# Package 'bnem'

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Type Package

**Title** Training of logical models from indirect measurements of perturbation experiments

**Version** 1.17.0

**Description** bnem combines the use of indirect measurements of Nested Effects Models (package mnem) with the Boolean networks of CellNOptR. Perturbation experiments of signalling nodes in cells are analysed for their effect on the global gene expression profile. Those profiles give evidence for the Boolean regulation of down-stream nodes in the network, e.g., whether two parents activate their child independently (OR-gate) or jointly (AND-gate).

**Depends** R (>= 4.1)

License GPL-3

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**Imports** CellNOptR, matrixStats, snowfall, Rgraphviz, cluster, flexclust, stats, RColorBrewer, epiNEM, mnem, Biobase, methods, utils, graphics, graph, affy, binom, limma, sva, vsn, rmarkdown

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<u> </u>	
absorptionII	
absorption	

## Description

applies absorption law to a disjuncitve normal form

# Usage

```
absorption(bString, model = NULL)
```

# Arguments

bString	a disjunctive normal form or binary vector according to model
model	Model object including the search space, if available. See CellNOntR::preprocessing.

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## Value

bString after absorption law

## Author(s)

Martin Pirkl

## Examples

```
graph <- c("A+B=C", "A=C")
absorption(graph)</pre>
```

 ${\it absorption} II$ 

Inverse absorption

## **Description**

applies "inverse" absorption law to a disjuncitve normal form

## Usage

```
absorptionII(bString, model = NULL)
```

## **Arguments**

bString a disjunctive normal form or binary vector according to model

model Model object including the search space, if available. See CellNOptR::preprocessing.

## Value

bString after "inverse" absorption law

## Author(s)

Martin Pirkl

```
graph <- c("A+B=C", "A=C")
absorptionII(graph)</pre>
```

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addNoise

Add noise

## **Description**

Adds noise to simulated data

## Usage

```
addNoise(sim, sd = 1)
```

## **Arguments**

 $sim \hspace{1cm} bnemsim \hspace{1cm} object \hspace{1cm} from \hspace{1cm} simBoolGtn$ 

sd standard deviation for the rnorm function

#### Value

noisy fold-change matrix

## Author(s)

Martin Pirkl

## **Examples**

```
sim <- simBoolGtn(Sgenes = 10, maxEdges = 10, negation=0.1,layer=1)
fc <- addNoise(sim,sd=1)</pre>
```

bcr

B-Cell receptor signalling perturbations

## **Description**

Processed data from experiments with a stimulated B-Cell receptor (bcr) and perturbed signalling genes. The raw data is available at https://www.ncbi.nlm.nih.gov/geo/ with accession id GSE68761. For the process steps we refer to the publication Martin Pirkl, Elisabeth Hand, Dieter Kube, Rainer Spang, Analyzing synergistic and non-synergistic interactions in signalling pathways using Boolean Nested Effect Models, Bioinformatics, Volume 32, Issue 6, 15 March 2016, Pages 893–900, https://doi.org/10.1093/bioinform Alternatively see also the function processDataBCR for details and for reproduction.

## Usage

bcr

#### References

Martin Pirkl, Elisabeth Hand, Dieter Kube, Rainer Spang, Analyzing synergistic and non-synergistic interactions in signalling pathways using Boolean Nested Effect Models, Bioinformatics, Volume 32, Issue 6, 15 March 2016, Pages 893–900, https://doi.org/10.1093/bioinformatics/btv680

## **Examples**

```
data(bcr)
```

bnem

Boolean Nested Effects Model main function

## **Description**

This function takes a prior network and normalized perturbation data as input and trains logical functions on that prior network

