, _proteinGroups; for PD - R-friendly headers enabled: _PSMs, _Proteins, _PeptideGroups, _ProteinGroups; for DIA-NN, Spectronaut and Generic: _Report
diann_addon_pg_qval	Numeric between 0 and 1. Applied only to DIA-NN data: diann_addon_pg_qval <= PG.Q.Value.
diann_addon_prec_qval	Numeric between 0 and 1. Applied only to DIA-NN data: diann_addon_prec_qval <= Q.Value.

Details

Function for easily importing the default software outputs and preparing for downstream analysis with mpwR within one folder. As default for MaxQuant "Reverse", "Potential contaminants" and "Only identified by site" are filtered out. As default for PD only "High" confidence identifications are included and for Found in Sample column(s) also only "High" identifications. Contaminants are filtered out. As default for Spectronaut only EG.Identified equals TRUE are included.

Value

A list - each list entry has filename and software info as well as stored data.

Author(s)

Oliver Kardell

Examples

```
## Not run:
prepare_mpwR(path = "DIRECTORY_TO_FILES")

## End(Not run)
```

---

write_experimental_design

Create template for experimental design

Description

Generation of a exp_design.csv file for using the import option with load_experimental_design.

Usage

```
write_experimental_design(path)
```

Arguments

path	Path to folder where exp_design file is generated.

Details

The generated exp_design.csv file can be used as starting point for importing with the load_experimental_design option for mpwR. Example entries are provided. The template file - exp_design.csv - is generated under the specified path.

Value

This function returns a csv-file with the following columns:

- analysis_name - name of your analysis.
- software - name of used software: DIA-NN, MaxQuant, PD, Spectronaut, Generic.
- path_to_folder - path to analysis folder.

Author(s)

Oliver Kardell

Examples

```
## Not run:
write_experimental_design(path = "DIRECTORY_WHERE_FILE_IS_GENERATED")

## End(Not run)
```

---

write_generic_template

Create generic template

Description

Generation of a template.csv file for generic input data. The template is provided in long-format.

Usage

```
write_generic_template(path_filename)
```

Arguments

path_filename	Path to folder where template is generated and user-defined filename

Details

The generated template.csv file can be used to create a software-independent input file for mpwR. Example entries are provided. The template file - filename_Report.csv - is generated. The appendix "_Report" is required for importing with mpwR. Note that the template is in long-format, so each ProteinGroup.ID has possible multiple entries depending on the number of Precursor.IDs.

Value

This function returns a csv-file with the following columns:

- Run_mpwR - name of file(s).
- ProteinGroup.IDs_mpwR - ProteinGroup with identifier(s) of protein(s) contained in the protein group.
- Protein.IDs_mpwR - Protein identifier(s).
- Peptide.IDs_mpwR - Sequence representation plus possible post-translational modifications.
- Precursor.IDs_mpwR - Sequence representation plus possible post-translational modifications including charge state.
- Stripped.Sequence_mpwR - The amino acid sequence of the identified peptide without modifications.
- Precursor.Charge_mpwR - Charge state of the precursor.
- Missed.Cleavage_mpwR - Number of missed enzymatic cleavages.
- Retention.time_mpwR - Retention time in minutes in the elution profile of the precursor ion.
- ProteinGroup_LFQ_mpwR - LFQ intensity column on proteingroup-level
- Peptide_LFQ_mpwR - LFQ intensity column on petide-level

Author(s)

Oliver Kardell

Examples

```
## Not run:
write_generic_template(path = "DIRECTORY_WHERE_FILE_IS_GENERATED/filename")

## End(Not run)
```

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