## Usage

```
bnem(
  search = "greedy",
  fc = NULL,
  expression = NULL,
  egenes = NULL,
  pkn = NULL,
  design = NULL,
  stimuli = NULL,
  inhibitors = NULL,
  signals = NULL,
 CNOlist = NULL,
 model = NULL,
  sizeFac = 10^{-10},
 NAFac = 1,
  parameters = list(cutOffs = c(0, 1, 0), scoring = c(0.1, 0.2, 0.9)),
 parallel = NULL,
 method = "cosine",
  relFit = FALSE,
  verbose = TRUE,
  reduce = TRUE,
  parallel2 = 1,
  initBstring = NULL,
  popSize = 100,
  pMutation = 0.5,
 maxTime = Inf,
 maxGens = Inf,
  stallGenMax = 10,
  relTol = 0.01,
```

```
priorBitString = NULL,
  selPress = c(1.2, 1e-04),
  fit = "linear",
  targetBstring = "none",
  elitism = NULL,
  inversion = NULL,
  selection = c("t"),
  type = "SOCK",
  exhaustive = FALSE,
  delcyc = FALSE,
  seeds = 1,
 maxSteps = Inf,
  node = NULL,
  absorpII = TRUE,
  draw = TRUE,
  prior = NULL,
 maxInputsPerGate = 2
)
```

#### Arguments

egenes

search	Type of search heuristic. Either "greedy", "genetic" or "exhaustive". "greedy" uses a greedy algorithm to move through the local neighbourhood of a initial hyper-graph. "genetic" uses a genetic algorithm. "exhaustive" searches through the complete search space and is not recommended.
fc	m x l matrix of foldchanges of gene expression values or equivalent input (nor-

malized pvalues, logodds, ...) for m E-genes and l contrasts. If left NULL, the gene expression data is used to calculate naive foldchanges.

expression Optional normalized m x 1 matrix of gene expression data for m E-genes and 1 experiments.

list object; each list entry is named after an S-gene and contains the names of

egenes which are potential children

pkn Prior knowledge network as output by CellNOptR::readSIF.

design Optional n x l design matrix with n S-genes and l experiments. If available. If

kept NULL, bnem needs either stimuli, inhibitors or a CNOlist object.

stimuli Character vector of stimuli names.

inhibitors Character vector of inhibitors.

signals Optional character vector of signals. Signals are S-genes, which can directly

regulate E-genes. If left NULL, all stimuli and inhibitors are defined as signals.

CNOlist Object (see package CellNOptR), if available.

model Model object including the search space, if available. See CellNOptR::preprocessing.

sizeFac Size factor penelizing the hyper-graph size.

NAFac factor penelizing NAs in the data.

parameters parameters for discrete case (not recommended); has to be a list with entries

cutOffs and scoring: cutOffs = c(a,b,c) with a (cutoff for real zeros), b (cutoff for real effects), c = -1 for normal scoring, c between 0 and 1 for keeping only relevant between -1 and 0 for keeping only a specific quantile of E-genes, and c > 1 for keeping the top c E-genes; scoring = c(a,b,c) with a (weight for real effects), c (weight for real zeros), b (multiplicator for effects/zeros between a

and c);

parallel Parallelize the search. An integer value specifies the number of threads on the

local machine or a list object as in list(c(1,2,3), c("machine1", "machine2", "machine3")) specifies the threads distributed on different machines (local or others).

method Scoring method can be "cosine", a correlation, or a distance measure. See ?cor

and ?dist for details.

relFit if TRUE a relative fit for each E-gene is computed (not recommended)

verbose TRUE for verbose output

reduce if TRUE reduces the search space for exhaustive search parallel2 if TRUE parallelises the starts and not the search itself

initBstring Binary vector for the initial hyper-graph.

popSize Population size (only "genetic").

pMutation Probability between 0 and 1 for mutation (only "genetic").

maxTime Define a maximal time (seconds) for the search.

maxGens Maximal number of generations (only "genetic").

stallGenMax Maximum number of stall generations (only "genetic").

relTol Score tolerance for networks defined as optimal but with a lower score as the

real optimum (only "genetic").

priorBitString Binary vector defining hyper-edges which are added to every hyper-graph. E.g.

if you know hyper-edge 55 is definitly there and to fix that, set priorBitString[55]

<- 1 (only "genetic").

selPress Selection pressure between 1 and 2 (if fit="linear") and greater 2 (for fit "non-

linear") for the stochastic universal sampling (only "genetic").

fit "linear" or "nonlinear fit for stochastic universal sampling

targetBstring define a binary vector representing a network; if this network is found, the com-

putation stops

elitism Number of best hyper-graphs transferred to the next generation (only "genetic").

inversion Number of worst hyper-graphs for which their binary strings are inversed (only

"genetic").

selection "t" for tournament selection and "s" for stochastic universal sampling (only "ge-

netic").

type type of the paralellisation on multpile machines (default: "SOCK")

exhaustive If TRUE an exhaustive search is conducted if the genetic algorithm would take

longer (only "genetic").

delcyc If TRUE deletes cycles in all hyper-graphs (not recommended).

seeds how many starts for the greedy search? (default: 1); uses the n-dimensional

cube (n = number of S-genes) to maximize search space coverage

maxSteps Maximal number of steps (only "greedy").

node vector of S-gene names, which are used in the greedy search; if node = NULL

all nodes are considered

absorpII Use inverse absorption (default: TRUE).

draw If TRUE draws the network evolution.

prior Binary vector. A 1 specifies hyper-edges which should not be optimized (only

"greedy").

maxInputsPerGate

If no model is supplied, one is created with maxInputsPerGate as maximum

number of parents for each hyper-edge.

#### Value

List object including the optimized hyper-graph, its corresponding binary vector for full hyper-graph and optimized scores.

#### Author(s)

Martin Pirkl

#### See Also

nem

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")</pre>
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)</pre>
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1,</pre>
maxInhibit = 2, signals = c("A", "B", "C", "D")
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)</pre>
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,</pre>
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)</pre>
initBstring <- rep(0, length(model$reacID))</pre>
res <- bnem(search = "greedy", model = model, CNOlist = CNOlist,
fc = fc, pkn = PKN, stimuli = "A", inhibitors = c("B", "C", "D"),
parallel = NULL, initBstring = initBstring, draw = FALSE, verbose = FALSE,
maxSteps = Inf)
```

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bnemBs Bootstraped Network	bnemBs	Bootstraped Network	
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## **Description**

Runs Bootstraps on the data

#### Usage

```
bnemBs(fc, x = 10, f = 0.5, replace = TRUE, startString = NULL, ...)
```

## **Arguments**

fc	m x l matrix of foldchanges of gene expression values or equivalent input (normalized pvalues, logodds,) for m E-genes and l contrasts. If left NULL, the gene expression data is used to calculate naive foldchanges.
X	number of bootstraps
f	percentage to sample, e.g. $f = 0.5$ samples only 50 the amount of E-genes as the original data
replace	if TRUE classical bootstrap, if FALSE sub-sampling without replacement
startString	matrix with each row being a string denoting a network to start inference several times with a specific network
• • •	additional parameters for the bnem function

#### Value

list with the accumulation of edges in x and the number of bootstraps in n

#### Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1,
maxInhibit = 2, signals = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)</pre>
```

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```
initBstring <- rep(0, length(model$reacID))
res <- bnemBs(search = "greedy", model = model, CNOlist = CNOlist,
fc = fc, pkn = PKN, stimuli = "A", inhibitors = c("B", "C", "D"),
parallel = NULL, initBstring = initBstring, draw = FALSE, verbose = FALSE,
maxSteps = Inf)</pre>
```

computeFc

Compute differential effects

#### **Description**

computes differential effects given an activation pattern (absolute gene expression or truth table)

## Usage

```
computeFc(CNOlist, y)
```

## **Arguments**

```
CNO1ist CNOlist object (see package CellNOptR), if available.

y activation pattern according to the annotation in CNOlist
```

#### Value

numeric matrix with annotated response scheme

#### Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction", fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1, maxInhibit = 2,
signals = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)</pre>
```

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convertGraph

Convert normal form

## **Description**

converts a disjunctive normal form into a conjunctive normal form and vice versa; input graph as disjunctive normal form like that: c("A+B=D", "C=D", "G+F=U", ...); output is the dual element also in disjunctive normal form;

## Usage

```
convertGraph(g)
```

## **Arguments**

g

graph in normal form

#### Value

converted graph normal form

## Author(s)

Martin Pirkl

## Examples

```
g <- "A+B=C"
g2 <- convertGraph(g)</pre>
```

dummyCNOlist

Create dummy CNOlist

## **Description**

creates a general CNOlist object from meta information

## Usage

```
dummyCNOlist(
   stimuli = NULL,
   inhibitors = NULL,
   maxStim = 0,
   maxInhibit = 0,
   signals = NULL
)
```

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## **Arguments**

stimuli Character vector of stimuli names.

Character vector of inhibitors.

maxStim maximal number of stimulated genes for a single experiment

maxInhibit maximal number of inhibited genes for a single experiment

signals Optional character vector of signals. Signals are S-genes, which can directly regulate E-genes. If left NULL, all stimuli and inhibitors are defined as signals.

#### Value

CNOlist object

#### Author(s)

Martin Pirkl

## **Examples**

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1, maxInhibit = 2,
signals = c("A", "B", "C", "D"))</pre>
```

epiNEM2Bg

Switch between epiNEM and B-NEM

## Description

Convert epiNEM model into general Boolean graph. Only needed for comparing accuracy of inferred network for bnem and epiNEM.

## Usage

```
epiNEM2Bg(t)
```

#### **Arguments**

t full epiNEM model

## Value

differential effects pattern

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#### Author(s)

Martin Pirkl

#### See Also

CreateTopology

#### **Examples**

```
topology <- epiNEM::CreateTopology(3, 1, force = TRUE)
topology <- unlist(unique(topology), recursive = FALSE)
extTopology <- epiNEM::ExtendTopology(topology$model, 100)
b <- epiNEM2Bg(extTopology)</pre>
```

findResiduals

Compute residuals

## **Description**

calculates residuals (data and optimized network do not match) and visualizes them

## Usage

```
findResiduals(
 bString,
 CNOlist,
 model,
  fc = NULL,
  expression = NULL,
  egenes = NULL,
  parameters = list(cutOffs = c(0, 1, 0), scoring = c(0.1, 0.2, 0.9)),
 method = "s",
  sizeFac = 10^{-10},
 main = "residuals for decoupled vertices",
 sub = paste0("green residuals are added effects (left positive,",
    " right negative) and red residuals are deleted ", "effects"),
  cut = TRUE,
  parallel = NULL,
 verbose = TRUE,
)
```

## Arguments

bString Binary vector denoting the network given a model

CNOlist CNOlist object (see package CellNOptR), if available.

model Model object including the search space, if available. See CellNOptR::preprocessing.

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fc	m x l matrix of foldchanges of gene expression values or equivalent input (normalized pvalues, logodds,) for m E-genes and l contrasts. If left NULL, the gene expression data is used to calculate naive foldchanges.
expression	Optional normalized m x l matrix of gene expression data for m E-genes and l experiments.
egenes	list object; each list entry is named after an S-gene and contains the names of egenes which are potential children
parameters	parameters for discrete case (not recommended); has to be a list with entries cutOffs and scoring: cutOffs = $c(a,b,c)$ with a (cutoff for real zeros), b (cutoff for real effects), $c = -1$ for normal scoring, c between 0 and 1 for keeping only relevant between -1 and 0 for keeping only a specific quantile of E-genes, and $c > 1$ for keeping the top c E-genes; scoring = $c(a,b,c)$ with a (weight for real effects), c (weight for real zeros), b (multiplicator for effects/zeros between a and c);
method	Scoring method can be "cosine", a correlation, or a distance measure. See ?cor and ?dist for details.
sizeFac	Size factor penelizing the hyper-graph size.
main	Main title of the figure.
sub	Subtitle of the figure.
cut	If TRUE does not visualize experiments/S-genes which do not have any residuals.
parallel	Parallelize the search. An integer value specifies the number of threads on the local machine or a list object as in $list(c(1,2,3), c("machine1", "machine2", "machine3"))$ specifies the threads distributed on different machines (local or others).
verbose	TRUE for verbose output
	additional parameters for epiNEM::HeatmapOP

#### Value

numeric matrices indicating experiments and/or genes, where the network and the data disagree

## Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1, maxInhibit = 2,
signal = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)</pre>
```

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```
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)
initBstring <- rep(0, length(model$reacID))
res <- bnem(search = "greedy", CNOlist = CNOlist, fc = fc, model = model,
parallel = NULL, initBstring = initBstring, draw = FALSE, verbose = FALSE,
maxSteps = Inf)
rownames(fc) <- seq_len(nrow(fc))
## val <- validateGraph(CNOlist = CNOlist, fc = fc, model = model,
## bString = res$bString, Egenes = 10, Sgene = 4)
residuals <- findResiduals(res$bString, CNOlist, model, fc = fc)</pre>
```

plot.bnem

plot bnem opbject

#### **Description**

plots the boolen network as disjunctive normal form

#### Usage

```
## S3 method for class 'bnem' plot(x, ...)
```

#### **Arguments**

x bnemsim object

... further arguments; see function mnem::plotDnf

## Value

plot of boolean network

#### Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1,
maxInhibit = 2, signals = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)</pre>
```

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```
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)
initBstring <- rep(0, length(model$reacID))
res <- bnem(search = "greedy", model = model, CNOlist = CNOlist,
fc = fc, pkn = PKN, stimuli = "A", inhibitors = c("B", "C", "D"),
parallel = NULL, initBstring = initBstring, draw = FALSE, verbose = FALSE,
maxSteps = Inf, seeds = 10)
plot(res)</pre>
```

plot.bnemBs

Plot Bootstrap result

## **Description**

Shows the result of a Boostrap with either edge frequencies or confidence intervals

## Usage

```
## $3 method for class 'bnemBs'
plot(
    x,
    scale = 3,
    shift = 0.1,
    cut = 0.5,
    dec = 2,
    ci = 0,
    cip = 0.95,
    method = "exact",
    ...
)
```

## Arguments

X	bnemBs object
scale	numeric value for scaling the edgewidth
shift	numeric value for shifting the edgewidth
cut	shows only edges with a fraction larger than cut
dec	integer for function round
ci	if TRUE shows confidence intervals
cip	range for the confidence interval, e.g. 0.95
method	method to use for conidence interval computation (see function binom.confint from package binom)
	additional parameters for the function mnem::plotDnf

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#### Value

plot of the network from the bootstrap

#### Author(s)

Martin Pirkl

## **Examples**

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")</pre>
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)</pre>
CNOlist <- dummyCNOlist("A", c("B","C","D"), maxStim = 1,</pre>
maxInhibit = 2, signals = c("A", "B", "C", "D")
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)</pre>
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,</pre>
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)</pre>
initBstring <- rep(0, length(model$reacID))</pre>
res <- bnemBs(search = "greedy", model = model, CNOlist = CNOlist,</pre>
fc = fc, pkn = PKN, stimuli = "A", inhibitors = c("B","C","D"),
parallel = NULL, initBstring = initBstring, draw = FALSE, verbose = FALSE,
maxSteps = Inf)
```

plot.bnemsim

plot simulation object

#### **Description**

plots the boolen network from a simulation as disjunctive normal form

#### **Usage**

```
## S3 method for class 'bnemsim' plot(x, ...)
```

#### Arguments

x bnemsim object

... further arguments; see function mnem::plotDnf

#### Value

plot of boolean network

18 processDataBCR

#### Author(s)

Martin Pirkl

## **Examples**

```
sim <- simBoolGtn()
plot(sim)</pre>
```

processDataBCR

BCR perturbation reproduction

## **Description**

Produce the application data from the BCR paper of Pirkl, et al., 2016, Bioinformatics. Raw data is available at https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE68761

## Usage

```
processDataBCR(path = "", combsign = FALSE)
```

## Arguments

path to the CEL.gz/Cel files

combsign if TRUE includes all covariates in ComBat analysis to estimate batch effects.

#### Value

list with the full foldchanges and epxression matrix, a reduced foldchange matrix and the design matrix for the computations

#### Author(s)

Martin Pirkl

```
## Not run:
processDataBCR()
## End(Not run)
data(bcr)
```

randomDnf 19

randomDnf

sample normal form

## Description

creates a random normal form or hyper-graph

## Usage

```
randomDnf(
  vertices = 10,
  negation = TRUE,
  max.edge.size = NULL,
  max.edges = NULL,
  dag = FALSE
)
```

## **Arguments**

vertices number of vertices

negation if TRUE, negations (NOT gates) are allowed

max.edge.size maximal number of inputs per edge
max.edges maximal number of hyper-edges
dag if TRUE, graph will be acyclic

## Value

random hyper-graph in normal form

## Author(s)

Martin Pirkl

```
g <- randomDnf(10)</pre>
```

20 reduceGraph

reduceGraph	Reduce graph

#### **Description**

reduces the size of a graph, if possible, to an equivalent sub-graph

## Usage

```
reduceGraph(bString, model, CNOlist)
```

## Arguments

bString binary vector indicating the sub-graph given a model		
model	Model object including the search space, if available. See CellNOptR::preprocessing.	
CNOlist	CNOlist object (see package CellNOptR), if available.	

#### Value

equivalent sub-graph denoted by a bString

#### Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1, maxInhibit = 2,
signal = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)
bString <- reduceGraph(rep(1, length(model$reacID)), model, CNOlist)</pre>
```

scoreDnf 21

scoreDnf

score a boolean network

#### **Description**

computes the score of a boolean network given the model and data

#### Usage

```
scoreDnf(
  bString,
  CNOlist,
  fc,
  expression = NULL,
  model,
  method = "cosine",
  sizeFac = 10^-10,
  NAFac = 1,
  parameters = list(cutOffs = c(0, 1, 0), scoring = c(0.25, 0.5, 2)),
  NEMlist = NULL,
  relFit = FALSE,
  verbose = FALSE
)
```

#### **Arguments**

bString	binary	string	denoting	the	boolean	network

CNOlist Object (see package CellNOptR), if available.

fc m x 1 matrix of foldchanges of gene expression values or equivalent input (nor-

malized pvalues, logodds, ...) for m E-genes and l contrasts. If left NULL, the

gene expression data is used to calculate naive foldchanges.

expression Optional normalized m x 1 matrix of gene expression data for m E-genes and 1

experiments.

model Model object including the search space, if available. See CellNOptR::preprocessing.

method Scoring method can be "cosine", a correlation, or a distance measure. See ?cor

and ?dist for details.

sizeFac Size factor penelizing the hyper-graph size.

NAFac factor penelizing NAs in the data.

parameters parameters for discrete case (not recommended); has to be a list with entries

cutOffs and scoring: cutOffs = c(a,b,c) with a (cutoff for real zeros), b (cutoff for real effects), c = -1 for normal scoring, c between 0 and 1 for keeping only relevant between -1 and 0 for keeping only a specific quantile of E-genes, and c > 1 for keeping the top c E-genes; scoring = c(a,b,c) with a (weight for real effects), c (weight for real zeros), b (multiplicator for effects/zeros between a

and c);

22 simBoolGtn

```
NEMlist NEMlist object (optional)
relFit if TRUE a relative fit for each E-gene is computed (not recommended)
```

verbose TRUE for verbose output

#### Value

```
numeric value (score)
```

## Author(s)

Martin Pirkl

## **Examples**

```
sim <- simBoolGtn()
scoreDnf(sim$bString, sim$CNOlist, sim$fc, model=sim$model)</pre>
```

simBoolGtn

Sample random network and simulate data

## Description

Draws a random prior network, samples a ground truth from the full boolean extension and generates data

## Usage

```
simBoolGtn(
 Sgenes = 10,
 maxEdges = 25,
  stimGenes = 2,
  layer = 1,
  frac = 0.1,
 maxInDeg = 2,
 dag = TRUE,
 maxSize = 2,
 maxStim = 2,
 maxInhibit = 1,
 Egenes = 10,
  flip = 0.33,
  reps = 1,
  keepsif = FALSE,
 negation = 0.25,
 allstim = FALSE,
  and = 0.25,
 positive = TRUE,
  verbose = FALSE
)
```

simBoolGtn 23

#### **Arguments**

Sgenes number of S-genes

maxEdges number of maximum edges (upper limit) in the DAG

stimGenes number of stimulated S-genes

layer scaling factor for the sampling of next Sgene layer of the prior. high (5-10) mean

more depth and low (0-2) means more breadth

frac fraction of hyper-edges in the ground truth (GTN)

maxInDeg maximum number of incoming hyper-edges

dag if TRUE, graph will be acyclic

maxSize maximum number of S-genes in a hyper-edge

maxStim maximum of stimulated S-genes in an experiment (=data samples)

maxInhibit maximum number of inhibited S-genes in an experiment (=data samples)

Egenes number of E-genes per S-gene, e.g. 10 S-genes and 10 E-genes will return 100

E-genes overall

flip fraction of inhibited E-genes

reps number of replicates

keepsif if TRUE does not delete sif file, which encodes the prior network

negation sample probability for negative or NOT edges

allstim full network in which all S-genes are also stimulated

and probability for AND-gates in the GTN

positive if TRUE, sets all stimulation edges to activation, else samples inhibitory edges

by 'negation' probability

verbose TRUE for verbose output

#### Value

list with the corresponding prior graph, ground truth network and data

## Author(s)

Martin Pirkl

```
sim <- simBoolGtn()
plot(sim)</pre>
```

24 simulateStatesRecursive

simulateStatesRecursive

Simulate states

## **Description**

simulates the activation pattern (truth table) of a hyper-graph and annotated perturbation experiments

## Usage

```
simulateStatesRecursive(CNOlist, model, bString, NEMlist = NULL)
```

#### **Arguments**

CNOlist	CNOlist object (see package CellNOptR), if available.			
model	Model object including the search space, if available. See CellNOptR::preprocessing.			
bString	binary vector denoting the sub-graph given model			
NEMlist	NEMlist object only for devel			

#### Value

return the truth tables for certain perturbation experiments as a numeric matrix

#### Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction", fileext=".sif")
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1, maxInhibit = 2,
signal = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)
states <- simulateStatesRecursive(CNOlist, model,
rep(1, length(model$reacID)))</pre>
```

transClose 25

transClose	transitive closure
transClose	transitive closur

#### **Description**

calculates transitive closure of a hyper-graph

## Usage

```
transClose(g, max.iter = NULL, verbose = FALSE)
```

## Arguments

g hyper-graph in normal form

max.iter maximal iteration till convergence

verbose TRUE for verbose output

## Value

transitive closure in normal form

## Author(s)

Martin Pirkl

## **Examples**

```
g <- c("A=B", "B=C")
gclose <- transClose(g)</pre>
```

transRed

transitive reduction

## Description

calculates transitive reduction of a hyper-graph in normal form

## Usage

```
transRed(g, max.iter = NULL, verbose = FALSE)
```

## Arguments

g hyper-graph in normal form

max.iter maximal number of iterations till convergence

verbose TRUE for verbose output

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#### Value

transitive reduction of the hyper-graph in normal form

#### Author(s)

Martin Pirkl

## **Examples**

```
\label{eq:gamma} \begin{array}{lll} g <- c("A=B", "A=C", "B=C", "B=D", "!A=D") \\ gred <- transRed(g) \end{array}
```

validateGraph

validate graph

## **Description**

plotting the observed differential effects of an effect reporter and the expected differential effects of the regulating signalling gene

## Usage

```
validateGraph(
  CNOlist,
  fc = NULL,
  expression = NULL,
 model,
 bString,
  Egenes = 25,
  Sgene = 1,
  parameters = list(cutOffs = c(0, 1, 0), scoring = c(0.1, 0.2, 0.9)),
  plot = TRUE,
  disc = 0,
  affyIds = TRUE,
  relFit = FALSE,
  xrot = 25,
  Rowv = FALSE,
  Colv = FALSE,
  dendrogram = "none",
  soft = TRUE,
  colSideColors = NULL,
  affychip = "hgu133plus2",
 method = "s",
  ranks = FALSE,
  breaks = NULL,
  col = "RdYlGn"
  sizeFac = 10^{-10},
```

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```
order = "rank",
  verbose = TRUE,
    ...
)
```

#### **Arguments**

CNOlist object (see package CellNOptR), if available.

fc m x l matrix of foldchanges of gene expression values or equivalent input (nor-

malized pvalues, logodds, ...) for m E-genes and l contrasts. If left NULL, the

gene expression data is used to calculate naive foldchanges.

expression Optional normalized m x l matrix of gene expression data for m E-genes and l

experiments.

model Model object including the search space, if available. See CellNOptR::preprocessing.

bString Binary string denoting the hyper-graph.

Egenes Maximal number of visualized E-genes.

Sgene Integer denoting the S-gene. See colnames(getSignals(CNOlist)[[1]]) to match

integer with S-gene name.

parameters parameters for discrete case (not recommended); has to be a list with entries

cutOffs and scoring: cutOffs = c(a,b,c) with a (cutoff for real zeros), b (cutoff for real effects), c = -1 for normal scoring, c between 0 and 1 for keeping only relevant between -1 and 0 for keeping only a specific quantile of E-genes, and c > 1 for keeping the top c E-genes; scoring = c(a,b,c) with a (weight for real effects), c (weight for real zeros), b (multiplicator for effects/zeros between a

and c);

plot Plot the heatmap. If FALSE, only corresponding information is printed.

disc Discretize the data.

affyIds Experimental. Turn Affymetrix Ids into HGNC gene symbols.

relFit if TRUE a relative fit for each E-gene is computed (not recommended)

xrot See function epiNEM::HeatmapOP
Rowv See function epiNEM::HeatmapOP
Colv See function epiNEM::HeatmapOP
dendrogram See function epiNEM::HeatmapOP

soft if TRUE, assigns weights to the expected pattern

colSideColors See function epiNEM::HeatmapOP

affychip Define Affymetrix chip used to generate the data (optional and experimental).

method Scoring method can be "cosine", a correlation, or a distance measure. See ?cor

and ?dist for details.

ranks if TRUE, turns data into ranks
breaks See function epiNEM::HeatmapOP
col See function epiNEM::HeatmapOP

sizeFac Size factor penelizing the hyper-graph size.

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```
order Order by "rank", "name" or "none"
verbose TRUE for verbose output
... additional arguments for epiNEM::HeatmapOP
```

#### Value

lattice object with matrix information

#### Author(s)

Martin Pirkl

```
sifMatrix <- rbind(c("A", 1, "B"), c("A", 1, "C"), c("B", 1, "D"),
c("C", 1, "D"))
temp.file <- tempfile(pattern="interaction",fileext=".sif")</pre>
write.table(sifMatrix, file = temp.file, sep = "\t",
row.names = FALSE, col.names = FALSE,
quote = FALSE)
PKN <- CellNOptR::readSIF(temp.file)</pre>
CNOlist <- dummyCNOlist("A", c("B", "C", "D"), maxStim = 1, maxInhibit = 2,</pre>
signal = c("A", "B", "C", "D"))
model <- CellNOptR::preprocessing(CNOlist, PKN, maxInputsPerGate = 100)</pre>
expression <- matrix(rnorm(nrow(slot(CNOlist, "cues"))*10), 10,</pre>
nrow(slot(CNOlist, "cues")))
fc <- computeFc(CNOlist, expression)</pre>
initBstring <- rep(0, length(model$reacID))</pre>
res <- bnem(search = "greedy", CNOlist = CNOlist, fc = fc,
model = model, parallel = NULL, initBstring = initBstring, draw = FALSE,
verbose = FALSE, maxSteps = Inf)
rownames(fc) <- seq_len(nrow(fc))</pre>
val <- validateGraph(CNOlist = CNOlist, fc = fc, model = model,</pre>
bString = res$bString, Egenes = 10, Sgene = 4)
```

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